

**Asymmetric Synthesis of Bioactive Molecules *via*
Hydrolytic Kinetic Resolution of Alkoxy Epoxides,
Dihydroxylation of Alkenes and Synthetic
Methodologies Involving Oxidative Cyclization
of Styrenes and Esterification of Aldehydes**

A THESIS

**SUBMITTED TO THE
UNIVERSITY OF PUNE**

**FOR THE DEGREE OF
DOCTOR OF PHILOSOPHY
IN
CHEMISTRY**

**By
Chaithanya Kiran, I. N.**

**UNDER THE GUIDANCE OF
Dr. A. Sudalai**

**Chemical Engineering and Process Development Division
CSIR-National Chemical Laboratory
Pune-411008, INDIA
March 2014**

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March 2014



DEDICATED TO

MY BELOVED BROTHER

I. LAKSHMI NARAYANA



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CERTIFICATE

This is to certify that the work incorporated in the thesis entitled “*Asymmetric Synthesis of Bioactive Molecules via Hydrolytic Kinetic Resolution of Alkoxy Epoxides, Dihydroxylation of Alkenes and Synthetic Methodologies Involving Oxidative Cyclization of Styrenes and Esterification of Aldehydes*” which is being submitted to the *University of Pune* for the award of *Doctor of Philosophy* in *Chemistry* by *Mr. Chaithanya Kiran, I. N.* was carried out by him under my supervision at the CSIR-National Chemical Laboratory, Pune. Such material as has been obtained from other sources has been duly acknowledged in the thesis.


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DECLARATION

I hereby declare that the thesis entitled *“Asymmetric Synthesis of Bioactive Molecules via Hydrolytic Kinetic Resolution of Alkoxy Epoxides, Dihydroxylation of Alkenes and Synthetic Methodologies Involving Oxidative Cyclization of Styrenes and Esterification of Aldehydes”* submitted for the degree of Doctor of Philosophy in Chemistry to the University of Pune, has not been submitted by me to any other university or institution. This work was carried out at the CSIR-National Chemical Laboratory, Pune, India.

March 2014

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I always love my brother Lakshmi Narayana and his family for their constant encouragement. I thank him for giving me wings to fly and allowing the spark in me to glow, yes this thesis is dedicated to you.

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Chaithanya Kiran, I. N.

ABBREVIATIONS

Ac	Acetyl
Ar	Aryl
Bn	Benzyl
Boc	<i>N-tert</i> -Butoxycarbonyl
(Boc) ₂ O	Di <i>tert</i> -butyl dicarbonate
<i>n</i> -Bu	<i>n</i> -Butyl
<i>n</i> -BuLi	<i>n</i> -Butyl lithium
<i>t</i> -Bu	<i>tert</i> -Butyl
Cbz	Benzyloxy carbonyl
CH ₂ Cl ₂	Methylene chloride
CHCl ₃	Chloroform
CH ₃ CN	Acetonitrile
CuSO ₄	Copper(II) sulfate
DBU	1,8-Diazabicyclo[5.4.0]undec-7-ene
DIBAL-H	Diisobutyl aluminium hydride
DMF	Dimethyl formamide
DMSO	Dimethyl sulphoxide
DMAP	<i>N,N</i> -dimethyl-4-aminopyridine
dr	Diastereomeric ratio
ee	Enantiomeric excess
Et	Ethyl
Et ₃ N	Triethylamine
Et ₂ O	Diethyl ether
EtOAc	Ethyl acetate
EtOH	Ethyl alcohol
g	Grams
h	Hours
HCl	Hydrochloric acid
HPLC	High pressure liquid chromatography
H ₂ SO ₄	Sulfuric acid
HNO ₃	Nitric acid
imid.	Imidazole
IR	Infra red
IBX	2-Iodoxybenzoic acid
K ₂ CO ₃	Potassium carbonate
KOH	Potassium hydroxide
LiAlH ₄	Lithium aluminum hydride
LiHMDS	Lithium hexamethyldisilazide
M+	Molecular ion

Me	Methyl
MeOH	Methyl alcohol
MOM	Methoxymethyl
min	Minutes
mg	Miligram
mL	Milliliter
mp	Melting point
MS	Mass spectrum
Ms	Mesyl
NaBH ₄	Sodium borohydride
NaHCO ₃	Sodium bicarbonate
NaOH	Sodium hydroxide
Na ₂ SO ₄	Sodium sulfate
NH ₄ Cl	Ammonium chloride
NH ₄ OH	Ammonium hydroxide
NBS	<i>N</i> -Bromosuccinimide
NMR	Nuclear Magnetic Resonance
NMO	<i>N</i> -Methyl morpholine <i>N</i> -oxide
PCC	Pyridinium chlorochromate
Pd/C	Palladium on activated charcoal
PDC	Pyridinium dichromate
Pd(OH) ₂	Palladium hydroxide
Ph	Phenyl
<i>p</i> -Ts	<i>p</i> -Tosyl
<i>p</i> -TSA	<i>p</i> -Toluene sulfonic acid
PhNO	Nitrosobenzene
Py	Pyridine
TBS	<i>tert</i> -Butyldimethylsilyl
TEMPO	(2,2,6,6-tetramethyl-1-piperidinyl)oxyl
THF	Tetrahydrofuran
TLC	Thin layer chromatography
TBAF	Tetrabutylammonium fluoride
TBDMSCI	<i>tert</i> -Butyldimethylsilyl chloride
TBDPSCI	<i>tert</i> -Butyldiphenylsilyl chloride
TFA	Trifluoroacetic acid

GENERAL REMARKS

1. All solvents were distilled and dried before use.
2. Petroleum ether refers to the fraction collected in the boiling range 60-80 °C.
3. Organic layers after every extraction were dried over anhydrous sodium sulfate.
4. Column Chromatography was performed over silica gel (230-400 mesh).
5. TLC analyses were performed over aluminum plates coated with silica gel (5-25 m) containing UV active G-254 additive.
6. IR spectra were recorded on a Perkin-Elmer model 683 B or 1605 FT-IR and absorptions were expressed in cm^{-1} .
7. ^1H and ^{13}C NMR spectra were recorded on Bruker FT AC-200 MHz, Bruker Avance 500 MHz and JEOL ECX 400 instruments using TMS as an internal standard. The following abbreviations were used: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br s = broad singlet, dd = doublet of doublet, dt = doublet of triplet and ddd = doublet of doublet of doublet.
8. Optical rotations were carried out on JASCO-181 digital polarimeter at 25 °C using sodium D light.
9. Enantiomeric excesses were determined on Agilent HPLC instrument equipped with a chiral column.
10. HRMS data were recorded on a Thermo Scientific Q-Exactive, Accela 1250 pump.
11. All melting points and boiling points are uncorrected and the temperatures are in centigrade scale.
12. Elemental analysis was done on Carlo ERBA EA 110B instrument.
13. The compounds, scheme and reference numbers given in each chapter refers to that particular chapter only.

ABSTRACT

The thesis entitled “**Asymmetric Synthesis of Bioactive Molecules via Hydrolytic Kinetic Resolution of Alkoxy Epoxides, Dihydroxylation of Alkenes and Synthetic Methodologies Involving Oxidative Cyclization of Styrenes and Esterification of Aldehydes**” is divided into four chapters.

The title of the thesis clearly reflects the objective, which is to synthesize enantiomerically pure bioactive molecules, drugs and to interface synthetic organic chemistry for the development of new methodologies. **Chapter 1** describes a short enantioselective synthesis of (+)-goniodiol and (-)-polysphorin analog using hydrolytic kinetic resolution of alkoxy epoxides. **Chapter 2** presents the synthesis of 3-substituted chiral phthalides using CN-assisted oxidative cyclization of cyano cinnamates and styrene derivatives and its application in the synthesis of (-)-matteucen C and butylphthalide. **Chapter 3** deals with asymmetric synthesis of 3-amino-1,2-diols *via* proline catalyzed α -aminooxylation and its application in the synthesis of (+)-*epi*-cytoxazone and formal synthesis of *N*-thiolated-2-oxazolidinone. **Chapter 4** presents an organocatalyzed asymmetric synthesis of rasagiline and NHC catalyzed esterification of aromatic aldehydes.

CHAPTER 1

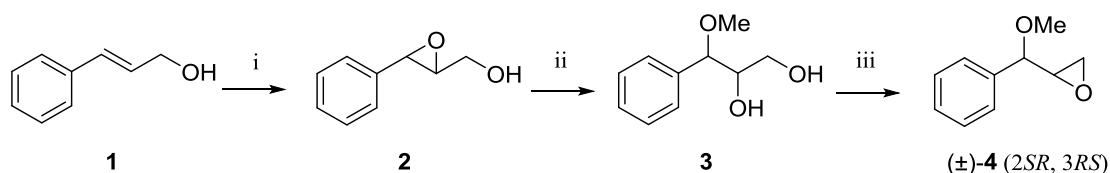
Short Enantioselective Synthesis of (+)-Goniodiol and (-)-Polysphorin Analog using Co-Catalyzed HKR of Alkoxy Epoxides

Jacobsen’s Hydrolytic Kinetic Resolution (HKR) has emerged as an effective method for obtaining chiral epoxides and 1,2-diols in a highly enantioenriched forms.¹ These compounds are important intermediates in the synthesis of various bioactive molecules.² In view of easy availability of chiral ligands and the simplicity of the reaction conditions with water being used as the nucleophile, HKR is being used extensively for providing several chiral building blocks in the synthesis of biologically active compounds.³ This chapter is divided into two sections. **Section I** deals with the enantioselective synthesis of (+)-goniodiol (**13**) and **Section II** describes enantioselective synthesis of (-)-polysphorin analog (**27**) using two-stereocentered HKR of alkoxy epoxides.

Section I: Concise Enantioselective Synthesis of (+)-Goniodiol, a Cytotoxic Lactone via Co-Catalyzed Hydrolytic Kinetic Resolution of *anti*-(2*SR*, 3*RS*)-3-Methoxy-3-phenyl-1,2-epoxypropane

(+)-Goniodiol (**13**), a styryl lactone, was isolated from petroleum ether extracts of leaves and twigs of *Goniothalamus sesquipedalis*.⁴ It exhibits potent and selective cytotoxic activity against human lung carcinoma and leukemia cells.⁵

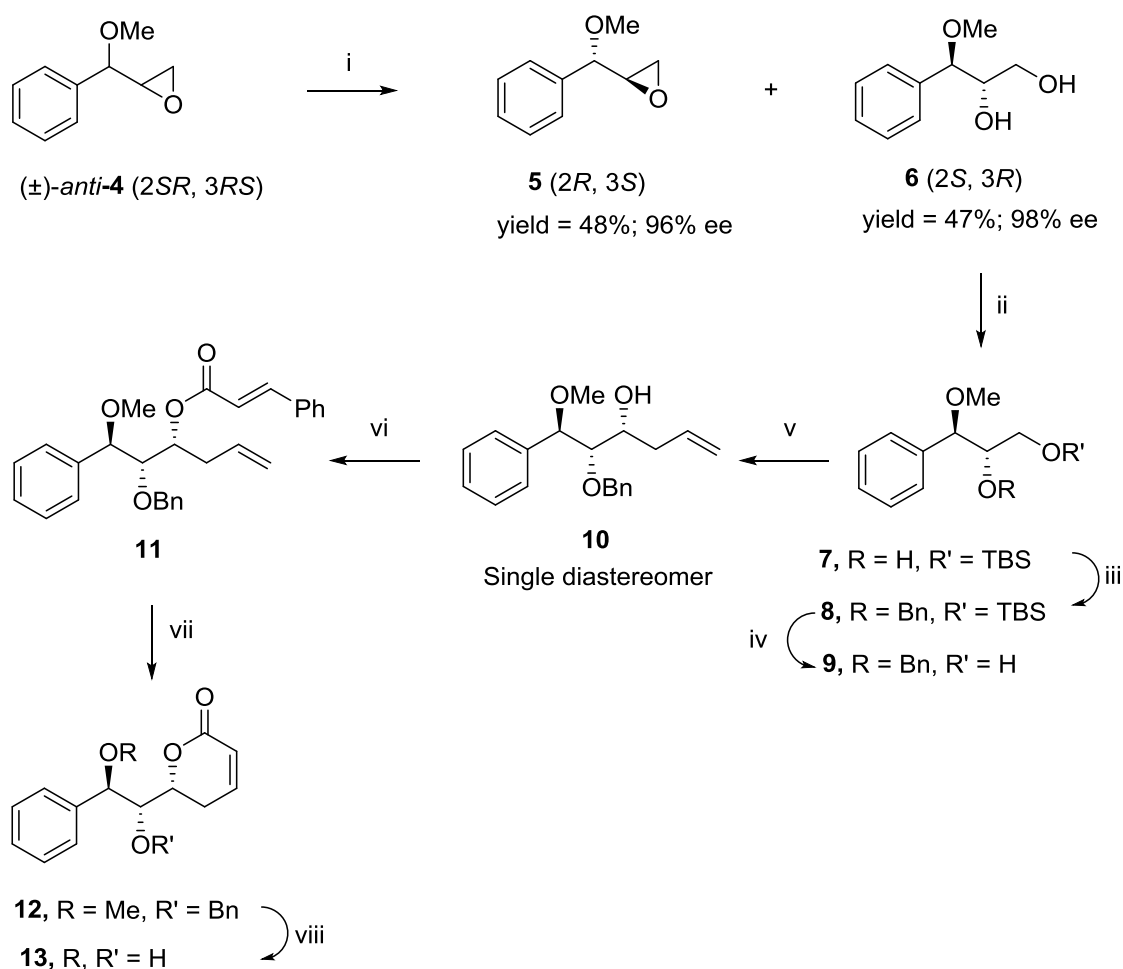
The asymmetric synthesis of (+)-goniodiol begins with commercially available cinnamyl alcohol and its transformation into racemic *anti*-methoxy epoxide **4** in 4 steps. Cinnamyl alcohol **1** on epoxidation with *m*CPBA gave the epoxy alcohol **2**. The regiospecific ring opening of epoxide **2** with MeOH was achieved quantitatively to give diol **3**, which was further converted to *anti*-methoxy epoxide **4** in a two-step reaction sequence: the selective monotosylation of primary alcohol (TsCl, Bu₂SnO, Et₃N, DMAP, CH₂Cl₂) followed by epoxide formation under basic conditions (K₂CO₃, MeOH) (**Scheme 1**).



Scheme 1: (i) *m*CPBA, CH₂Cl₂, 14 h, 0 °C, 88%; (ii) camphor sulfonic acid (10 mol %), MeOH, 1 h, 96%; (iii) (a) TsCl, Bu₂SnO (2 mol%), Et₃N, DMAP (10 mol%), CH₂Cl₂, 2 h; (b) K₂CO₃, MeOH, 3 h, 76% (over two steps).

Racemic *anti*-methoxy oxirane **4** was subjected to HKR by using (*S*, *S*)-salen-Co-OAc catalyst to give highly enantiomerically pure methoxy epoxide **5** (48% yield, 96% ee), and methoxy diol **6** (47% yield, 98% ee).⁷ The primary hydroxyl group in **6** was selectively protected as its TBS ether **7**, followed by its secondary hydroxyl protection as its benzyl ether **8** (NaH, BnBr in DMF). Further TBS group was selectively deprotected to give the primary alcohol **9** in 91% yield, which was oxidized using IBX in DMSO to give the corresponding aldehyde. To this crude aldehyde in CH₂Cl₂ at -78 °C was added TiCl₄ and tributyl allyl stannane which produced homoallylic alcohol **10**, as a single diastereomer (¹H NMR analysis) in 73% yield.⁶ Homo allylic alcohol **10** was esterified with (*E*)-cinnamoyl chloride to give cinnamate ester **11**, which on RCM Grubbs' second generation catalyst gave α -pyrone **12** in 76% yield. Both methyl and benzyl ethers in pyrone **12** were cleaved in a single

step using BBr_3 to afford (+)-goniodiol **13** in 12.25% overall yield and 98% ee (Scheme 2).⁸

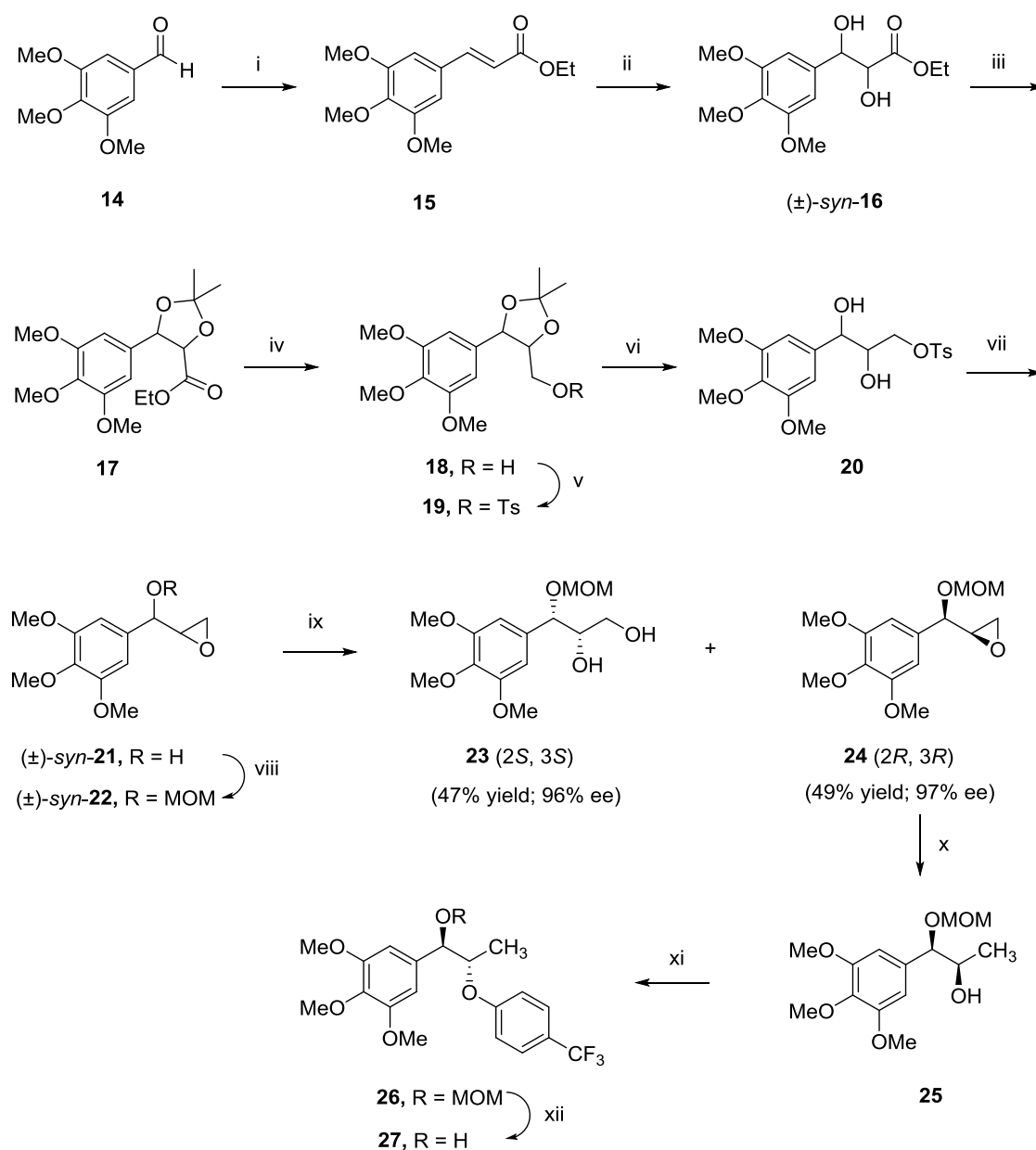


Scheme 2: (i) (*S,S*)-Co(salen)OAc, (0.5 mol%), THF, H_2O (0.5 equiv), 0 °C, 14 h; (ii) TBDMSCl, imid., CH_2Cl_2 , 0 °C to 25 °C, 1 h, 90%; (iii) NaH, DMF, BnBr, 0 °C to 25 °C, 2 h, 95%; (iv) TBAF, THF, 25 °C, 8 h, 91%; (v) IBX, DMSO, 2 h, 25 °C then TiCl_4 , tributyl allyl stannane, CH_2Cl_2 , -78 °C, 30 min, 73% (over two steps); (vi) (*E*)-cinnamoyl chloride, dry pyridine, DMAP, CH_2Cl_2 , 0 °C to 25 °C, 12 h, 64%; (vii) Grubbs' II generation catalyst (10 mol%), CH_2Cl_2 reflux, 8 h, 76%; (viii) BBr_3 , CH_2Cl_2 , -78 °C, 6 h, 71%.

SECTION II: Enantioselective Synthesis of (-)-Polysporin Analog via Co-Catalyzed Hydrolytic Kinetic Resolution of *syn*-(2*SR*, 3*SR*)-3-(Methoxymethyl)oxy-3-(3,4,5-trimethoxyphenyl)-1,2-epoxypropane

Polysporin neolignan isolated from *Piper polysporum* C in China,⁹ and from the leaves and stems of *Rhaphidopora decursiva* in Vietnam¹⁰ was shown to possess *in vitro* antimalarial activity (IC_{50} 400ng/ml).¹¹ Several other members of this family of

neolignans have also been shown to display interesting biological properties. Polysphorin analog (**27**) was found to be highly effective against malaria during the hepatic phase of the disease.¹²



Scheme 3: (i) $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$, CH_2Cl_2 , 25 °C, 5 h, 96%; (ii) $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$ (2 mol%), NMO, acetone: H_2O (4:1), 0 to 25 °C, 20 h, 93%; (iii) 2,2-dimethoxypropane, camphor sulfonic acid (10 mol %), CH_2Cl_2 , 25 °C, 6 h, 98%; (iv) LiAlH_4 , THF, 0 to 25 °C, 1 h, 92%; (v) *p*-TsCl, Et_3N , DMAP (10 mol%), CH_2Cl_2 , 25 °C, 4 h, 94%; (vi) camphor sulfonic acid (10 mol %), CH_3OH , 40 °C, 1 h, 96%; (vii) K_2CO_3 , CH_3OH , 25 °C, 3 h, 88%; (viii) MOMCl, $i\text{Pr}_2\text{NEt}$, CH_2Cl_2 , 25 °C, 8 h, 92%; (ix) (*S,S*)-Co(salen)OAc (0.5 mol%), H_2O (0.5 equiv), 0 °C, 14 h; (x) LiAlH_4 , THF, 0 to 25 °C, 4 h, 96%; (xi) PPh_3 , $i\text{PrO}_2\text{C}-\text{N}=\text{N}-\text{CO}_2\text{Pr}^i$, 4-(trifluoromethyl)phenol, THF, 70 °C, 12 h, 74%; (xii) conc. HCl, CH_3OH , 25 °C, 4 h, 86%.

The complete synthetic sequence for polysphorin analog (**27**), commencing from the precursor aldehyde **14**, is shown in **Scheme 3**. Wittig olefination of aldehyde **14** resulted in cinnamate ester **15** (96% yield), which was subjected to Os-catalyzed Upjohn dihydroxylation to give the *syn* dihydroxy ester **16** in 93% yield. Acetonide protection of diol in **16** was achieved in 98% yield (2,2 - dimethoxypropane, CSA in CH₂Cl₂). Reduction of ester **17** using LiAlH₄ furnished alcohol **18** in quantitative yield, which was further converted to epoxy alcohol **21** in a three-step reaction sequence: (i) tosylation of primary alcohol (TsCl, Et₃N, DMAP); (ii) acetonide deprotection (CSA, MeOH, 40 °C) followed by (iii) epoxide formation under basic conditions (K₂CO₃, MeOH). Secondary alcohol in epoxy alcohol **21** was protected as its MOM ether to give *syn*-racemic epoxide **22**, precursor for HKR in 92% yield. Racemic oxirane **22** was subjected to two-setereocentred HKR by using (*S*, *S*)-salen-Co-OAc catalyst to give highly enantiomerically pure diol **23** (47% yield, 96% ee) and epoxide **24** (49% yield, 97% ee).⁷ Regioselective reductive ring opening of the epoxide **24** was achieved with LiAlH₄ to afford secondary alcohol **25** in 96% yield. The Mitsunobu reaction of alcohol **25** with *p*-trifluoromethyl phenol gave the protected neolignan **26** in 74% yield. Subsequent deprotection of MOM group in **26** using conc.HCl gave the (-)-polysphorin analog (**27**) in 86% yield.

CHAPTER 2

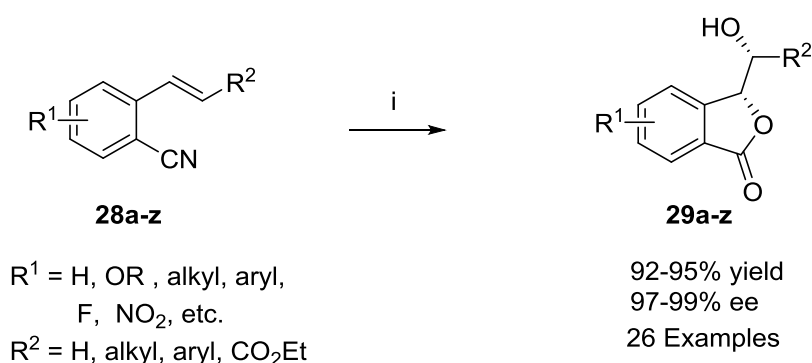
CN-Assisted Oxidative Cyclization of Cyano Cinnamates and Styrene Derivatives: A Facile Entry to 3-Substituted Chiral Phthalides and its Application to the Synthesis of (-)-Matteucen C and Butylphthalide

Sharpless asymmetric dihydroxylation (ADH) is one of the most effective methods for the preparation of chiral diols. This chapter deals with development of a novel method for the synthesis of chiral phthalides **29a-z** via CN-assisted oxidative cyclization of cyano cinnamates followed by its application in the synthesis of (-)-matteucen C and butylphthalide. This chapter is divided into two sections.

SECTION I: CN-Assisted Oxidative Cyclization of Cyano Cinnamates and Styrene Derivatives: a Facile Entry to 3-Substituted Chiral Phthalides

Chiral phthalides [isobenzofuran-1(3*H*)-ones] comprising of 5-membered lactones are found in a large number of plant products displaying broad and potent biological

activities such as anticonvulsant, anesthesia, antiischemic, antiHIV, anticancer and antibiotics.¹³ In this section, we describe a single-step oxidative cyclization of cyanocinnamates and styrene substrates that affords 3-substituted phthalides in high yields *via* synergetic acceleration of CN and osmate ester groups present in proximity positions. When ethyl 2-cyanocinnamate **28a** was subjected to a typical AD-mix- β process for 7 h, with THF as co-solvent for better solubility, the corresponding chiral phthalide **29a** was obtained exclusively in 99% ee.



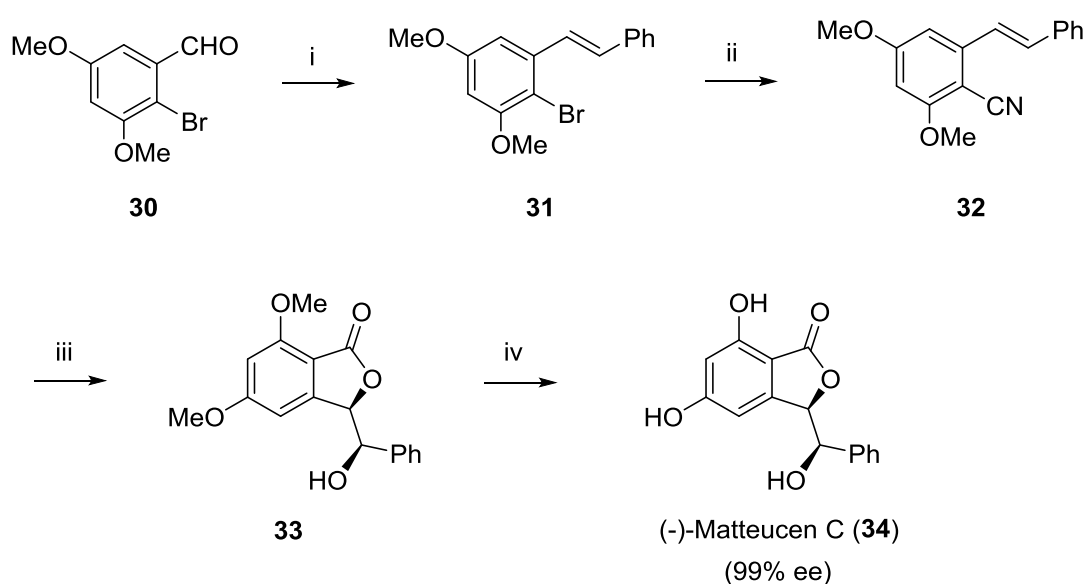
Scheme 4: (i) AD-mix- β , *tert*-BuOH:THF:H₂O (0.5:0.5:1), 25 °C, 3-7 h.

Encouraged by this result, we examined the scope of the reaction with other cyano cinnamate esters and styrene derivatives **28b-z**. In every case, the reaction proceeded rapidly in 3-7 h giving the desired phthalides **29b-z** in excellent yields and ees (upto 99%) (**Scheme 4**). The higher reactivity of cyano substituted cinnamates and styrenes were substantiated by carrying out several competitive experiments involving 1:1 molar equivalents of aromatic substrates with and without cyano substitution. The results clearly show that cyano substituted substrates react almost 10-12 times faster than the one without cyano substitution, giving excellent yields of chiral phthalides.

SECTION II: First Enantioselective Synthesis of (-)-Matteucen C and Facile Synthesis of antiIschemic Stroke Drug, 3-Butylphthalide

Matteucen C (**34**), isolated from Chinese medicinal herb, is used in the treatment of hemostatics and relieving ostalgia.¹⁴ 3-butylphthalide (**40**), is an anticonvulsant drug for the treatment of stroke.¹⁵ This section describes a short and practical enantioselective synthesis of (-)-matteucen C (**34**) and 3-butylphthalide (**40**), by employing CN-assisted oxidative cyclization of the corresponding *o*-cyano styrene

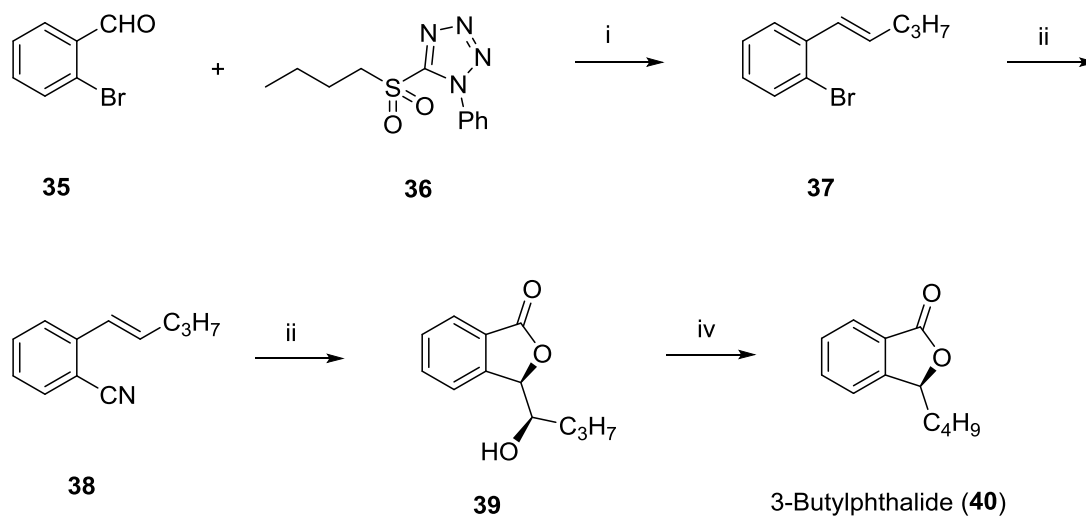
derivative **32** and **38** as the key step. Our synthesis of (-)-matteucen C commenced with 2-bromo-3,5-dimethoxy benzaldehyde (**30**), which was subjected to Wittig reaction to afford *trans*-stilbene derivative **31** in 82% yield. Rosenmund-von Braun reaction of bromo stilbene **31** gave cyano stilbene **32** with excess of CuCN under reflux conditions in DMF. Cyano stilbene **32** was then subjected to AD-mix- β process to give chiral phthalide **33** in 93% yield and 99% ee *via* CN-assisted “one-pot” oxidative cyclization. Demethylation of chiral phthalide **33** with BBr₃ in CH₂Cl₂ gave (-)-matteucen C in 43.67% overall yield and 99% ee (**Scheme 5**). Synthesis of (-)-matteucen C (**34**) was achieved for the first time, thereby confirming its structural and stereochemical assignments.



Scheme 5: (i) PhCH₂Ph₃P⁺T⁻, *n*-BuLi, THF, 0 to 25 °C, 3 h, 82%; (ii) CuCN (3.5 equiv), DMF, reflux, 14 h, 83%; (iii) AD-mix- β , *tert*-BuOH:THF:H₂O (0.5:0.5:1), 25 °C, 7 h, 93%; (iv) BBr₃, CH₂Cl₂, 25 °C, 12 h, 69%.

Similarly, our synthesis of 3-butylphthalide (**40**), started with *o*-bromobenzaldehyde (**35**), which on Julia-Kocienski olefination with butyl sulfone **36** gave (*E*)-2-bromostyrene **37** in 82% yield. Rosenmund-von Braun reaction of bromostyrene **37** with CuCN gave cyanostyrene **38**, which was then subjected to AD-mix- β process to give chiral phthalide **39** in 93% yield and 99% ee *via* CN-assisted “one-pot” oxidative

cyclization. Barton-McCombie protocol was utilized for deoxygenation of alcohol **39** to give 3-butylphthalide (**40**) in 86% yield (**Scheme 6**).



Scheme 6: (i) NaHMDS, THF, -78 °C to 25 °C, 14 h, 82%; (ii) CuCN (3.5 equiv), dry DMF, reflux, 14 h, 87%; (iii) AD-mix- β , *tert*-BuOH:THF:H₂O (0.5:0.5:1), 25 °C, 7 h, 93%; (iv) (a) 1,1-thiocarbonyldiimidazole, DMAP (10 mol%), CH₂Cl₂, 18 h; (b) AIBN (10 mol%), Bu₃SnH, toluene, reflux, 15 min, 86%.

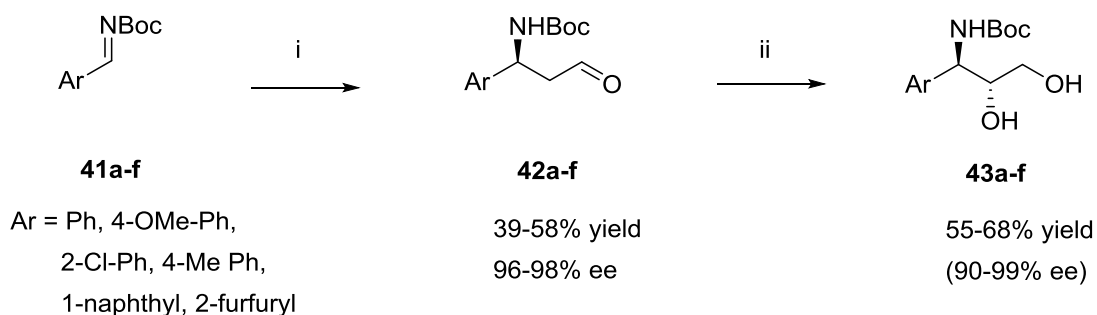
CHAPTER 3

A New Concise Method for the Synthesis of Chiral 3-Amino-1,2-Diols, (+)-*epi*-Cytosaxone and Formal Synthesis of *N*-Thiolated-2-Oxazolidinone *via* Proline-Catalyzed α -Aminooxylation of β -Aminoaldehydes

Asymmetric organocatalysis in organic chemistry has provided several new methods for obtaining chiral compounds in an environmentally benign manner. This chapter is divided into two sections. **Section I** deals with the development of novel flexible organocatalytic route to the synthesis of chiral 3-amino-1,2-diols *via* proline catalyzed α -aminooxylation of chiral β -aminoaldehydes. **Section II** describes the application of this protocol for the enantioselective synthesis of (+)-*epi*-cytosaxone and formal asymmetric synthesis of *N*-thiolated-2-oxazolidinone.

SECTION I: Proline-Catalyzed α -Aminoxylation of β -Aminoaldehydes: A Highly Stereoselective Synthesis of 3-Amino-1,2-Alkane Diols

The enantiomerically pure *syn*- or *anti*-3-amino-1,2-alkane diols¹⁶ are valuable building blocks for asymmetric synthesis of numerous biologically active natural products and pharmacologically relevant therapeutic agents such as taxol side chain, glycosphingolipids, (-)-cytoxazone, (+)-*epi*-cytoxazone, oxazolidinones, etc. Proline, an abundant, inexpensive aminoacid available in both enantiomeric forms has emerged as a practical and versatile organocatalyst, efficient for α -functionalization¹⁷ of aldehydes and ketones. In this section, an elegant and efficient protocol for the synthesis of chiral 3-amino-1,2-diols **43a-f** as single diastereomer *via* proline catalyzed α -aminoxylation of β -aminoaldehydes **42a-f** in good yields and high ee is described. Chiral pure β -aminoaldehydes **42a-f**, the starting materials for α -aminoxylation were efficiently prepared from the corresponding Boc-protected aryl aldimines **41a-f** following literature protocol (L-proline, CH₃CN, 0 °C).

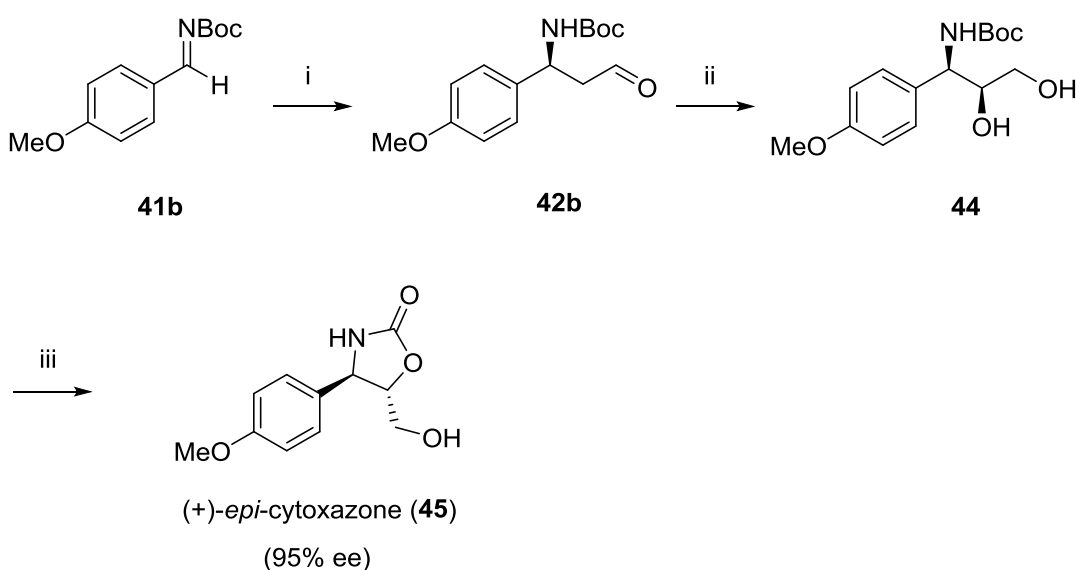


Scheme 7: (i) CH₃CHO, L-proline (20 mol%), CH₃CN, 0 °C, 3 h; (ii) (a) PhNO (0.8 equiv), L-proline (20 mol%), CH₃CN, -10 °C, 20 h then NaBH₄ CH₃OH, 10 min; (b) Cu(OAc)₂·H₂O (15 mol%), CH₃OH, 25 °C, 10 h.

Several β -aminoaldehydes **42a-f** when subjected to L-proline catalyzed α -aminoxylation with PhNO followed by its reduction with NaBH₄ gave the corresponding 3-amino-1,2-diol derivatives **43a-f**. In every case, the reaction proceeded smoothly to give a single diastereomer of *anti*-3-amino-1,2-diols **43a-f**, with the intact of excellent enantioselectivity. For instance, substrates having electron-rich or electron-deficient substituent on the aromatic ring including 1-naphthyl ring system and heteroaryl gave the desired *anti*-3-amino-1,2-diols in excellent stereoselectivity (**Scheme 7**).

SECTION II: Enantioselective Synthesis of Cytokine Modulator (+)-*epi*-Cytosaxone and Formal Synthesis of Antibacterial *N*-Thiolated-2-Oxazolidinone

(+)-*epi*-Cytosaxone (**45**) containing a novel 4,5-disubstituted-2-oxazolidinone moiety was isolated from *Streptomyces sp.*¹⁸ and was found to exhibit cytokine-modulating activity by inhibiting the signaling pathway of Th2 cells. Recently, studies have shown that *N*-thiolated 2-oxazolidinone (**50**) possesses antibacterial activity against methicillin-resistant *Staphylococcus aureus* and *Bacillus anthracis*.¹⁹ In this section, a concise protecting group-free enantioselective synthesis of (+)-*epi*-Cytosaxone (**45**) and 2-oxazolidinone (**49**) using proline catalyzed α -aminoxylation of β -amino aldehyde is presented (Scheme 8 and 9).

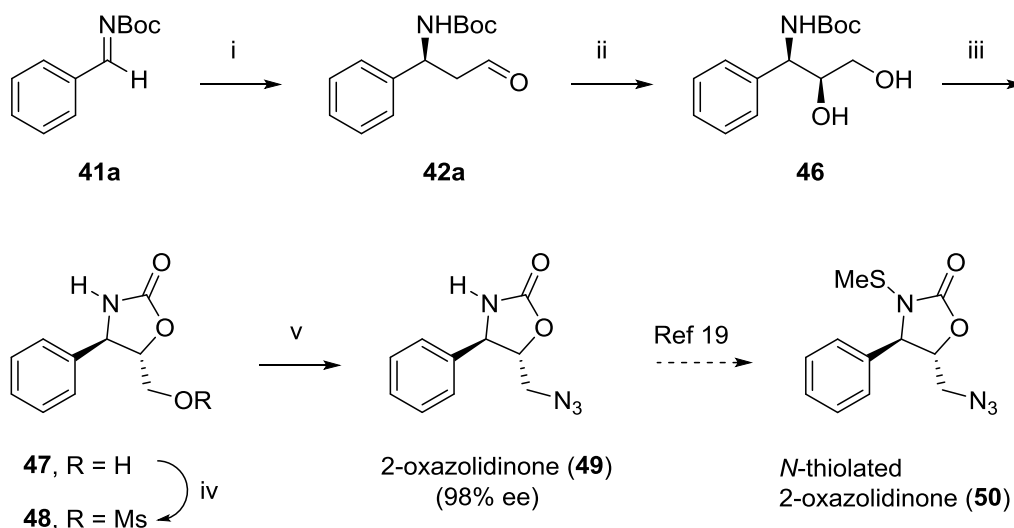


Scheme 8: (i) CH₃CHO, L-proline (20 mol%), CH₃CN, 0 °C, 3 h, 56%; (ii) (a) PhNO (0.8 equiv), D-proline (20 mol%), CH₃CN, -10 °C, 18 h; then NaBH₄, CH₃OH, 0 °C, 10 min; (b) Cu(OAc)₂·H₂O, CH₃OH, 25 °C, 16 h, 68% (over two steps); (iii) NaH, dry THF, 25 °C, 3 h, 90%.

The L-proline-catalyzed Mannich reaction of acetaldehyde with *N*-Boc-arylaldimine **41b** gave enantiopure β -aminoaldehyde **42b** in 56% yield and 98% ee.²⁰ The β -amino aldehyde **42b** was then subjected to D-proline catalyzed α -aminoxylation, which involves a two-step reaction sequence: (i) reaction of β -aminoaldehyde **42b** with

nitrosobenzene as the oxygen source in the presence of D-proline in CH₃CN at -10 °C followed by treatment with NaBH₄ in MeOH gave the crude aminoxy alcohol *in situ* and (ii) subsequent reduction of the crude aminoxy product with 30 mol% Cu(OAc)₂·H₂O yielded chiral 3-amino-1,2-diol **44** in 68% yield, >99% dr and 98% ee (over two steps). Finally, the regioselective intramolecular cyclization of **44** has been achieved to give (+)-*epi*-Cytoxazone (**45**) (Scheme 8).

The synthetic strategy utilized for the synthesis of (+)-2-oxazolidinone (**49**), precursor to *N*-thiolated-2-oxazolidinone (**50**) (Scheme 9) was similar to the one employed for (+)-*epi*-cytoxazone (**45**) (see Scheme 8). Thus oxazolidinone (**49**) was prepared in 3 steps [(i) Mannich reaction, (ii) proline-catalyzed α -aminoxylation and (iii) intramolecular cyclization] starting from imine **41a** in 33% yield (over three steps). Mesylation of primary alcohol **47** gave the mesylate **48**, which on treatment with NaN₃ in DMF at 60 °C afforded (+)-2-oxazolidinone (**49**) in 98% ee.



Scheme 9: (i) CH₃CHO, L-proline (20 mol%), CH₃CN, 0 °C, 3 h, 55%; (ii) (a) PhNO (0.8 equiv), D-proline (20 mol%), CH₃CN, -10 °C, 18 h; then NaBH₄, CH₃OH, 0 °C, 10 min; (b) Cu(OAc)₂·H₂O, CH₃OH, 25 °C, 16 h, 64% (over two steps); (iii) NaH, dry THF, 25 °C, 3 h, 94%; (iv) MsCl, Et₃N, CH₂Cl₂, 0 °C, 1 h; (v) NaN₃, DMF, 60 °C, 12 h, 82% (over two steps).

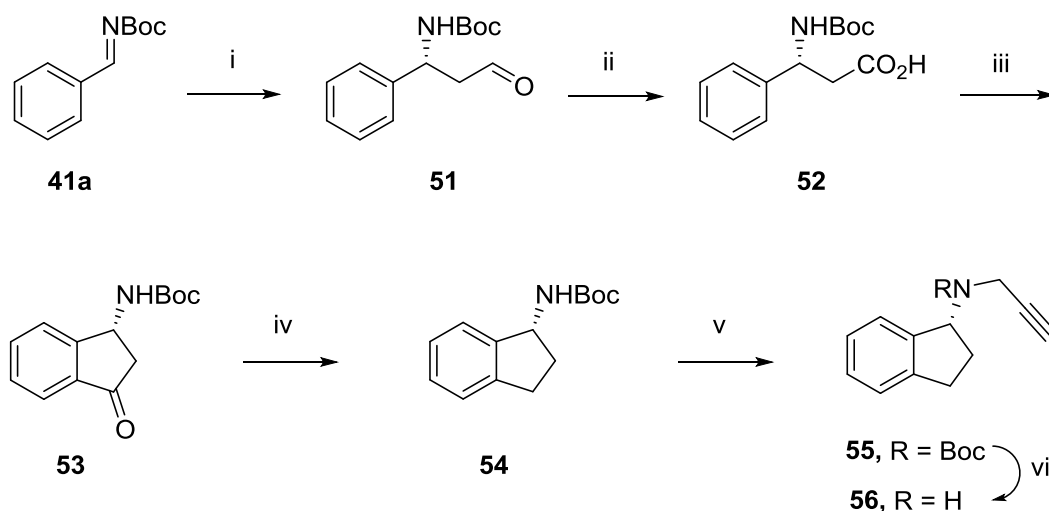
CHAPTER 4

Asymmetric Synthesis of Rasagiline and NHC-catalyzed Esterification of Aromatic Aldehydes

This Chapter is divided into two sections. **Section I** deals with the asymmetric synthesis of anti-Parkinson drug, rasagiline *via* proline-catalyzed Mannich reaction and **Section II** describes the esterification of aromatic aldehydes with alcohols under aerobic condition using *N*-heterocyclic carbene as catalyst.

SECTION I: Enantioselective Synthesis of Rasagiline an anti-Parkinson Drug

Rasagiline (Azilect) (**56**) is an irreversible inhibitor of monoamine oxidase^{21a} used as a monotherapy in early Parkinson's disease or as an adjunct therapy in more advanced cases.^{21b} The present section describes the enantioselective synthesis of rasagiline (**56**) *via* proline-catalyzed Mannich reaction, followed by Friedel-Crafts' intramolecular acylation as the key reactions (**Scheme 10**). Our synthesis of rasagiline (**56**) started from Boc-protected benzaldimine **41a**, which on Mannich reaction with acetaldehyde (D-proline, CH₃CN, 0 °C) afforded β -aminoaldehyde **51** in 62% yield and 98% ee.

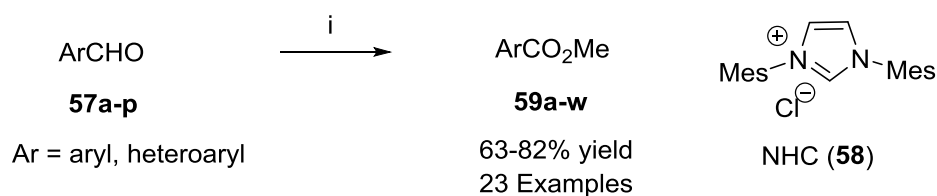


Scheme 10: (i) CH₃CHO, D-proline (20 mol%), CH₃CN, 0 °C, 3 h, 62%; (ii) NaClO₂, NaH₂PO₄, *tert*-BuOH:H₂O (5:1), 25 °C, 30 min, 93%; (iii) ClSO₃H, CH₂Cl₂, 25 °C, 2 h, then (Boc)₂O, Et₃N, cat. DMAP, CH₂Cl₂, 25 °C, 3 h, 83%; (iv) NH₂NH₂·H₂O, KOH, diethylene glycol, reflux, 4 h, 88%; (v) NaH, propargyl bromide, DMF, 25 °C, 4 h, 81%; (vi) 37% aq. HCl, dioxane, 25 °C, 30 min, 97%.

The Pinnick oxidation of β -aminoaldehyde **51** with NaClO_2 and NaH_2PO_4 gave the corresponding β -amino carboxylic acid **52**, which on Friedel-Crafts' intramolecular acylation reaction with ClSO_3H , followed by the protection of primary amine [$(\text{Boc})_2\text{O}$, Et_3N , cat. DMAP] afforded indanone **53**. Wolff-Kishner reduction of indanone **53** afforded carbamate **54** in 88% yield. Alkylation of carbamate **54** followed by subsequent deprotection of the Boc protecting group with 37% aq. HCl in dioxane furnished rasagiline (**56**).

SECTION II: *N*-Heterocyclic Carbene-Catalyzed Esterification of Aromatic Aldehydes with Alcohols under Aerobic Condition

Aromatic esters are important and useful structural elements finding tremendous applications in wide range of fields encompassing solvents, lubricants, plasticizing agents, perfumes, pharmaceuticals, agrochemicals, etc. In recent years, *N*-heterocyclic carbenes (NHCs) have emerged as an important and powerful class of organocatalysts²² with tremendous applications in a variety of synthetic transformations and as versatile ligands in transition metal catalysis. In this section, we describe a simple, organocatalytic procedure for the direct oxidative esterification of a variety of aromatic aldehydes **57a-p** with various alcohols that affords the corresponding aromatic esters **59a-w** in good yields using imidazolium NHC (**58**) as catalyst and molecular O_2 as oxidant at ambient conditions (**Scheme 11**).



Scheme 11: (i) NHC (**58**) (10 mol%), DBU (20 mol%), alcohol (1.2 equiv), O_2 (1 atm).

Several aromatic aldehydes underwent oxidative esterification with methanol smoothly under mild conditions. Remarkably, substrates with electron-withdrawing groups showed higher reactivity as compared to electron-releasing substituents. Heteroaromatic aldehydes such as 3-pyridine carboxaldehyde and furfural also gave the corresponding esters in high yields.

A wide range of alcohols were then examined for oxidative esterification with 4-nitrobenzaldehyde as the substrate, both primary and secondary alcohols including allylic, propargylic and benzylic alcohols underwent this reaction to give the corresponding esters in excellent yields. In order to gain insight into the mechanistic details of the reaction, control experiments were carried out and based on result and literature precedents,²³ a catalytic cycle was proposed.

References:

1. Tokunaga, M.; Larrow, J. F.; Kakiuchi, F.; Jacobsen, E. N. *Science* **1997**, *277*, 936.
2. (a) Yu, Q.; Wu, Y.; Xia, L. J.; Tang, M. H.; Wu, Y. L. *J. Chem. Soc. Chem. Commun.* **1999**, 129; (b) Kumar, P.; Naidu, S. V.; Gupta, P. *Tetrahedron* **2007**, *63*, 2745.
3. Kumar, P.; Gupta, P. *Synlett* **2009**, *9*, 1367.
4. (a) Talapatra, S. K.; Basu, D.; Talapatra, B. *Ind. J. Chem. Sect. B* **1985**, *24B*, 29; (b) Lan, Y. H.; Chang, F. R.; Liaw, C. C.; Wu, C. C.; Chiang, M. Y.; Wu, Y. C. *Planta Med.* **2005**, *71*, 153.
5. (a) Tsubuki, M.; Kanai, K.; Honda, T. *J. Chem. Soc., Chem. Commun.* **1992**, 1640; (b) Tsubuki, M.; Kanai, K.; Nagase, H.; Honda, T. *Tetrahedron* **1999**, *55*, 2493.
6. Keck, G. E.; Boden, E. P. *Tetrahedron Lett.* **1984**, *25*, 265.
7. Reddy, R. S.; Chouthaiwale, P. V.; Suryavanshi, G.; Chavan, V. B.; Sudalai, A. *Chem. Commun.* **2010**, *46*, 5012.
8. Langille, N. F.; Panek, J. S. *Org. Lett.* **2004**, *6*, 3203.
9. Arison, B. H.; Hwang, S. B. *Acta Pharmacol. Sin.* **1991**, *5*, 345.
10. Lee, A. L.; Ley, S. V. *Org. Biomol. Chem.* **2003**, *1*, 3957.
11. Zhang, H. J.; Tamez, P. A.; Hoang, V. D.; Tan, G. T.; Van Hung, N.; Xuan, L. T.; Huong, L. M.; Cuong, N. M.; Thao, D. T.; Soejarto, D. D.; Fong, H. H. S.; Pezzuto, J. M. *J. Nat. Prod.* **2001**, *64*, 772.
12. Choppin, S.; Colobert, F.; Hanquet, G.; Leroux, F.; Mahmoudi, N.; Mazier, D. *EP 2177496 A1/2010*.
13. For recent reviews of synthesis and biological applications of phthalides, see: (a) Lin, G.; Chan, S. S. -K.; Chung, H. S.; Li, S. L. *Stud. Nat. Prod. Chem.* **2005**, 611; (b) Beck, J. J.; Chou, S. C. *J. Nat. Prod.* **2007**, *70*, 891; (c) Liu, J.; Li, F.; Kim, E. L.; Li, J. L.; Hong, J.; Bae, K. S.; Chung, H. Y.; Kim, H. S.; Jung, J. H. *J. Nat. Prod.* **2011**, *74*, 1826.
14. Shao, P.; Zhang, W.; Li, B.; Jiao, W. H.; Wu, L. J.; Yao, X. S. *Chem. Pharm. Bull.* **2010**, *58*, 1650.
15. (a) Li, J.; Si, Y.; Che, Y. *Org. Lett.* **2011**, *13*, 2670; (b) Barton, D. H. R.; Devries, J. X. *J. Chem. Soc.* **1963**, 1916; (c) Sato, H.; Yorozu, H.; Yamaoka, S. *Biomed. Res. Trace Elem.* **1993**, *14*, 385.
16. (a) Bloch, R. *Chem. Rev.* **1998**, *98*, 1407 and references cited therein; (b) Hutin, P.; Larcheveque, M. *Tetrahedron Lett.* **2000**, *41*, 2369; (c) Righi, G.; Chionne, A.; Bonini, C. *Eur. J. Org. Chem.* **2000**, 3127; (d) Righi, G.; Ronconi, S.; Bonini, C. *Eur. J. Org. Chem.* **2002**, 1573; (e) Bickley, J.

-
- F.; Roberts, S. M.; Runhui, Y.; Skidmore, J.; Smith, C. B. *Tetrahedron*, **2003**, *59*, 5731.
17. (a) Kotkar, S. P.; Chavan, V. B.; Sudalai, A. *Org. Lett.* **2007**, *9*, 1001; (b) Kotkar, S. P.; Sudalai, A. *Tetrahedron Lett.* **2006**, *47*, 6813; (c) Rawat, V.; Chouthaiwale, P. V.; Chavan, V. B.; Suryavanshi, G.; Sudalai, A. *Tetrahedron Lett.* **2010**, *51*, 6565.
18. Kakeya, H.; Morishita, M.; Osono, M.; Ishizuka, M.; Osada, H. *J. Antibiot.* **1998**, *51*, 1126.
19. (a) Mishra, R. K.; Revell, K. D.; Coates, C. M.; Turos, E.; Dickeyb, S.; Limb, D. V. *Bioorg. Med. Chem.* **2006**, *16*, 2081; (b) Steven, R. W.; Hisao, I.; Marvin, J. M.; *Tetrahedron Lett.* **1985**, *26*, 3891.
20. Yang, J. W.; Chandler, C.; Stadler, M.; Kampen, D.; List, B. *Nature* **2008**, *452*, 453.
21. (a) Oldfield, V.; Keating, G. M.; Perry, C. M. *Drugs* **2007**, *67*, 1725; (b) Gallagher, D. A.; Schrag, A. *CNS Drugs* **2008**, *22*, 563; (c) Binda, C.; Hubalek, F.; Li, M. *J. Med. Chem.*, **2005**, *48*, 8148.
22. (a) Moore, J. L.; Rovis, T. *Top. Curr. Chem.* **2009**, *291*, 77; (b) Phillips, E. M.; Chan, A.; Scheidt, K. A. *Aldrichimica Acta* **2009**, *42*, 55; (c) Nair, V.; Vellalath, S.; Babu, B. P. *Chem. Soc. Rev.* **2008**, *37*, 2691; (d) Enders, D.; Niemeier, O.; Henseler, A. *Chem. Rev.* **2007**, *107*, 5606.
23. (a) Arde, P.; Ramanjaneyulu, B. T.; Reddy, V.; Saxena, A.; Anand, V. *Org. Biomol. Chem.* **2012**, *10*, 848; (b) Meng, J. J.; Gao, M.; Wei, Y. P.; Zhang, W. Q. *Chem. Asian. J.* **2012**, *7*, 872; (c) Phillips, E. M.; Riedrich, M.; Scheidt, K. A. *J. Am. Chem. Soc.* **2010**, *132*, 13179.

CHAPTER 1

**Short Enantioselective Synthesis of (+)-
Goniodiol and (-)-Polysphorin Analog using Co-
Catalyzed HKR of Alkoxy Epoxides**

Section I:

Concise Enantioselective Synthesis of (+)-Goniodiol, a Cytotoxic Lactone via Co-Catalyzed Hydrolytic Kinetic Resolution of *anti*-(2*SR*, 3*RS*)-3-Methoxy-3-phenyl-1,2-epoxy propane

1.1.1 Introduction

The genus *Goniothalamus* (Annonoaceae) consists of over 115 species of shrubs and treelets growing abundantly in the rain forests of tropical Asia.¹ The extracts and leaves of *Goniothalamus* species have traditionally been used as folk medicine: e.g. in the treatment of edema and rheumatism,² abortifacient,³ labor pain,⁴ etc. Recently, (+)-goniodiol (**1**), belonging to styryllactone family, was isolated from *Goniothalamus sesquipedalis*.⁴

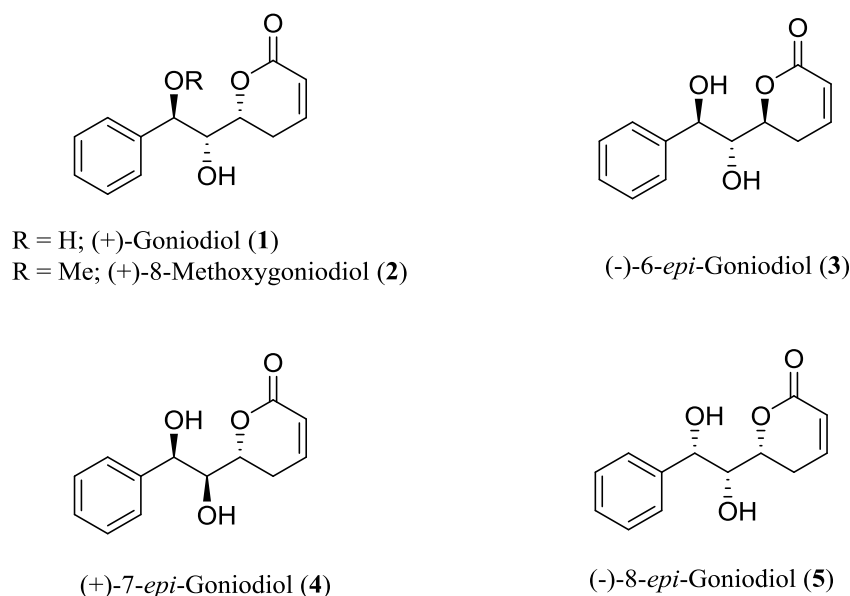


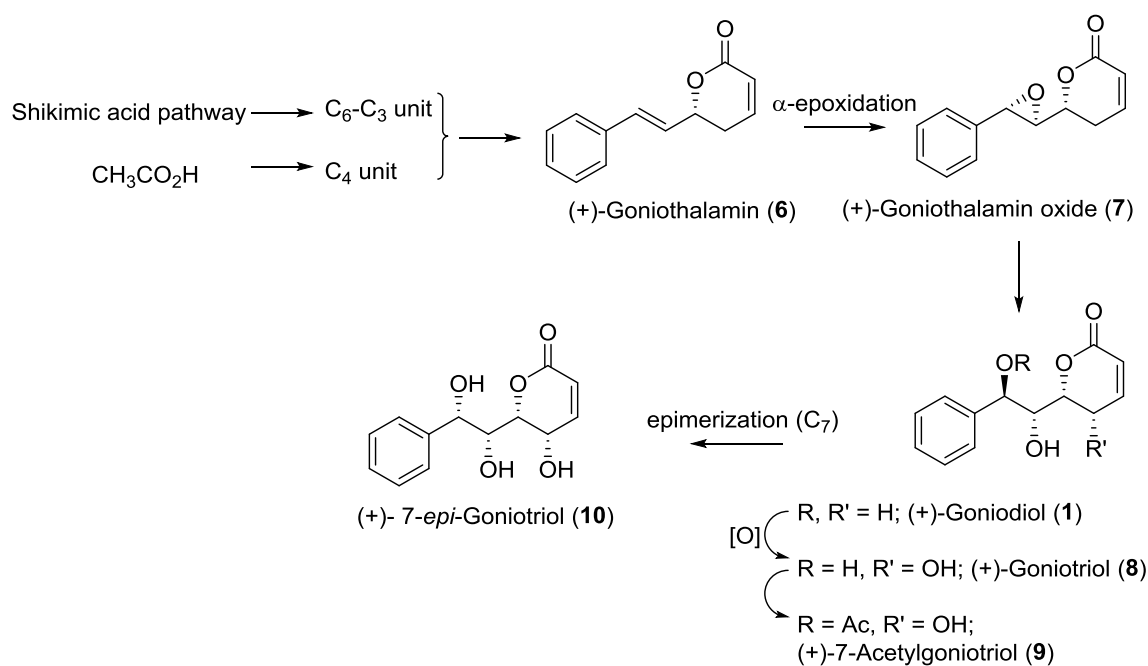
Fig. 1: Structures of bioactive styryllactones

Fig. 1 shows the structures of some of the isomers of goniodiol **1-5**. These bioactive natural products (**1-5**) with relatively small and densely functionalized molecules are

found to exhibit significant cytotoxic activity⁵ including antimicrobial and antifungal properties as well as antibiotic potential. In particular, (+)-goniodiol (**1**) was found to show significant cytotoxic selectivity against several human tumor cell lines e.g. human lung carcinoma (A-549) (ED_{50} 1.22×10^{-1} $\mu\text{g/ml}$).^{4a,6} Its unique structural features coupled with potent biological activities make them ideal targets for testing new synthetic methodology. Consequently, there have been consistent efforts toward styryllactones **1-5** resulting in a number of their total synthesis.⁷⁻¹⁸

1.1.2 Pharmacology and Mode of Action

The structure and relative configuration of (+)-goniodiol (**1**) were established by NMR spectral studies^{4a,6} and X-ray crystallography.⁶ (+)-Goniodiol (**1**) contains a 6-substituted 5,6-dihydro- α -pyrone moiety, which is widely distributed in the plant kingdom. Natural products containing this lactone unit also possess a wide range of biological activity such as insect antifeedants, antifungal, plant growth inhibitors, etc.



Scheme 1: Proposed biosynthetic pathways for styryllactones

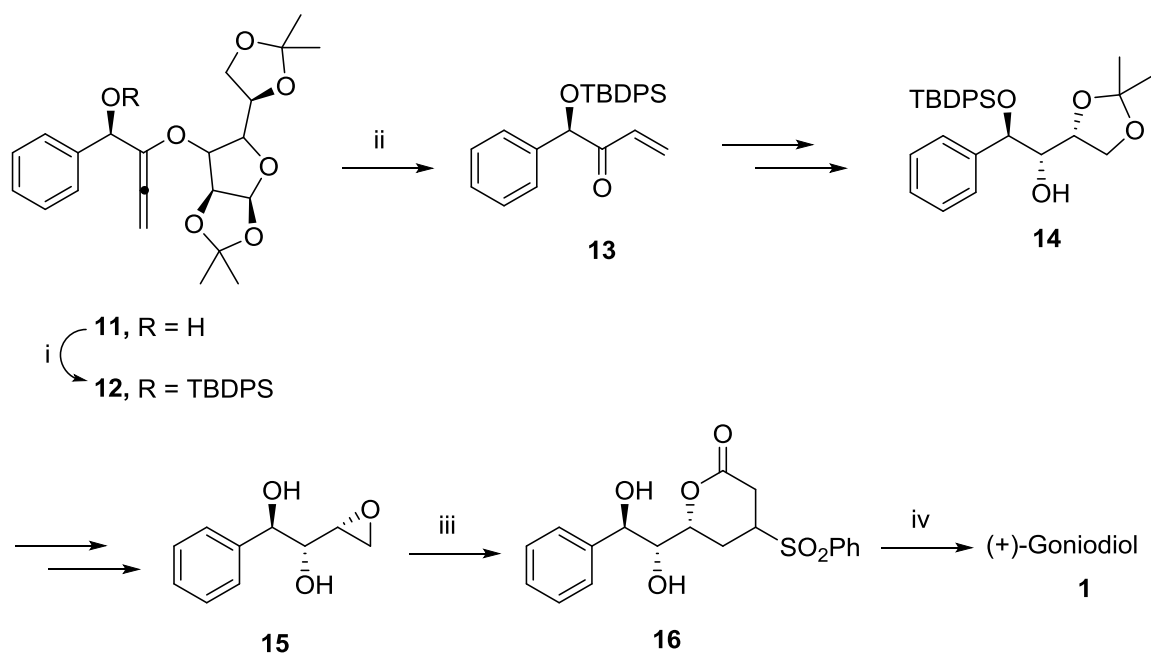
The α,β -unsaturated δ -lactone functionality is presumed to be responsible for the biological activities, due to its ability to act as a Michael acceptor, enabling these molecules to bind to a target enzyme. Literature reveals that (+)-goniodiol (**1**) is considered to be the biosynthetic precursor to other styryllactones. Thus, (+)-goniodiol (**1**) may be more important and can be an immediate precursor to many higher order functionalized styryllactones **8-10** (**Scheme 1**).¹⁹

1.1.3 Review of Literature

Literature search revealed that several reports are available for the synthesis of (+)-goniodiol (**1**) involving chiral pool, chemo-enzymatic approach or enantioselective syntheses, several of which are described below.

Vatele's approach (1996)⁷

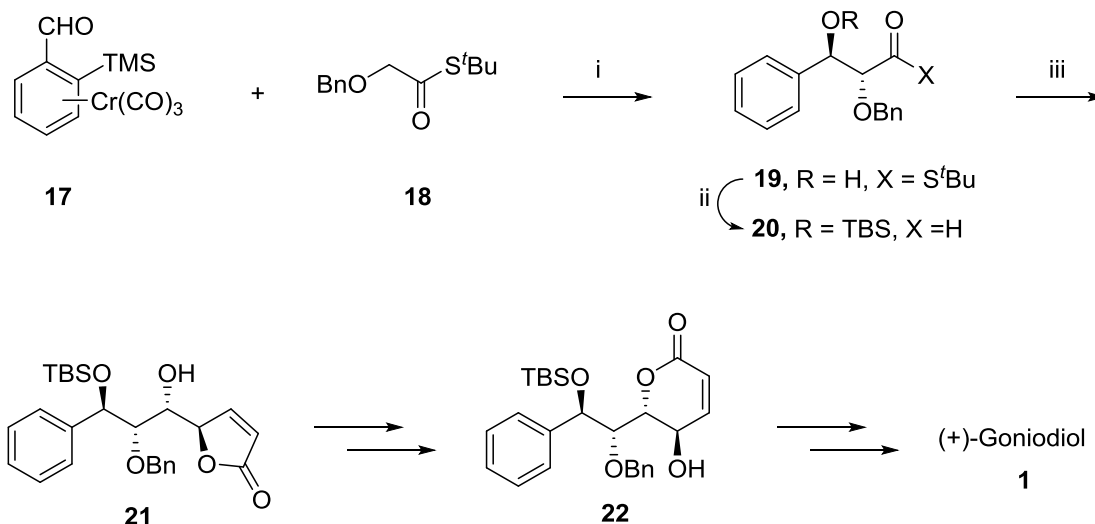
Vatele *et al.* have achieved the synthesis of (+)-goniodiol (**1**) using a chiral pool approach commencing from allenic alcohol **11**, which was derived from D-glucose by a diastereoselective addition of the lithium salt of 3-O-allenyl diacetone-D-glucose to benzaldehyde. The allenic alcohol **11** was subjected to O-silylation **12**, followed by acidic hydrolysis to give unsaturated ketol **13**. Protected ketol **13** was converted to alcohol **14** (dr = 92:8) using simple standard transformations, where diastereoselective dihydroxylation and reduction using L-Selectride served as key steps. Epoxide **15** was obtained from alcohol **14** in four linear steps. Further, epoxide **15** was treated with lithio derivative of methyl 3-phenylsulfonyl orthopropionate in the presence of $\text{BF}_3 \cdot \text{Et}_2\text{O}$ to give the sulfonyl lactone **16**. Finally, DBU-induced elimination gave (+)-goniodiol (**1**) (**Scheme 2**).



Scheme 2: (i) *t*-BuPh₂SiCl, DMAP, CH₂Cl₂, 3 days; (ii) 50% CF₃CO₂H-CH₂Cl₂, 3 h, 81% (over 2 steps); (iii) (a) 2-methoxypropene, camphorsulfonic acid, CH₂Cl₂, 10 min, 25 °C, 68%; (b) methyl-3-phenylsulfonyl orthopropionate, *n*-BuLi, BF₃·Et₂O, THF, -78 °C, 2 h; then 3M H₂SO₄, 50 °C, 3 h; (iv) DBU, CH₂Cl₂, 1 h, 0 °C, 60%.

Hanaoka's approach (1997)⁸

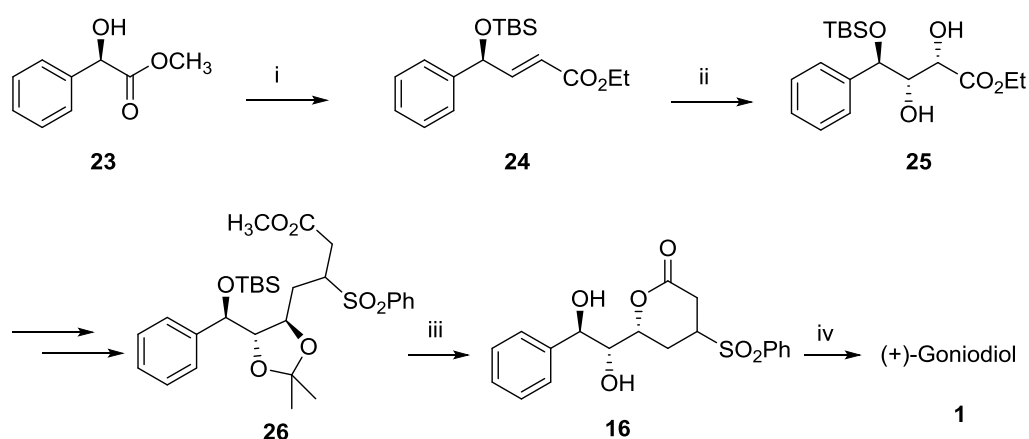
Hanaoka *et al.* have reported a useful method of synthesis of (+)-goniodiol **1** using a highly stereoselective asymmetric aldol reaction of chromium complexed *o*-TMS-benzaldehyde **17** as key step. Reaction of aldehyde **17** with ethanethioate **18** in the presence of TiCl₄ afforded the *anti*-aldol product **19** in 59% yield. *anti*-Aldol product **19** was then converted to aldehyde **20** using simple transformations; Further, aldol reaction of aldehyde **20** with 2-trimethylsilyloxy furan in the presence of Ti(O^{*i*}Pr)₂Cl₂ produced δ -lactone **21** in 56% yield. Five-membered δ -lactone **21** was then converted to six-membered lactone **22** in two steps, which was finally transformed to (+)-goniodiol (**1**) in five linear steps (**Scheme 3**).



Scheme 3: (i) TiCl₄, Et₃N, CH₂Cl₂, -78 °C, 2 h then TBAF- HF, CH₃CN:THF, -78 °C to 0 °C, 30 min then hv, Et₂O, 0 °C, 2 h, 67%; (ii) (a) thallium trinitrate, CH₃OH, 4 h, 71%; (b) TBSCl, imid., CH₂Cl₂, 71%; (c) DIBAL-H, benzene, 25 °C, 3 h; (iii) Ti(OⁱPr)₂Cl₂, 2-trimethyl silyloxyfuran, CH₂Cl₂, -78 °C, 2 h, 56%.

Vatele's approach (1998)⁹

Vatele *et al.* have commenced their synthesis from commercially available methyl (*R*)-mandelate **23**.

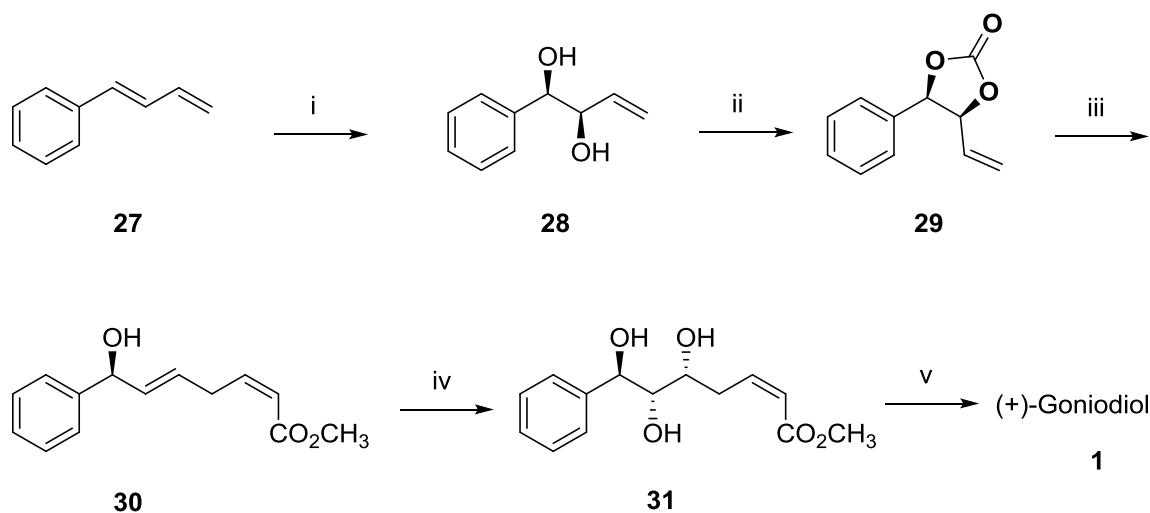


Scheme 4: (i) (a) TBDMSCl, imid., DMF, 25 °C, 12 h, 98%; then DIBAL-H, Et₂O, hexane, -78 °C, 20 min, 82%; (b) Ph₃P=CHCO₂Et, toluene, reflux, 30 min, 94%; (ii) cat. OsO₄, NMO, acetone:H₂O (5:1), 25 °C, 5 h, 85%; (iii) CF₃CO₂H:H₂O (4:1), 25 °C, 18 h, 83 %; (iv) DBU, CH₂Cl₂, 0 °C, 1 h, 97 %.

The (*E*)-unsaturated ester **24** was obtained from methyl (*R*)-mandelate **23** by sequential protection of the hydroxy group, reduction of ester and Wittig reaction. Ester **24** on OsO₄-catalyzed diastereoselective dihydroxylation gave dihydroxy ester **25**, with high *anti* selectivity (89:11 ratio) in 85% yield. Sulfone **26** obtained from dihydroxy ester **25** in four steps; was then converted to known intermediate **16** under acidic condition. Finally, DBU-induced elimination afforded goniodiol (**1**) (**Scheme 4**).

Lin's approach (2002)¹⁰

Lin *et al.* have reported the synthesis of (+)-goniodiol (**1**) by employing Sharpless asymmetric dihydroxylation as the key reaction. Thus, (*E*)-1-phenyl-1,3-butadiene **27** was subjected to regioselective asymmetric dihydroxylation, which resulted in a slightly greater preference for the internal double bond over terminal double bond with a ratio of 2.2 to 1 and an ee of 95% for diol **28**.



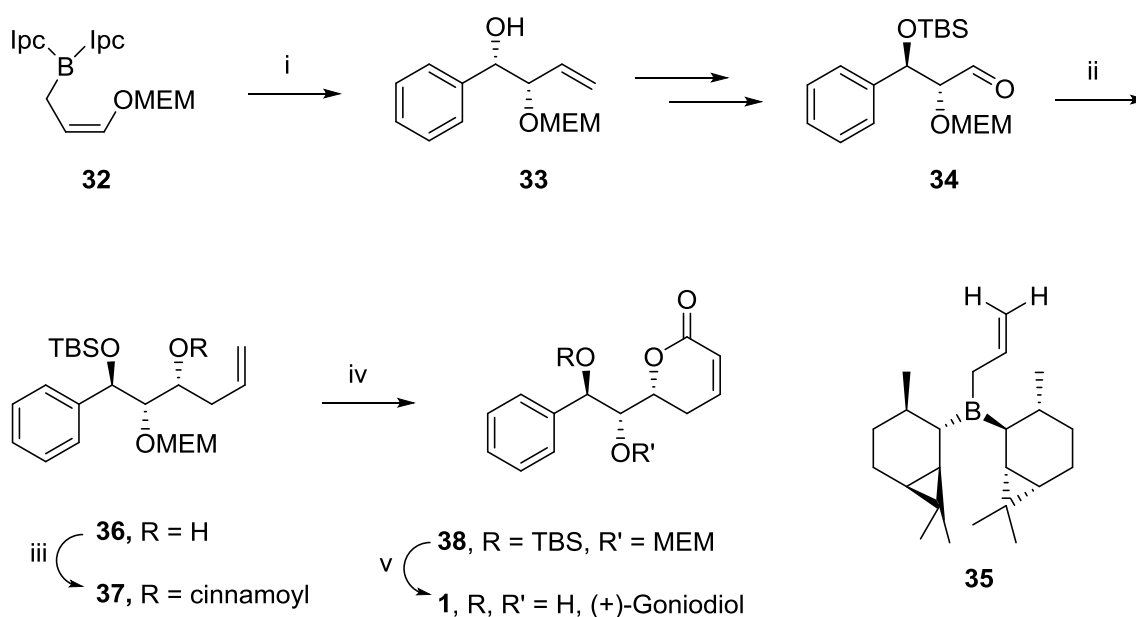
Scheme 5: (i) AD-mix- β , CH₃SO₂NH₂, 0 °C, 30 h, 58%; (ii) triphosgene, Et₃N, CH₂Cl₂, 0 °C, 2 h, 98%; (iii) cat. PdCl₂(CH₃CN)₂, (*Z*)-vinyltributylstannane, DMF:H₂O (4:1), 0 °C, 1 h, 71%; (iv) AD-mix- β , CH₃SO₂NH₂, 0 °C, 22 h, 83%; (v) *p*-TSA, toluene, 80 °C, 84%.

Protection of diol **28** with triphosgene furnished allylic cyclic carbonate **29**, which was coupled with (*Z*)-vinyltributylstannane in the presence of PdCl₂(CH₃CN)₂ (5

mol%) to afford the (*E*)-substituted allylic alcohol **30**. Sharpless dihydroxylation of **30** with AD-mix- β furnished the triol **31** (de 96:4). Finally, lactone formation under acidic condition provided (+)-goniodiol (**1**) (Scheme 5).

Ramachandran's approach (2002)¹¹

Ramachandran *et al.* have employed asymmetric alkoxyallylboration as the key reaction. Thus, alkoxyallylboration of benzaldehyde with (+)-*B*- γ -methoxyethoxy methoxyallyl diisopinocampheylborane **32** afforded α -alkoxyhomoallylic alcohol **33** in 71% yield and 98% ee.



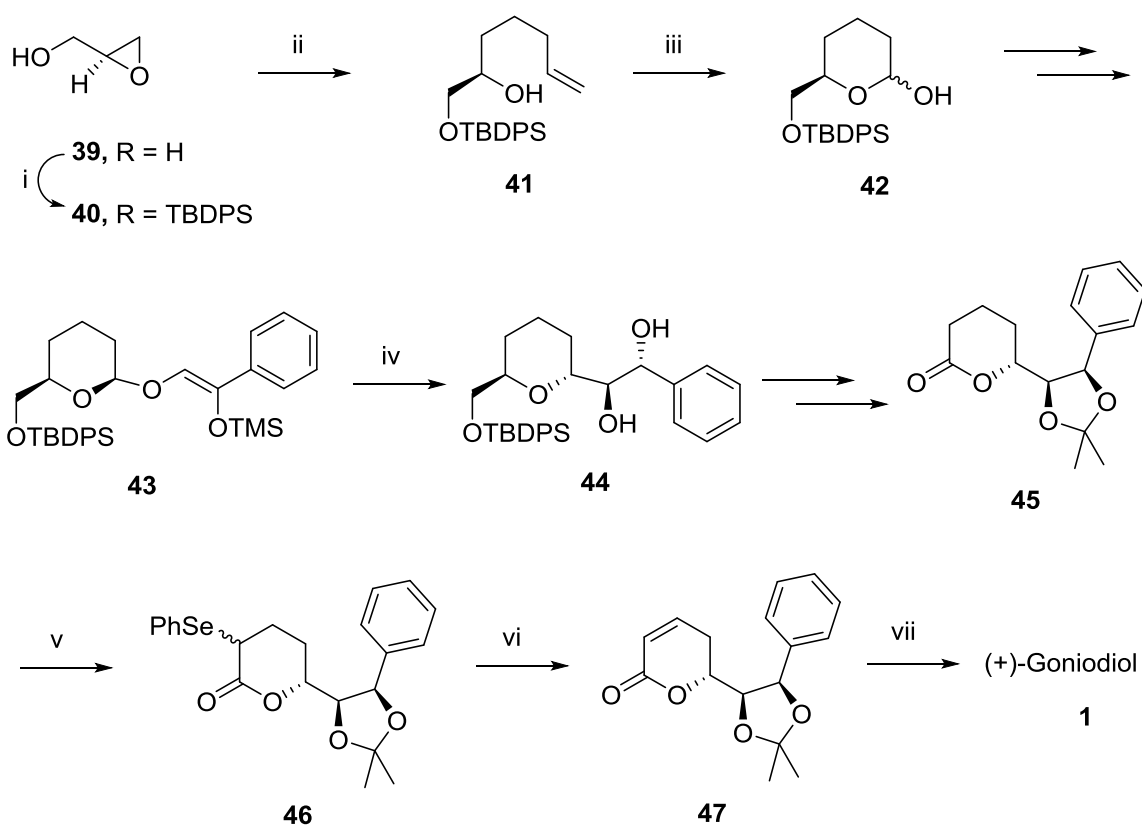
Scheme 6: (i) (a) PhCHO, -100 °C; (b) NaOH/H₂O₂, 25 °C, 71%; (ii) (a) (+)-**35**, ether:pentane, -100 °C; (b) NaOH, H₂O₂, 25 °C, 6 h, 76%; (iii) (*E*)-cinnamoyl chloride, pyridine, DMAP, CH₂Cl₂, 0 °C, 15 h, 63%; (iv) Grubbs' I generation catalyst, toluene, 120 °C, 3 h, 76%; (v) HCl:THF(1:8), 65%.

With simple transformations, α -alkoxyhomoallylic alcohol **33** was converted to aldehyde **34**, allylboration of which with **35**, provided the corresponding homoallylic alcohol **36** in 76% yield and 92% diastereomeric excess. The alcohol **36** was converted as its cinnamate ester **37** followed by ring closing metathesis with Grubbs'

first generation catalyst provided α -pyrone **38**. Both TBS and MEM groups were deprotected in a single step with HCl in THF to afford (+)-goniodiol (**1**) (Scheme 6).

Ley's approach (2006)¹²

Ley *et al.* have achieved the synthesis of (+)-goniodiol (**1**) using a chiral pool approach starting from (*S*)-(-)-glycidol **39**. Silyl protection of alcohol and subsequent regioselective addition of but-3-enylmagnesium bromide afforded alkenol **41**.



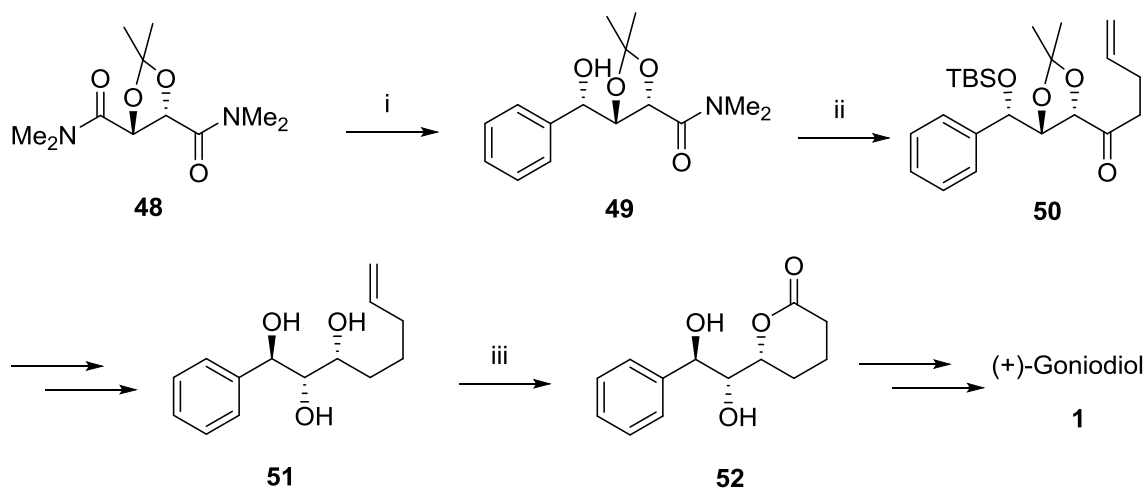
Scheme 7: (i) TBDPSCl, DMAP, Et₃N, CH₂Cl₂, 25 °C, 3 days, 86%; (ii) CH₂=CH(CH₂)₂MgBr, Li₂CuCl₄, THF, -30 °C, 100%; (iii) O₃, CH₂Cl₂, -78 °C, then PPh₃, 25 °C, 100%; (iv) (a) TMSOTf, CH₂Cl₂, -30 °C, 43%; (b) NaBH₄, CH₃OH, 0 °C, 100%; (v) LDA, THF, -78 °C then PhSeBr; (vi) CH₂Cl₂:30% aq. H₂O₂ (2 : 1), 0 °C, 10 min, 82% (over 2 steps); (vii) 50% aq. AcOH, 80 °C, 30 min, 97%.

Ozonolysis of olefin in **41** followed by reductive work-up gave lactol **42**. Lactol **42** was then converted to *anti*-diol **44** (d.e. = 95%) through silyl enol ether **43** using

simple transformations, where Grignard addition and diastereoselective reduction served as key steps. Subsequent protection and oxidations produced lactone **45** in five steps from *anti*-diol **44**. Introduction of α,β -unsaturation into the lactone molecule was achieved efficiently in two steps *via* sequential treatment with PhSeBr under basic condition to give selenide **46**, followed by oxidative elimination with H₂O₂ resulting in lactone **47**. Finally, deprotection of acetonide group gave (+)-goniodiol (**1**) (Scheme 7).

Prasad's approach (2007)¹³

Prasad *et al.* have employed chiral pool approach commencing from *D*-(-)-iso propylene dioxy tartaric amide **48**, which on reaction with PhMgBr and stereoselective reduction under Luche condition resulted in the formation of alcohol **49** (dr = 94:6) in 82% yield.



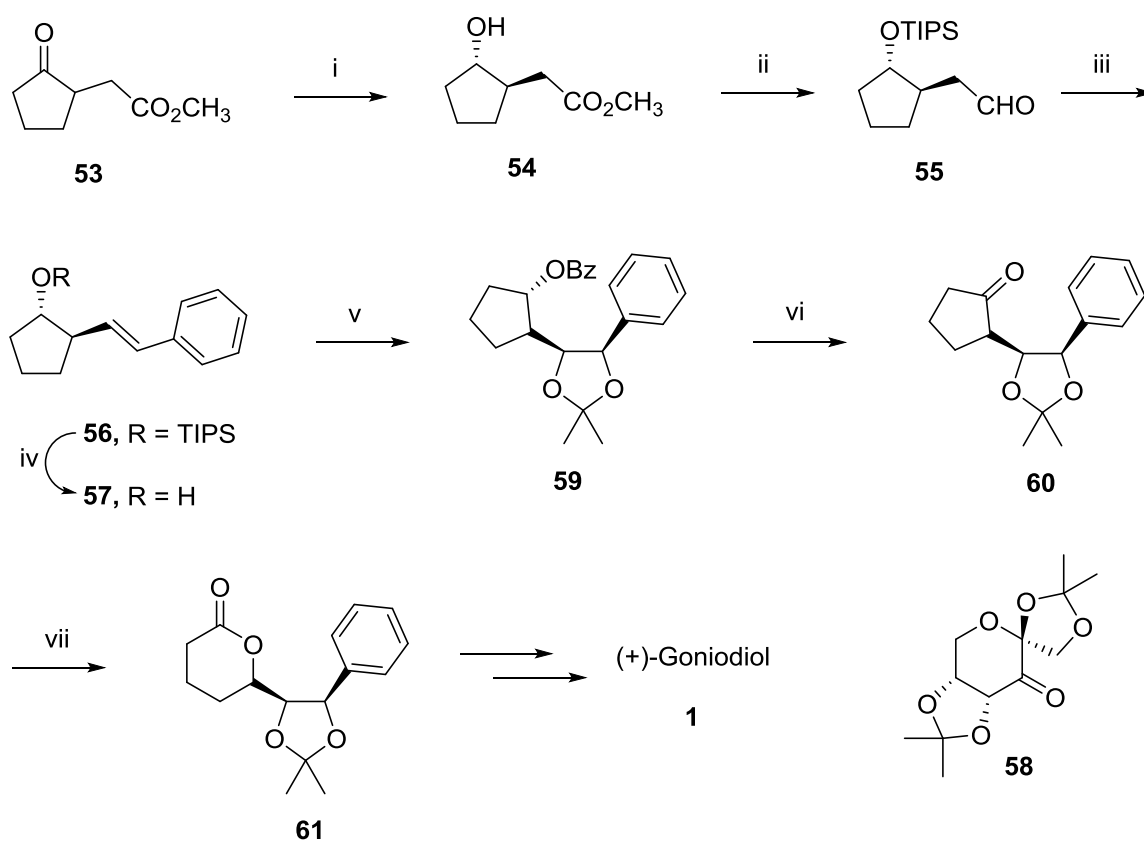
Scheme 8: (i) (a) PhMgBr, THF, -10 °C, 0.5 h, 92%; (b) NaBH₄, CeCl₃, -78 °C, 2 h; (ii) (a) TBDMSCl, imid., DMAP, DMF, 25 °C, 6 h, 90%; (b) 3-butenylmagnesium bromide, -10 °C, THF, 0.5 h, 93%; (iii) (a) O₃/Me₂S, CH₂Cl₂:MeOH, -78 °C, 6 h; (b) Ag₂CO₃/Celite, toluene reflux, 0.5 h, 76%.

Protection of the benzylic hydroxy group followed by the addition of 3-butenyl magnesium bromide afforded ketone **50**. Triol **51** was obtained from ketone **50** using simple transformations, where in Barton-McCombie deoxygenation and Mitsunobu

reactions served as key steps. Ozonolysis of olefin moiety followed by oxidation of the resulting lactol with Ag_2CO_3 , gave lactone **52**. Selenation and deselenation of lactone **52** resulted in α,β -unsaturated lactone, which afforded (+)-goniodiol (**1**) (Scheme 8).

Yamauchi's approach (2008)¹⁴

Yamauchi *et al.* have reported the synthesis of (+)-goniodiol (**1**) by employing the enzymatic reduction of keto ester as the key reaction.

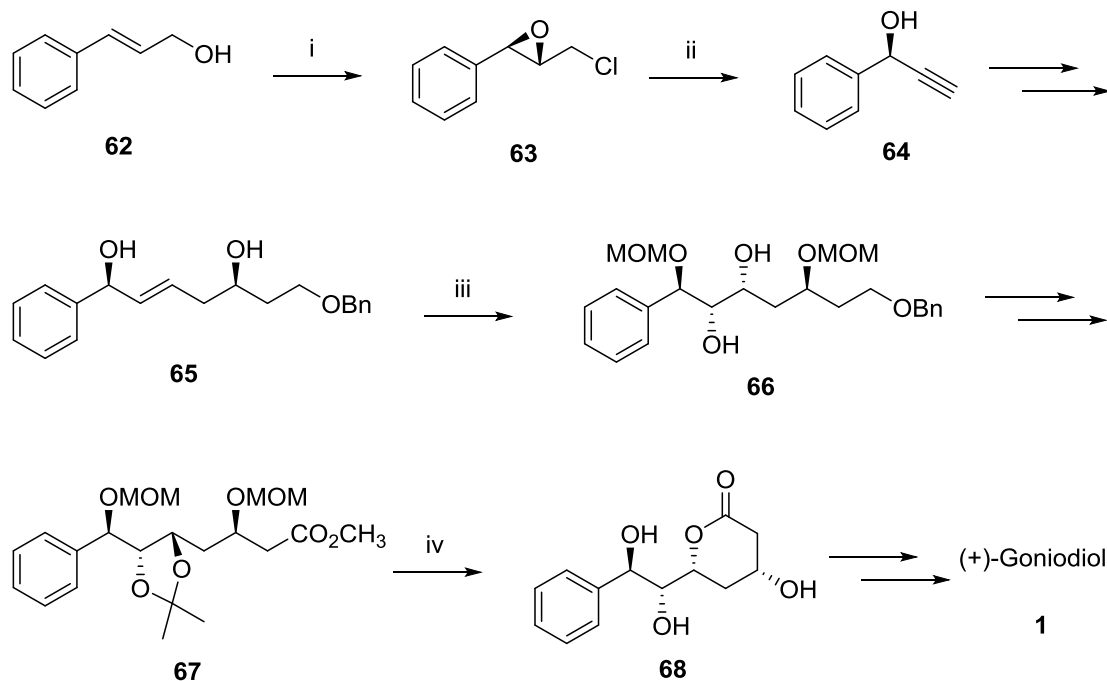


Scheme 9: (i) Baker's yeast, 30 °C, 48 h, 36%; (ii) (a) TIPSOTf, 2,6-lutidine, CH_2Cl_2 , 25 °C, 20 h, 99%; (b) LiAlH_4 , 0 °C, 1 h; (c) PCC, CH_2Cl_2 , 25 °C, 16 h, 82%; (iii) (a) PhMgBr , 0 °C, 20 min, 94%; (b) KHSO_4 , toluene reflux, 1.5 h, 84 %; (iv) TBAF, THF, 25 °C, 4 h, 100%; (v) (a) ketone **58**, 25 °C, 18 h, 83%; (b) BzCl , pyridine, 25 °C, 9 h, 98%; (c) 2-methoxypropene, *p*-TSA, DMF, 25 °C, 8 h, 41%; (vi) (a) NaOH , EtOH, 25 °C, 14 h, 97 %; (b) PCC, CH_2Cl_2 , 25 °C, 22 h, 91% (over 2 steps); (vii) *m*CPBA, CH_2Cl_2 , 25 °C, 8 h, 74%.

Thus, the racemic keto ester **53** was subjected to enzymatic reduction with Baker's yeast to give (1*R*, 2*S*)-hydroxy ester **54** in 36% yield. The hydroxy group of **54** was protected as its silyl ether, followed by reduction of ester group and oxidation of alcohol with PCC gave aldehyde **55**. Aldehyde **55** was subjected to Grignard reaction followed by dehydration, giving *trans*-olefin **56** as a single isomer. Desilylation and treatment of olefin **57** with D-fructose-derived ketone **58** gave epoxides as an inseparable diastereomers in 83% yield. Pure stereoisomer **59** was however obtained after benzylation; Hydrolysis of epoxide to diol under acidic condition followed by its protection as acetonide was carried out. Base hydrolysis followed by PCC oxidation of **59** gave ketone **60**. Ketone **60** was then converted to the known lactone **45** by Baeyer-Villiger oxidation, thus constituting a formal synthesis of (+)-goniodiol (**1**) (Scheme 9).

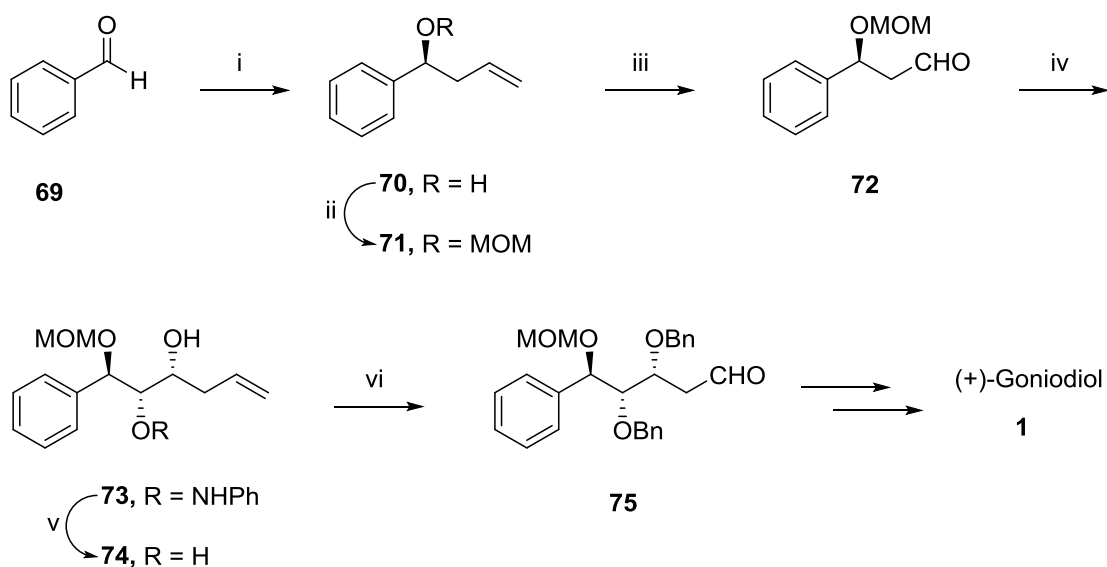
Yadav's approach (2008), (2011)^{15,16}

Yadav *et al.* have achieved the synthesis of (+)-goniodiol (**1**) by employing Sharpless asymmetric epoxidation as the key reaction. Thus, Sharpless asymmetric epoxidation of cinnamyl alcohol **62** gave the desired epoxyalcohol in 82% yield, which was subsequently converted to chiral propargylic alcohol **64** in 72% yield. Regioselective ring opening of (*S*)-2-(2-(benzyloxy)ethyl)oxirane by reaction with lithium acetylide from **64**; followed by reduction with LiAlH₄ gave allylic alcohol **65**. Protection of diol **65** as its MOM ether and Sharpless asymmetric dihydroxylation afforded diol **66** in 80% yield. Ester **67** was obtained from diol **66** using simple transformations in four steps. Cyclization of ester **67** was achieved using *p*-TSA to afford lactone **68**, which was further converted to (+)-goniodiol (**1**) by elimination of hydroxyl group in the lactone moiety (Scheme 10).



Scheme 10: (i) (a) D-(-)-DET, $\text{Ti}(\text{O}^i\text{Pr})_4$, TBHP, CH_2Cl_2 , 30 °C, 82%; (b) Ph_3P , CCl_4 , NaHCO_3 , 88%; (ii) *n*-BuLi, THF, 72%; (iii) (a) MOMCl, Hunig's base, CH_2Cl_2 , 25 °C, 92%; (b) AD-mix- β , $\text{CH}_3\text{SO}_2\text{NH}_2$, *tert*-BuOH:H₂O (1:1), 0 °C; (iv) MeOH, *p*-TSA, 78%.

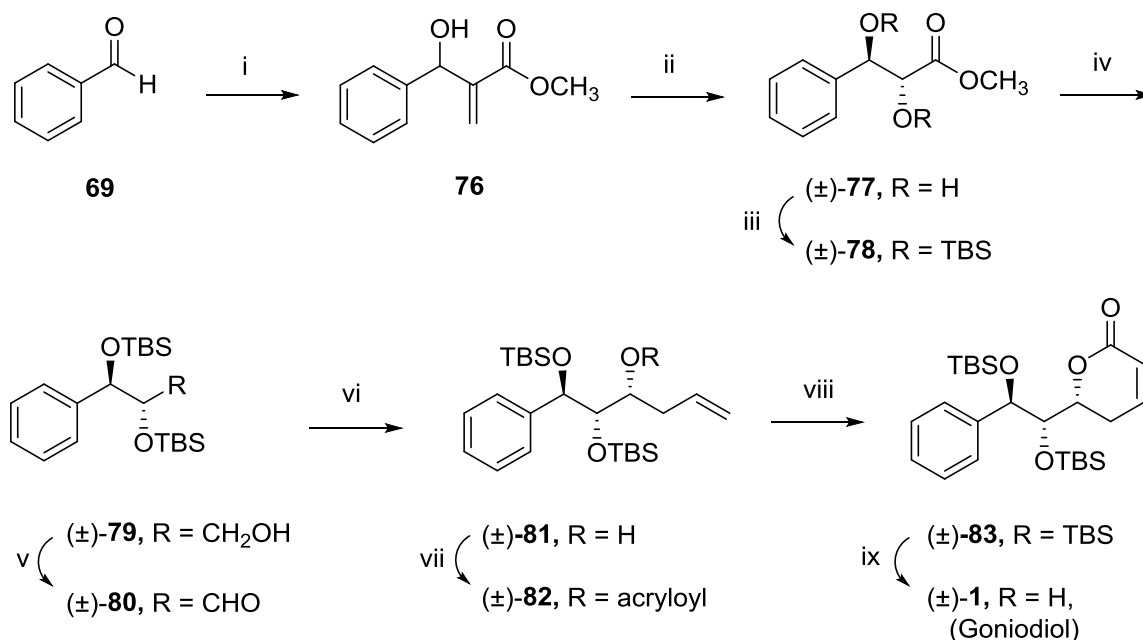
Yadav *et al.* have also developed another approach commencing from commercially available benzaldehyde **69**, which was converted to homoallylic alcohol **70** (98% ee) *via* Keck allylation using (*S*)-BINOL. The protection of alcohol **70** and subsequent oxidative cleavage provided aldehyde **72**. *L*-Proline-catalyzed α -aminoxylation of aldehyde **72** followed by *in situ* In-mediated allylation provided *syn*-alcohol **73** in 65% yield, dr = 6:4 (*syn:anti*). *syn*-Alcohol **73** was subjected to oxidative cleavage to provide aldehyde **75**, which on chain elongation with Wittig ylide and under cyclization conditions provided (+)-goniodiol (**1**) (**Scheme 11**).



Scheme 11: (i) (*S*)-BINOL, $\text{Ti}(\text{O}^i\text{Pr})_4$, CH_2Cl_2 reflux, 2 h, 25 °C, 10 min, then allyltributyltin, -78 °C, 24 h, 91%; (ii) Pr_2NEt , MOMCl, CH_2Cl_2 , 25 °C, 15 h, 92%; (iii) (a) cat. OsO_4 , NMO, acetone: H_2O (2:1), 25 °C, 24 h; (b) NaIO_4 , THF: H_2O (2:1), 85%; (iv) PhNO, *L*-proline, CHCl_3 , 0 °C, 4 h, allyl bromide, NaI, In, DMSO, 25 °C, 90 min, 65%; (v) $\text{Cu}(\text{OAc})_2$, EtOH, 12 h, 25 °C; (vi) (a) NaH, BnBr, THF, 25 °C, 12 h, 90%; (b) cat. OsO_4 , NMO, acetone: H_2O (2:1), 25 °C, 24 h; (c) NaIO_4 , THF: H_2O , 85%.

Coelho's approach (2011)¹⁷

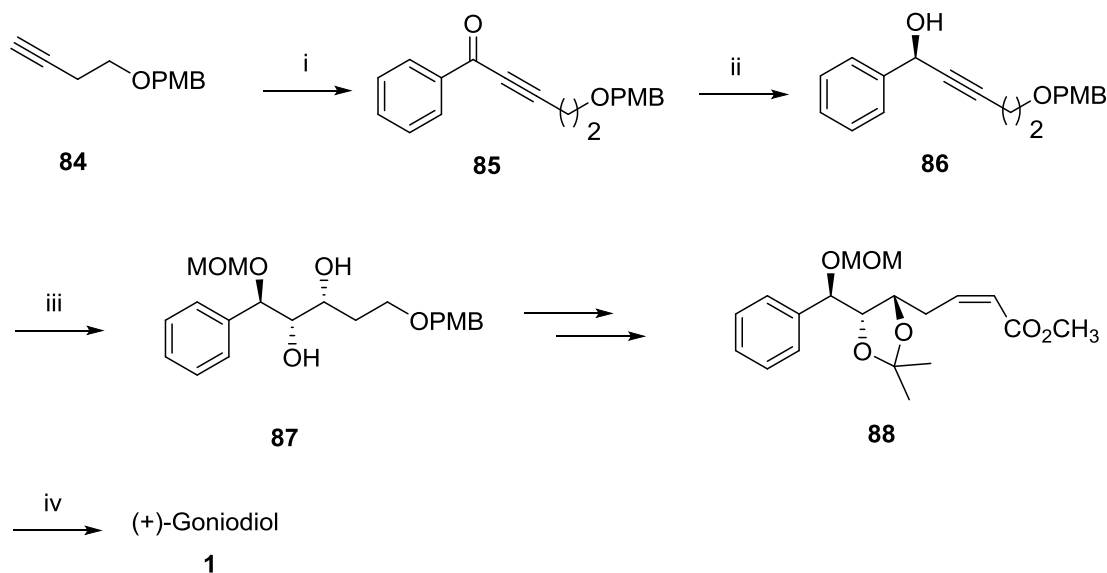
Coelho *et al.* have synthesized (±)-goniodiol (**1**) using Morita-Baylis-Hillman adduct. The reaction between benzaldehyde **69** and methyl acrylate gave the Morita-Baylis-Hillman adduct **76** in 85% yield. Ozonolysis under reductive conditions afforded *anti*-dihydroxylated ester **77** (95% de), which was protected as its silyl ether **78**. The aldehyde **80** was obtained by reduction of ester **78**, followed by the oxidation of alcohol **79** with IBX. Allylation using allyltributylstanane on aldehyde **80** gave allyl alcohol **81** (*syn:anti* = 83:17) in 79% yield, which on reaction with acryloyl chloride provided ester **82**. Ring closing metathesis of ester **82** gave α,β -unsaturated cyclohexenone **83**, which on treatment with HF/pyridine gave (±)-goniodiol (**1**) (Scheme 12).



Scheme 12: (i) methyl acrylate, $25\text{ }^\circ\text{C}$, 96 h, 85%; (ii) (a) O_3 , MeOH, $-78\text{ }^\circ\text{C}$, then $(CH_3)_2S$, 1 h; (b) $NaBH_4$, CH_2Cl_2 , $-78\text{ }^\circ\text{C}$, 12 h, 70%; (iii) TBSCl, imid., $25\text{ }^\circ\text{C}$, 12 h, 98%; (iv) DIBAL-H, CH_2Cl_2 , $-78\text{ }^\circ\text{C}$, 1 h, 79%; (v) IBX, DMSO, $25\text{ }^\circ\text{C}$, 1 h, 88%; (vi) allyltributyltin, $TiCl_4$, CH_2Cl_2 , $-78\text{ }^\circ\text{C}$, 1 h, 79%; (vii) acryloyl chloride, NEt_3 , CH_2Cl_2 , $0\text{ }^\circ\text{C}$, 83%; (viii) cat. Grubbs' I generation, CH_2Cl_2 reflux, 18 h, 64%; (ix) HF, pyridine, CH_3CN , $25\text{ }^\circ\text{C}$, 24 h, 74%.

Sabitha's approach (2011)¹⁸

Sabitha *et al.* have developed a useful synthetic method for (+)-goniodiol (**1**) using CBS-reduction and Sharpless asymmetric dihydroxylation as the key reactions. Thus, asymmetric CBS-reduction of ketone **85** derived from homopropargylic alcohol derivative **84**, gave chiral propargylic alcohol **86** (99% ee) in 66% yield. Reduction of propargylic alcohol **86** with $LiAlH_4$ followed by Sharpless asymmetric dihydroxylation with AD-mix- β provided diol **87** as a single isomer in 90% yield. Ester **88** was obtained from diol **87** in four steps, where Still-Gennari reagent was used for chain elongation. Finally, the ester **88** on treatment with catalytic amount of *p*-TSA afforded (+)-goniodiol (**1**) via tandem deprotection and *in situ* cyclization process (Scheme 13).



Scheme 13: (i) (a) LHMDS, benzaldehyde, THF, -78 °C to 25 °C, 3 h, 90%; (b) IBX, DMSO, CH₂Cl₂, 0 °C to 25 °C, 92%; (ii) (*R*)-(Me)-CBS, BH₃.SMe₂, THF, -25 °C, 1 h, 66%; (iii) (a) LiAlH₄, THF, 0 °C to 25 °C, 3 h, 90%; (b) MOMCl, ^tPr₂NEt, CH₂Cl₂, 0 °C to 25 °C, 3 h, 96%; (c) AD-mix-β, CH₃SO₂NH₂, *tert*-BuOH:H₂O (1:1), 0 °C, 10 h, 90%; (iv) *p*-TsOH, EtOH, 50 °C, 16 h, 84%.

1.1.4 Present Work:

1.1.4.1 Objective:

Even though several methods are reported for the synthesis of (+)-goniodiol (**1**),⁷⁻¹⁸ some of them suffer from certain limitations such as use of chiral building blocks, exotic reagents, involvement of longer reaction sequences, expensive and hazardous reagents, low overall yields, low diastereomeric ratios, etc. In this context, a more practical method for the synthesis of (+)-goniodiol (**1**) is highly desirable.

Retrosynthetic analysis of (+)-goniodiol (**1**) reveals that homoallylic alcohol **103** could be visualized as the key intermediate. The homoallylic alcohol **103** could be achieved by means of Lewis acid-mediated diastereoselective allylation of aldehyde obtained from *anti*-methoxy diol **99**. The *anti*-methoxy diol **99** could in turn be obtained by performing Co-catalyzed two-stereocentered hydrolytic kinetic resolution

of the corresponding racemic *anti*-methoxy epoxide **97**. The requisite racemic *anti*-methoxy epoxide **97** could be easily prepared from cinnamyl alcohol **62** (Fig. 2).

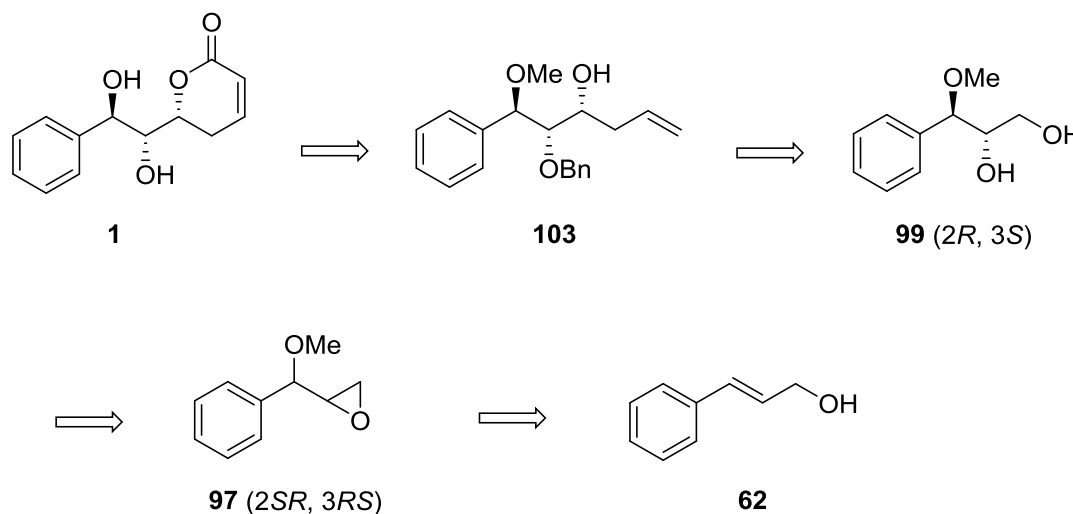


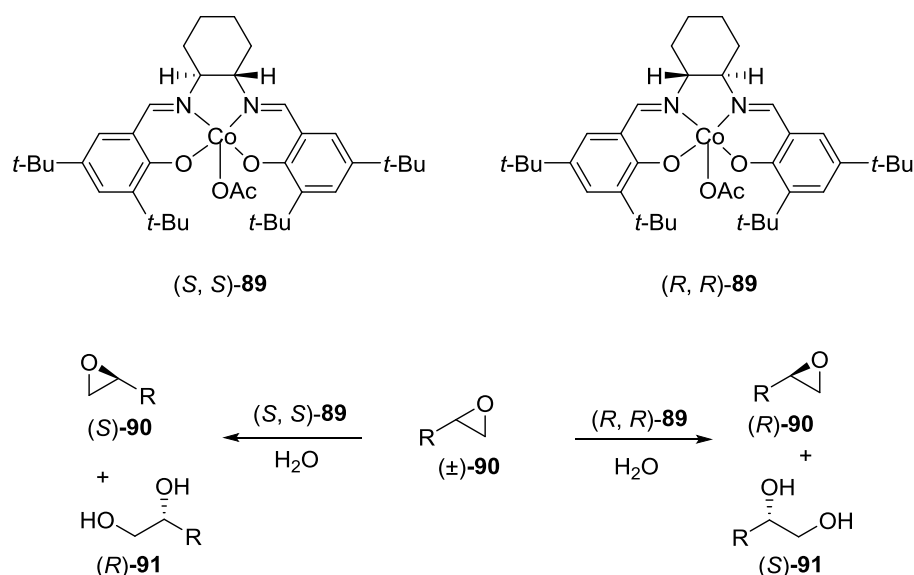
Fig. 2: Retrosynthetic analysis of (+)-goniodiol (**1**)

Since this chapter employs Co-catalyzed two-stereocentered HKR (Hydrolytic Kinetic Resolution),²⁰ as a key chiral inducing step, a brief account of the same is presented in the following section.

1.1.4.2 Hydrolytic Kinetic Resolution (HKR)

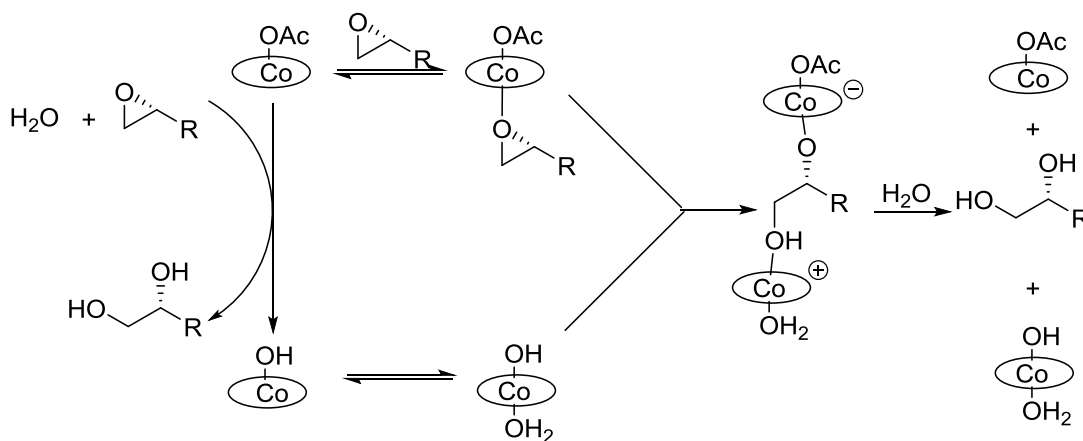
The importance of epoxides in organic synthesis arises partly from the occurrence of the strained three-membered ring unit in a number of interesting natural products²¹ but more so because the ring opening of epoxides allows straightforward elaboration to useful new functionality, often with generation of new carbon-carbon bonds. Indeed, reactions of epoxides with nucleophiles, Lewis acids, radicals, reducing agents, oxidizing agents, acids and bases have all been well-documented and utilized in synthesis.²² Thus, epoxides are versatile building blocks for organic synthesis. However, terminal epoxides are arguably the most important subclass of these

compounds, and no general and practical method exists for their production in enantiomerically pure form. Terminal epoxides are available very inexpensively as racemic mixtures, and kinetic resolution is an attractive strategy for the production of optically active epoxides, given an economical and operationally simple method. Readily accessible synthetic catalysts (for example: chiral cobalt-salen complexes, **89**)²³ have been used for the efficient asymmetric hydrolysis of terminal epoxides. This process uses water as the only reagent, no added solvent, and low loadings of a recyclable catalyst (0.5 mol%), and it affords highly valuable terminal epoxides and 1,2-diols in high yields with high enantiomeric enrichment. One of the most attractive features of kinetic resolution processes in general is the fact that the enantiomeric composition of unreacted substrate can be controlled by adjusting the degree of conversion, and virtually enantiopure material can be obtained at appropriately high conversions. However, in the absence of straightforward substrate racemization protocols, kinetic resolutions have the significant disadvantage of a 50% maximum yield of substrate recovery. With a specific interest in devising a practical method for obtaining highly enantioenriched terminal epoxides, the following criteria must be met in order for a kinetic resolution approach to be viable:²⁴ (1) The racemic epoxides must be inexpensive or easily accessible from commercial starting materials; (2) The catalyst for the resolution must be readily available in both enantiomeric forms. The catalyst would be used in small quantities in the resolution and would be recyclable; (3) The nucleophile used for the ring opening should be inexpensive and easily handled; (4) The resolved epoxides must be obtained in good yield and very high enantiopurity and must be easily separated from the ring-opened products; (5) Ideally, the ring-opened byproducts should also be valuable chiral building blocks and be obtainable in high enantiomeric excess.



Scheme 14: Jacobsen's Hydrolytic Kinetic Resolution (HKR) of racemic epoxide, $(\pm)\text{-90}$

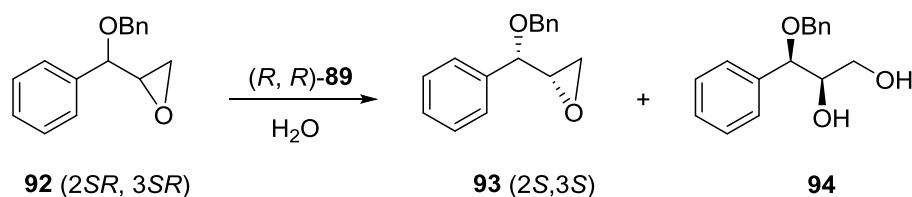
The (salen)Co complex **89** has been well-established to catalyze the efficient hydrolytic kinetic resolution (HKR) of a variety of terminal epoxides **90** (**Scheme 14**).²⁵ One of the most remarkable features of this method is the consistently high stereoselectivity obtained in the hydrolysis of a wide range of terminal epoxides, with the relative rate of reaction for the two enantiomers of the substrate (k_{rel}) >500 for some substrates and >100 for almost all examined.²⁶ The asymmetric induction in epoxide opening is imparted by both chiral complexes working cooperatively rather than by either complex alone. A mechanistic picture have been disclosed wherein the rate and stereoselectivity-determining step involves one Co(III) complex acting as a Lewis acid to activate the epoxide while another serves to activate water as a nucleophile *via* a (salen)Co–OH complex (**Scheme 15**).²⁷ The rate of this step, and therefore of the overall reaction, depends strongly on the identity of the counterion in the (salen)Co–OAc precatalyst.²⁸ In contrast, the stereoselectivity in the HKR was shown to be quite insensitive to counterion effects.^{28a}



Scheme 15: Proposed catalytic mechanism for epoxide hydrolysis

This method appeared to hold considerable promise with regard to meeting all of the five criteria outlined above. First, racemic 1,2-epoxides are generally available directly from commercial suppliers at low cost or are obtainable in one-step from inexpensive olefins or aldehydes. In fact, certain racemic epoxides, such as propylene oxide, epichlorohydrin, styrene oxide, and butadiene monoepoxide, are commodity chemicals and are no more expensive than common organic solvents. Second, the ligands for catalyst **89** had previously been commercialized and manufactured on a ton scale in the context of (salen)Mn epoxidation catalysts.²⁹ The cobalt analogues (*R,R*)-**89** and (*S,S*)-**89** proved equally accessible, and these are also now available in bulk. Third, water is perhaps the ideal reagent for effecting the resolution reaction: it is inexpensive and safe, and the rate of the ring-opening reaction can be controlled simply by modulating the rate of addition of water to the epoxide-catalyst mixture. Fourth, for those examples that were described in the preliminary report, highly enantioenriched epoxides were recovered from the HKR. Finally, the HKR provided useful enantioenriched 1,2-diols, including many that are otherwise not readily accessible using existing asymmetric dihydroxylation methods.³⁰ Two useful methods for the generation of complex **89** have been developed.

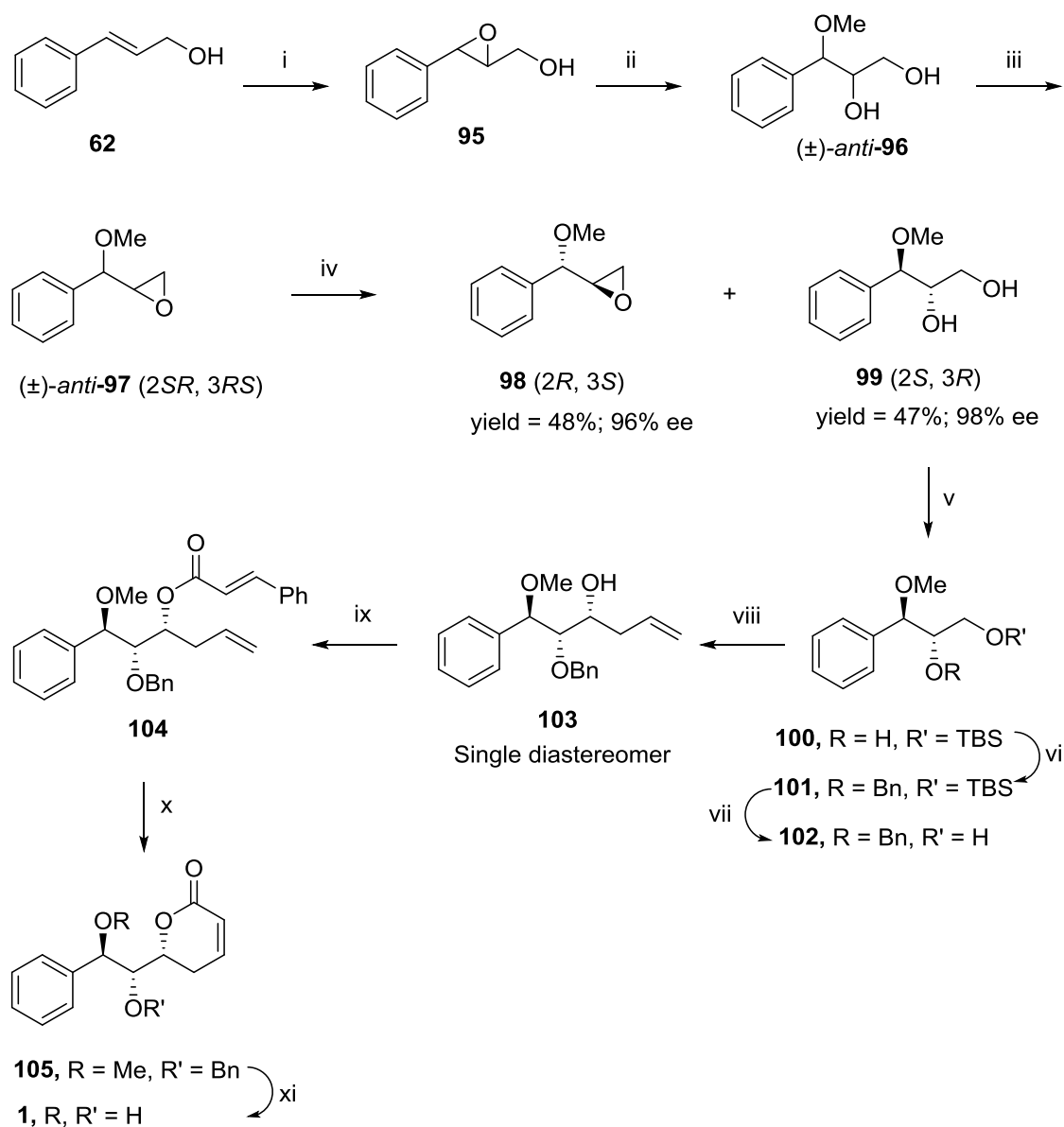
Method A involves isolation of **89** as a crude solid prior to the HKR. The Co(II) salen complex is dissolved in toluene to generate ~ 1 M solution, and acetic acid (2 equiv) is added. The resulting solution is stirred open to air at room temperature for 30 min. All volatile materials are removed *in vacuo*, affording **89** as a brown solid residue that can be used without further purification. Method B involves *in situ* generation of **89** by suspension of the Co(II) salen complex in epoxide and addition of HOAc under an aerobic atmosphere. Despite these achievements, HKR has only been applied to the resolution of simple terminal epoxides with one stereocentre.³¹ To overcome this drawback, Sudalai *et al.* have extended the scope of the applicable substrates to cover multi-functionalized molecules with two stereocentres. This study related to HKR of alkoxy epoxides (*ex*: benzyloxy epoxide **92**) with two stereocentres, wherein the relative stereochemistry between the alkoxy and the epoxide functions are established prior to the HKR step and thus a single asymmetric reaction is employed to form compounds with two asymmetric centers (**Scheme 16**).^{20a}



Scheme 16: Hydrolytic Kinetic Resolution (HKR) of two-stereocentered benzyloxy epoxide, **92**

1.1.5 Results and Discussion

The synthetic scheme for (+)-goniodiol (**1**), wherein two-stereocentred Co-catalyzed HKR of racemic epoxide and Lewis acid-mediated diastereoselective allylation of aldehyde constituting two key steps for the introduction of chirality into the molecule is shown in **Scheme 17**.



Scheme 17: (i) *m*CPBA, CH₂Cl₂, 14 h, 0 °C, 88%; (ii) camphor sulfonic acid (10 mol %), MeOH, 1 h, 96%; (iii) (a) TsCl, Bu₂SnO (2 mol%), Et₃N, DMAP (10 mol%), CH₂Cl₂, 2 h; (b) K₂CO₃, MeOH, 3 h, 76% (over two steps); (iv) (*S,S*)-Co(salen)OAc, (0.5 mol%), THF, H₂O (0.5 equiv), 0 °C, 14 h; (v) TBDMSCl, imid., CH₂Cl₂, 0 °C to 25 °C, 1 h, 90%; (vi) NaH, DMF, BnBr, 0 °C to 25 °C, 2 h, 95%; (vii) TBAF, THF, 25 °C, 8 h, 91%; (viii) IBX, DMSO, 2 h, 25 °C then TiCl₄, tributyl allyl stannane, CH₂Cl₂, -78 °C, 30 min, 73% (over two steps); (ix) (*E*)-cinnamoyl chloride, dry pyridine, DMAP, CH₂Cl₂, 0 °C to 25 °C, 12 h, 64%; (x) Grubbs' II generation catalyst (10 mol%), CH₂Cl₂ reflux, 8 h, 76%; (xi) BBr₃, CH₂Cl₂, -78 °C, 6 h, 71%.

The present synthesis of (+)-goniodiol (**1**) started from commercially available cinnamyl alcohol **62**, which on epoxidation with *m*CPBA gave the epoxy alcohol **95** in 88% yield. The formation of epoxy alcohol **95** was confirmed from its ¹H NMR spectrum, which showed a doublet at δ 3.92 (d, *J* = 2.2 Hz, 1H) and a multiplet at δ 3.19-3.23 (m, 1H) corresponding to two methine protons of epoxide moiety. Its ¹³C NMR spectrum showed two typical carbon signals at δ 55.6 and 62.5 indicative of the methine carbons of epoxide, thus confirming the formation of epoxy alcohol **95** (Fig. 3).

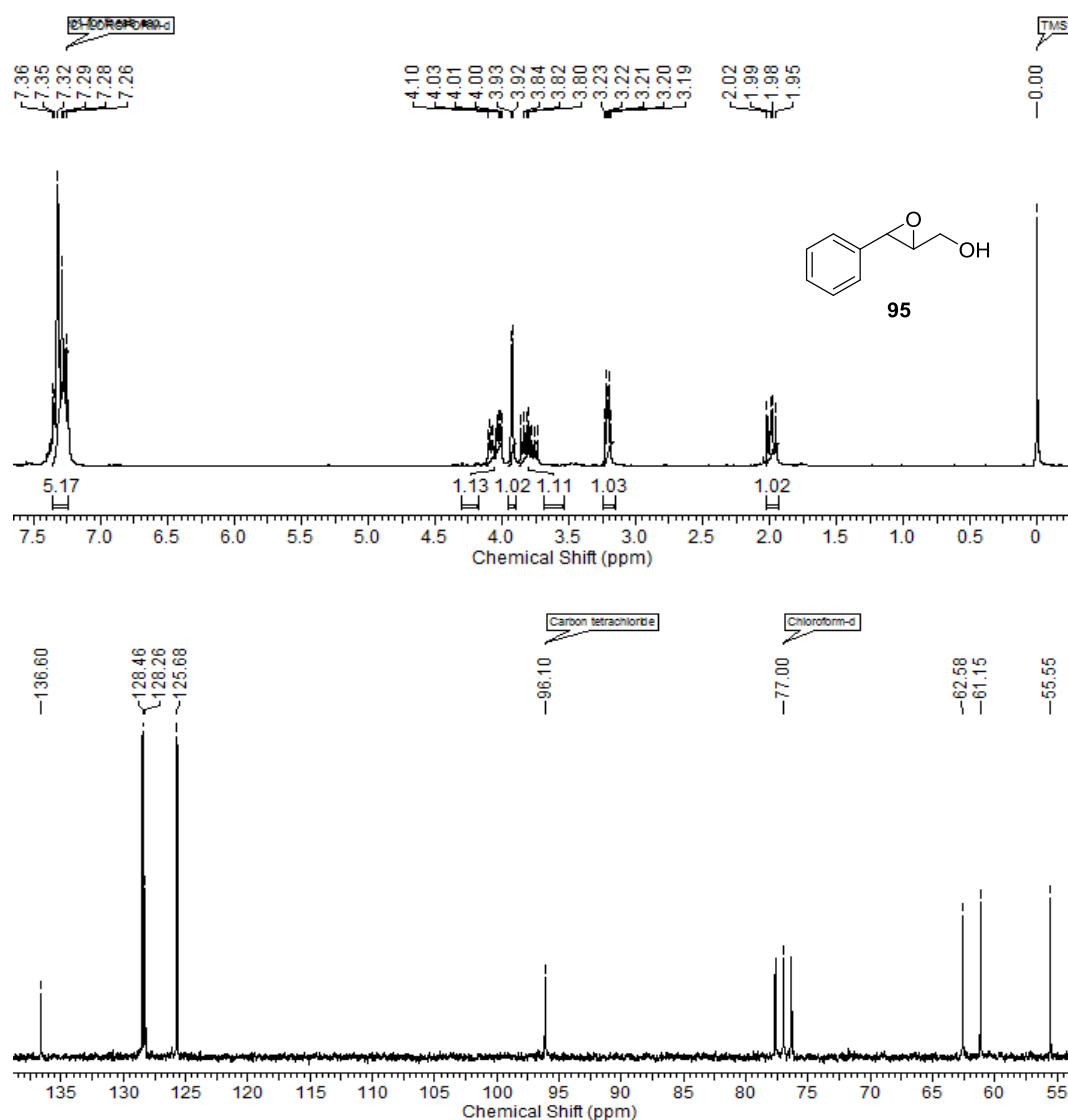


Fig. 3: ¹H and ¹³C NMR spectra of epoxy alcohol **95**

The regiospecific opening of epoxide **95** with MeOH was accomplished quantitatively to give the racemic *anti*-methoxy diol **96**. Its ^1H NMR spectrum showed a characteristic multiplet at δ 3.63-3.75 (m, 3H) for methylene ($-\text{CH}_2-\text{OH}$) protons while its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 62.2 and 75.7 due to methylene and methine carbons attached to hydroxyl groups respectively (**Fig. 4**).

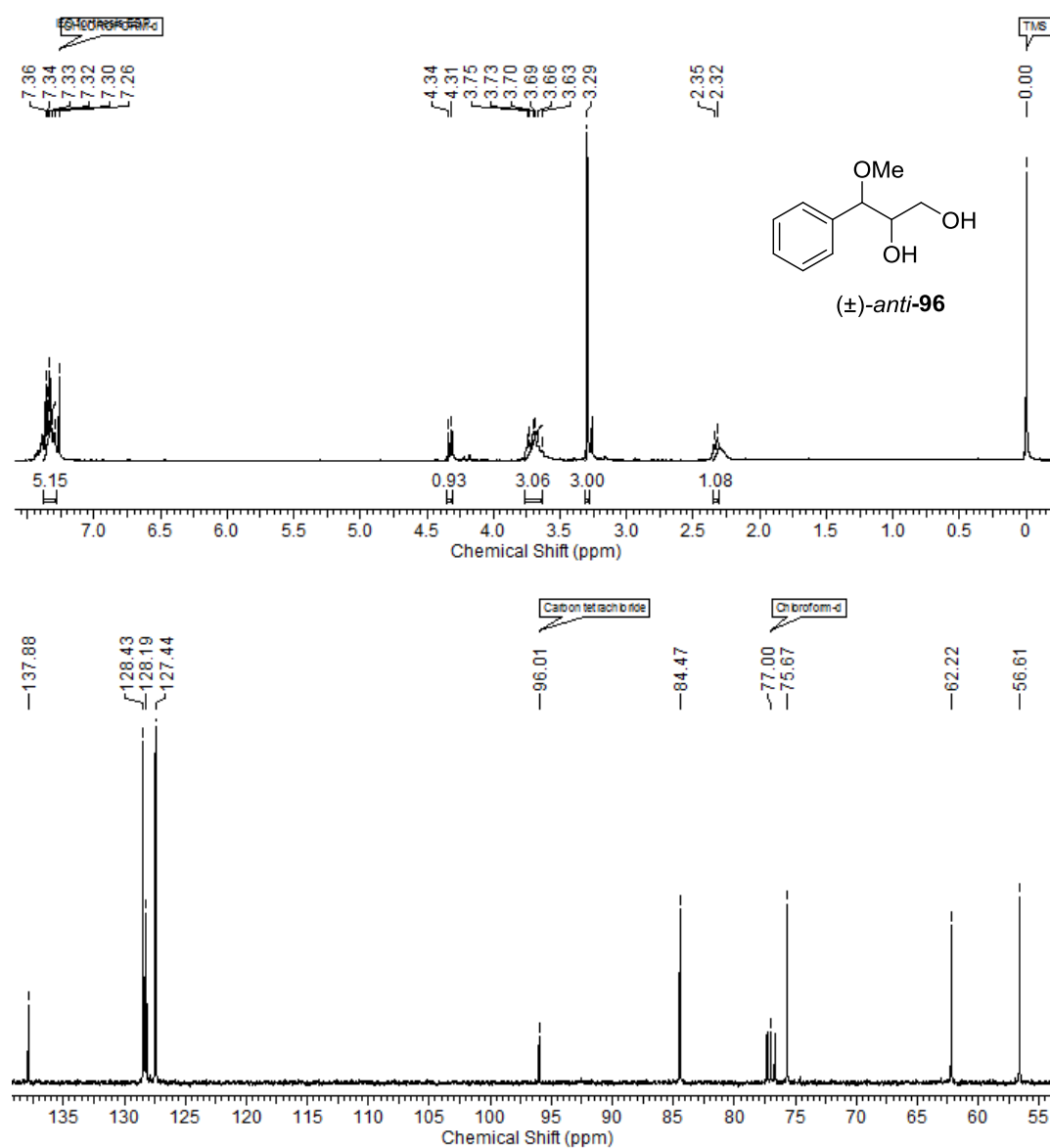


Fig. 4: ^1H and ^{13}C NMR spectra of racemic *anti*-methoxy diol **96**

Racemic *anti*-methoxy diol **96** was further converted to racemic *anti*-methoxy epoxide **97** in a two-step reaction sequence: i) selective monotosylation of primary alcohol (TsCl, cat. Bu₂SnO, Et₃N, DMAP, CH₂Cl₂); ii) formation of epoxide under basic conditions (K₂CO₃, MeOH). The formation of racemic *anti*-methoxy epoxide **97** was confirmed from its ¹H NMR spectrum, which showed typical proton signals at δ 2.59 (dd, *J* = 2.6, 4.8 Hz, 1H), 2.73 (t, *J* = 4.2 Hz, 1H) and 3.14-3.21 (m, 1H) corresponding to methylene and methine protons of epoxide respectively. Its ¹³C NMR spectrum showed characteristic carbon signals at δ 43.8 and 55.0 corresponding to methylene and methine carbons of the epoxide moiety respectively (**Fig. 5**).

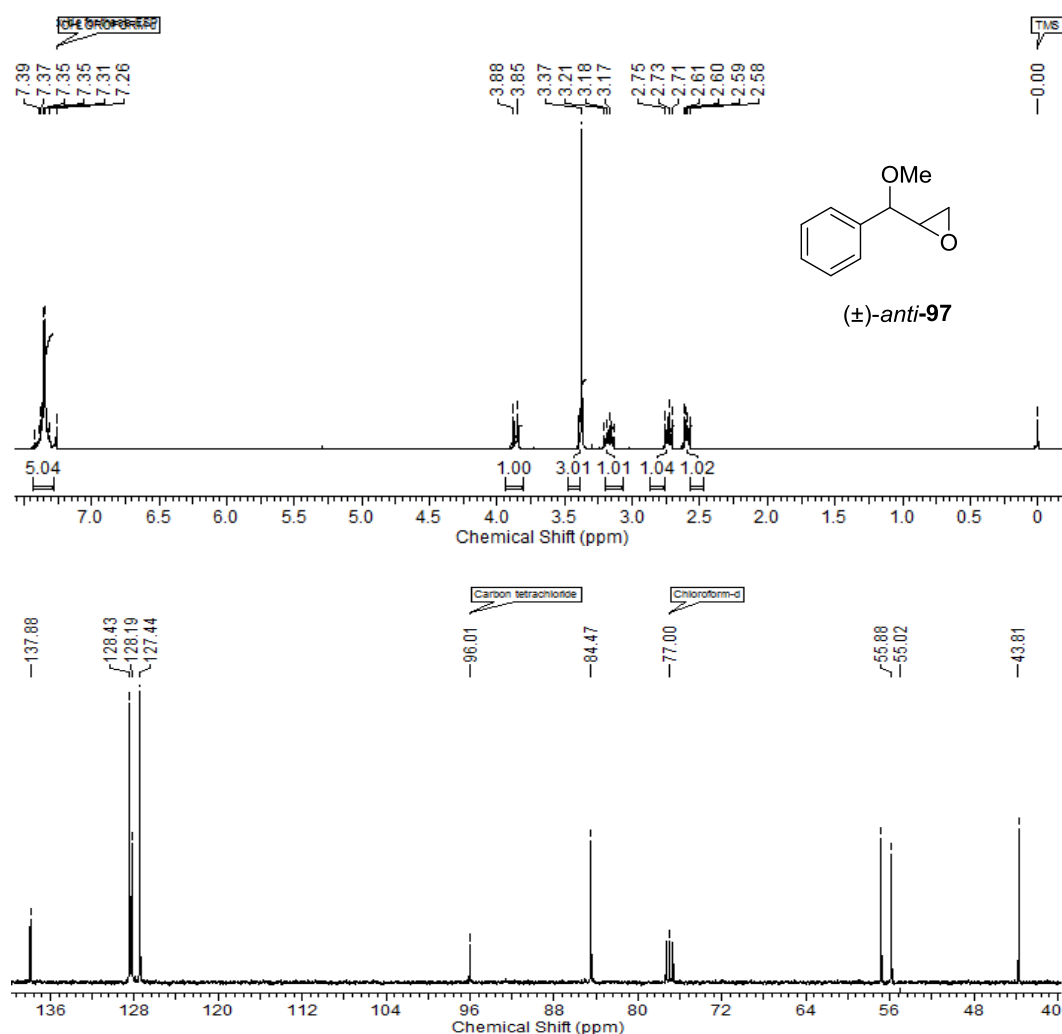


Fig. 5: ¹H and ¹³C NMR spectra of racemic *anti*-methoxy epoxide **97**

Then, *anti*-(2*SR*, 3*RS*)-3-methoxy-3-phenyl-1,2-epoxypropane **97** was subjected to HKR with (*S*, *S*)-salen Co(OAc) complex^{20a,b} (0.5 mol%) and H₂O (0.5 equiv), which produced the corresponding chiral epoxide **98** (48% yield, 96% ee) and chiral *anti*-diol **99** (47% yield, 98% ee) in high optical purity.

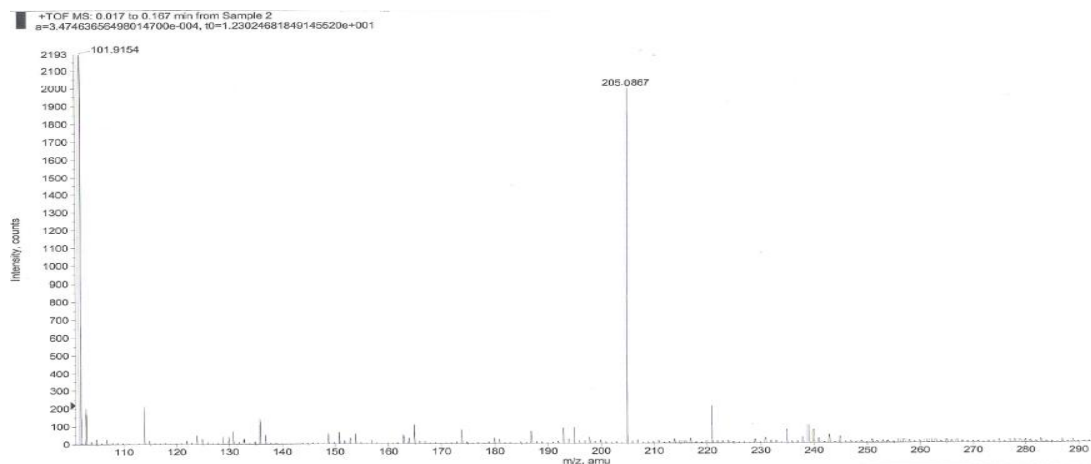
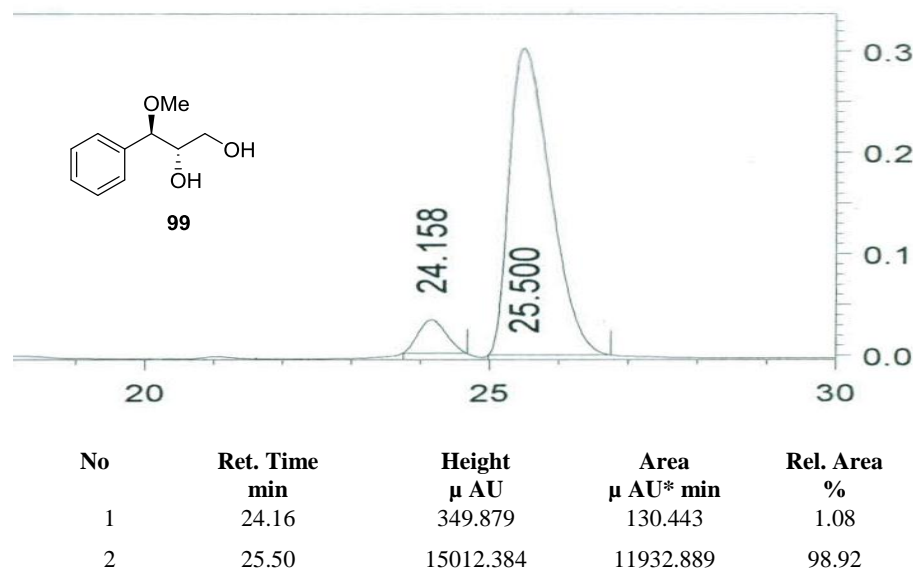


Fig. 6: HPLC chromatogram and mass spectra of chiral *anti*-methoxy diol **99**

The chiral diol **99** was readily separated from epoxide **98** by the column chromatographic purification. The optical purity of chiral diol **99** was determined to be 98% ee from chiral HPLC analysis (Chiralcel OJ-H, *n*-hexane/ *i*PrOH, 86:14, 0.5

mL/min) retention time 24.16 (1.08%) and 25.50 (98.92%). The mass spectrum also confirmed the formation of diol **99** (Fig.6).

The primary hydroxyl function in diol **99** was selectively protected as TBS ether (TBSCl, imid.)³² to give the corresponding silyl ether **100** in 90% yield. The formation of silyl ether **100** was confirmed from its ¹H NMR spectrum, which showed a doublet at δ 0.07 (d, $J = 2.5$ Hz, 6H) corresponding to methyl protons attached to silicon atom.

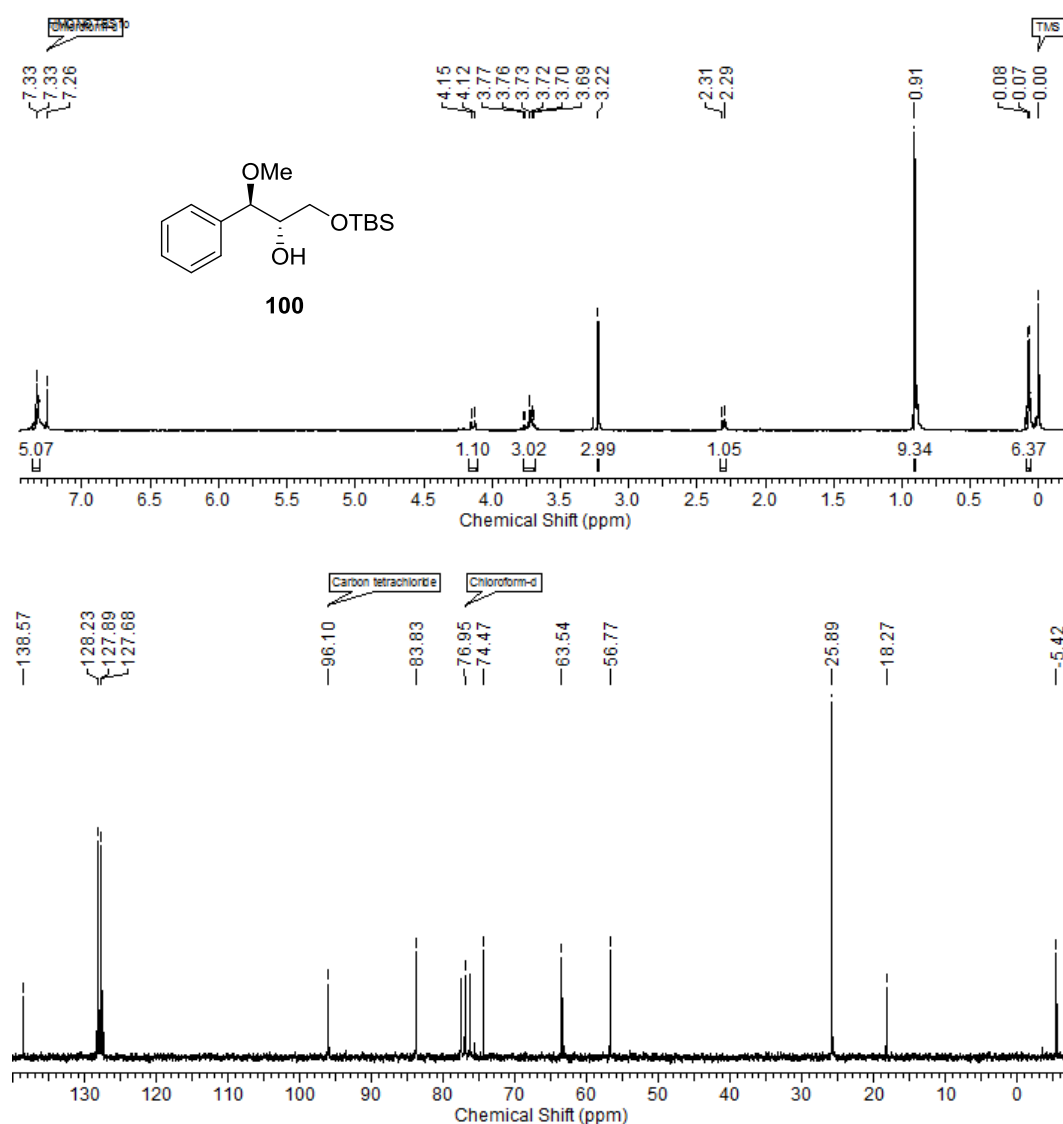


Fig. 7: ¹H and ¹³C NMR spectra of silyl ether **100**

Its ^{13}C NMR spectrum showed characteristic carbon signals at δ -5.4 and 25.9 corresponding to methyl and quaternary carbons of silyl group respectively (**Fig. 7**).

Protection of secondary alcohol in **100** as benzyl ether (BnBr, NaH) was then carried out to produce the protected ether **101**. Formation of **101** was confirmed by its ^1H NMR spectrum, which showed a characteristic singlet at δ 4.43 due to benzylic proton while its ^{13}C NMR spectrum showed a typical carbon signal at δ 73.7 for benzylic carbon (**Fig. 8**).

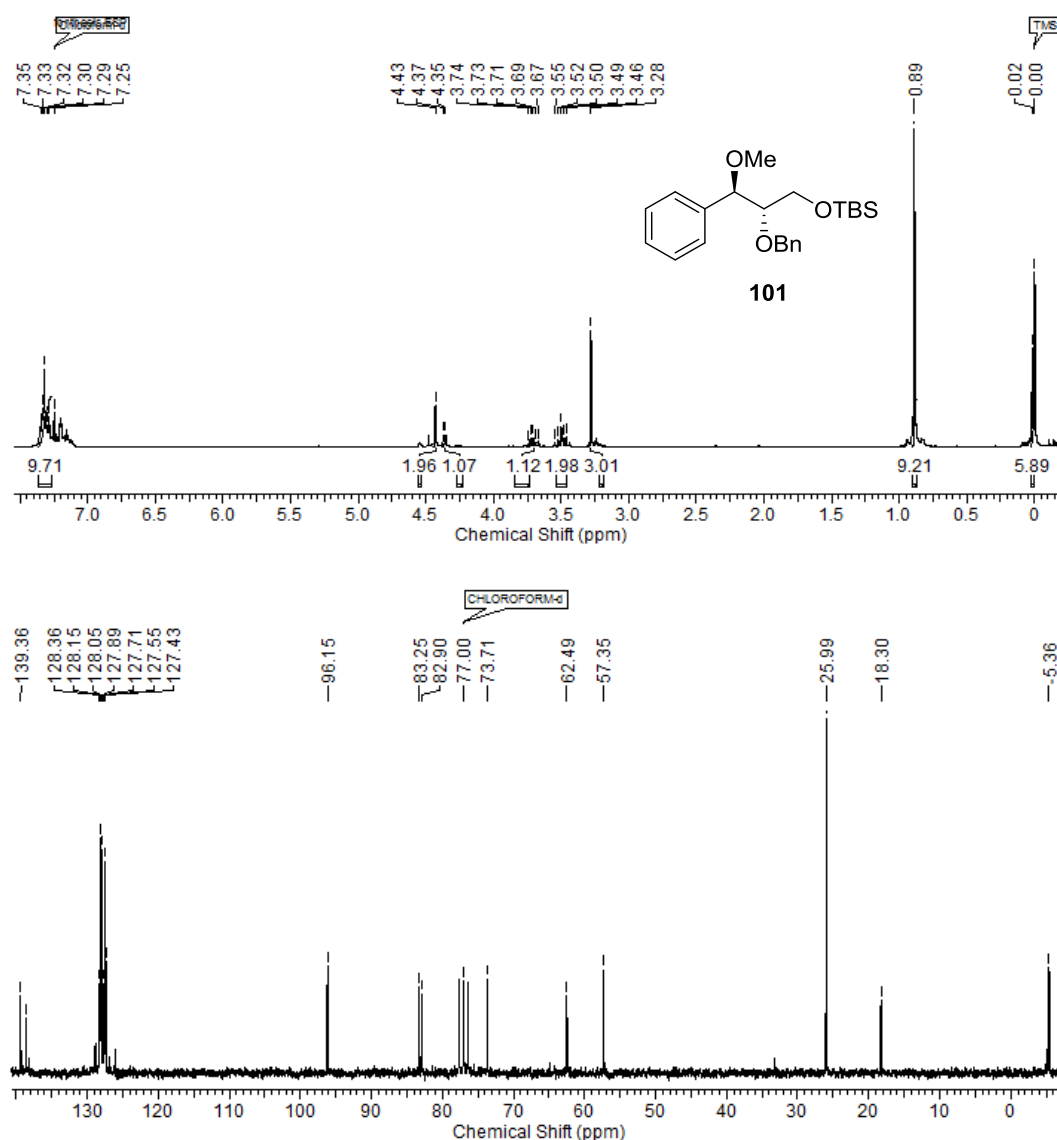


Fig. 8: ^1H and ^{13}C NMR spectra of benzyl ether **101**

The selective deprotection of TBS group was achieved using tetrabutylammonium fluoride to give the primary alcohol **102** in 91% yield. The disappearance of proton signals at δ 0.9 and 0.02 in the ^1H NMR spectrum of **102** due to the TBS group confirmed the deprotection of silyl ether (**Fig. 9**).

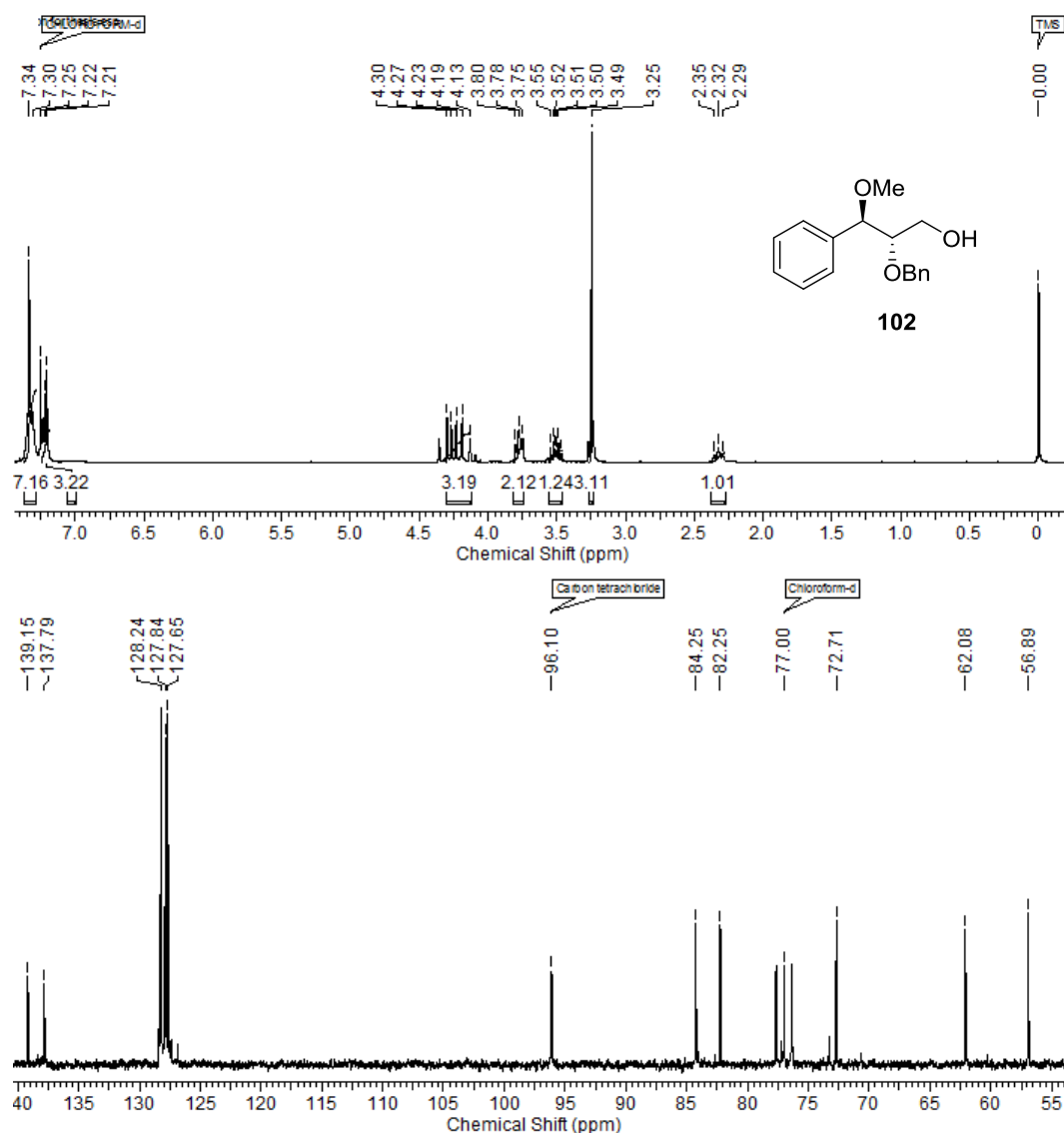


Fig. 9: ^1H and ^{13}C NMR spectra of alcohol **102**

The smooth IBX oxidation of alcohol **102** at 25 °C gave the corresponding crude aldehyde, which was found to be quite unstable on exposure both to moisture as well as air. Hence, the crude aldehyde was immediately subjected to Ti-mediated

diastereoselective allylation (TiCl₄, tri-*n*-butyl allyl stannane, CH₂Cl₂, -78 °C) to furnish the key intermediate homoallylic alcohol **103** in 73% yield over two steps as a single diastereomer as determined from its ¹H NMR spectrum of the crude product.³³

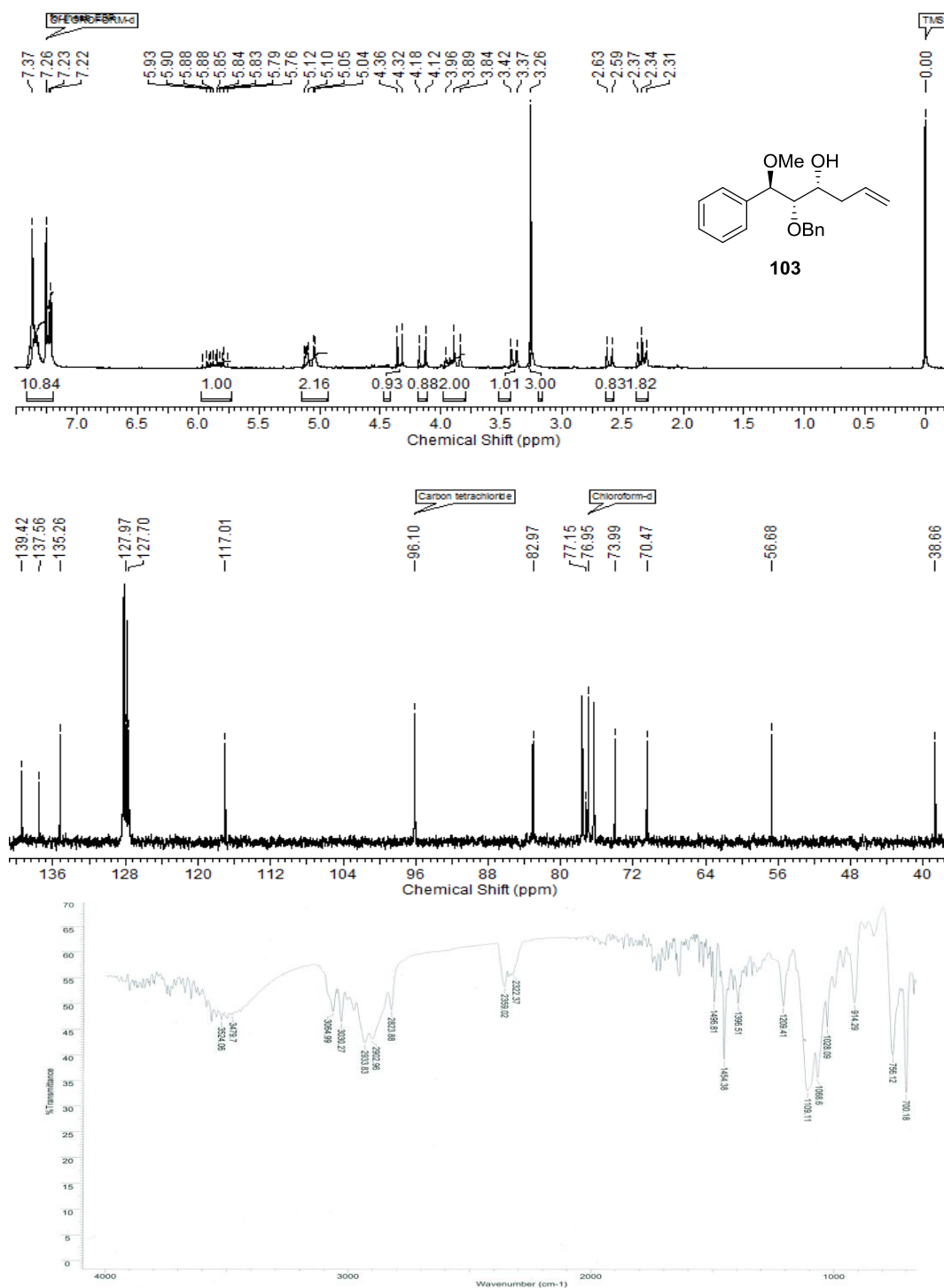


Fig. 10: ¹H, ¹³C NMR and IR spectra of homoallylic alcohol **103**

Its ^1H NMR spectrum showed two typical multiplets at δ 5.03-5.10 (m, 2H) and 5.76-5.93 (m, 1H) corresponding to olefinic protons. It was further substantiated from its ^{13}C NMR spectrum, by the appearance of characteristic olefinic carbon signals at δ 117.0 and 135.3 confirming the formation of homoallylic alcohol **103**. Its IR spectrum displayed a characteristic broad absorption band at 3479 cm^{-1} indicating the presence of OH functional group (**Fig. 10**).

Esterification of alcohol **103** with (*E*)-cinnamoyl chloride in the presence of pyridine and catalytic amount of DMAP in CH_2Cl_2 at $25\text{ }^\circ\text{C}$ gave the corresponding ester **104** in 64% yield.¹¹

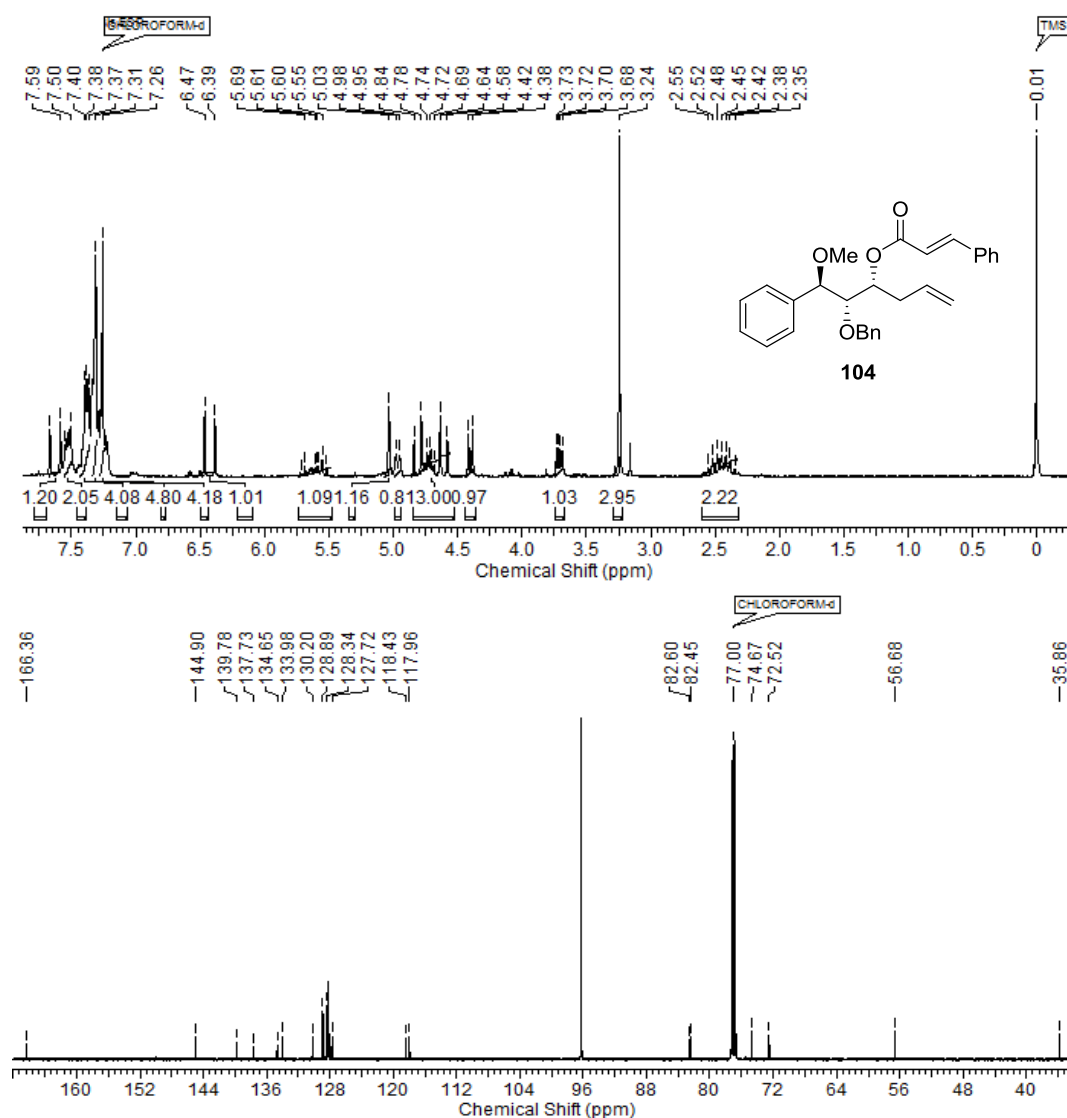


Fig. 11: ^1H and ^{13}C NMR of ester **104**

The ^1H NMR spectrum of **104** showed two doublets at δ 6.43 (d, $J = 16.0$ Hz, 1H) and 7.63 (d, $J = 16.0$ Hz, 1H) establishing the presence of α - and β - CH proton of COCH=CHPh . This was further ascertained by the presence of carbon signals at δ 117.9 and 144.9 corresponding to α - and β - carbon respectively of COCH=CHPh in its ^{13}C NMR spectrum. Its IR spectrum displayed a strong absorption band at 1714 cm^{-1} indicating the presence of ester carbonyl group (**Fig. 11**).

Ring closing metathesis of cinnamate ester **104** with Grubbs' second generation catalyst led to the isolation of α -pyrone **105** in 76% yield.

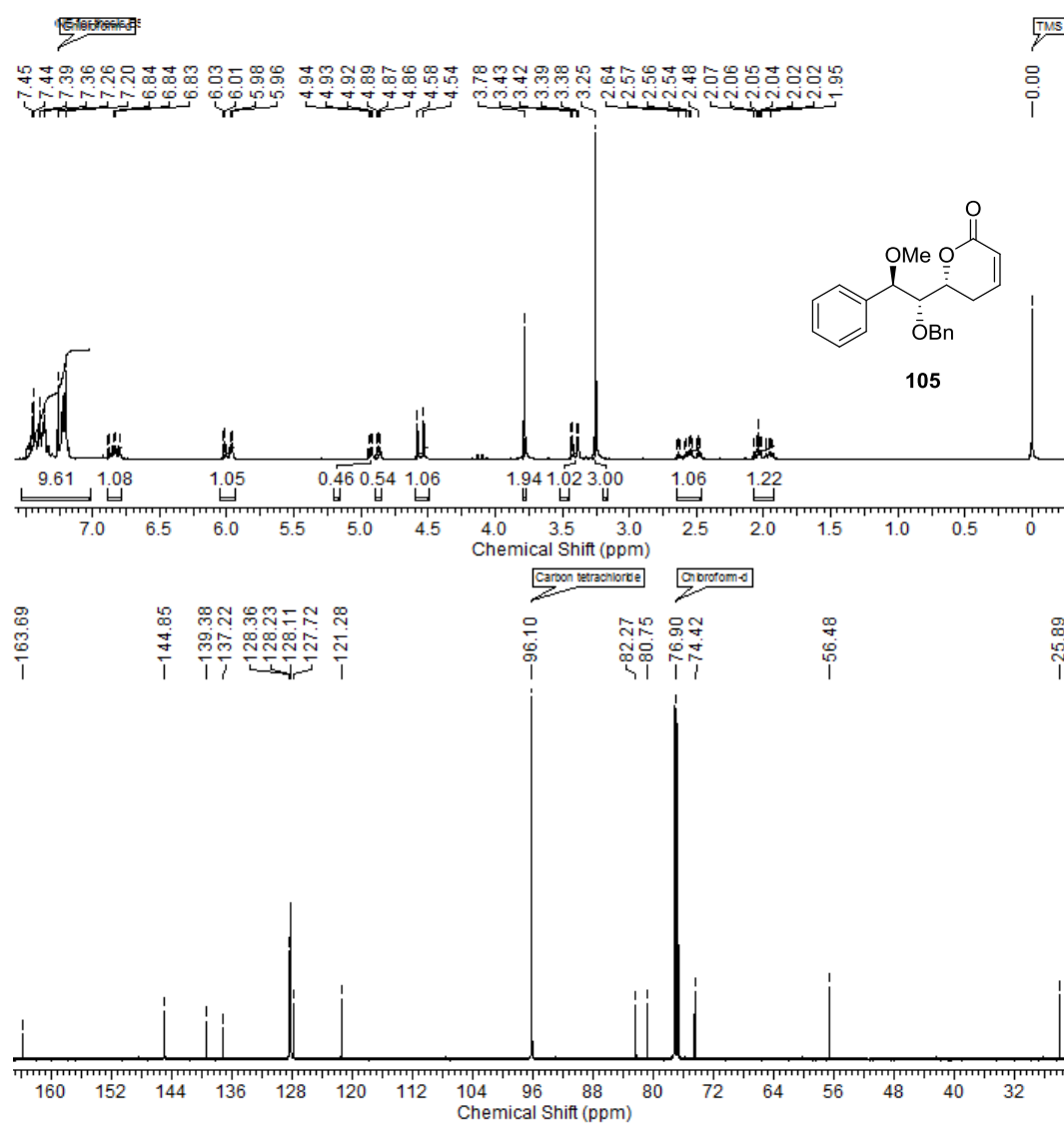


Fig. 12: ^1H and ^{13}C NMR spectra of α -pyrone **105**

The ^1H NMR spectrum of **105** showed two typical multiplets at δ 5.99 and 6.84 due to olefinic protons. Its ^{13}C NMR spectrum displayed a typical carbon signal at δ 163.7 due to lactone carbonyl carbon, confirming the formation of α -pyrone **105** (Fig. 12). Its IR spectrum showed a strong absorption band at 1720 cm^{-1} due to carbonyl of lactone moiety. Finally, simultaneous demethylation and debenzylation of **105** were achieved successfully with BBr_3 (CH_2Cl_2 , $-78\text{ }^\circ\text{C}$, 6 h) to afford (+)-goniodiol (**1**) in 71% yield.³⁴

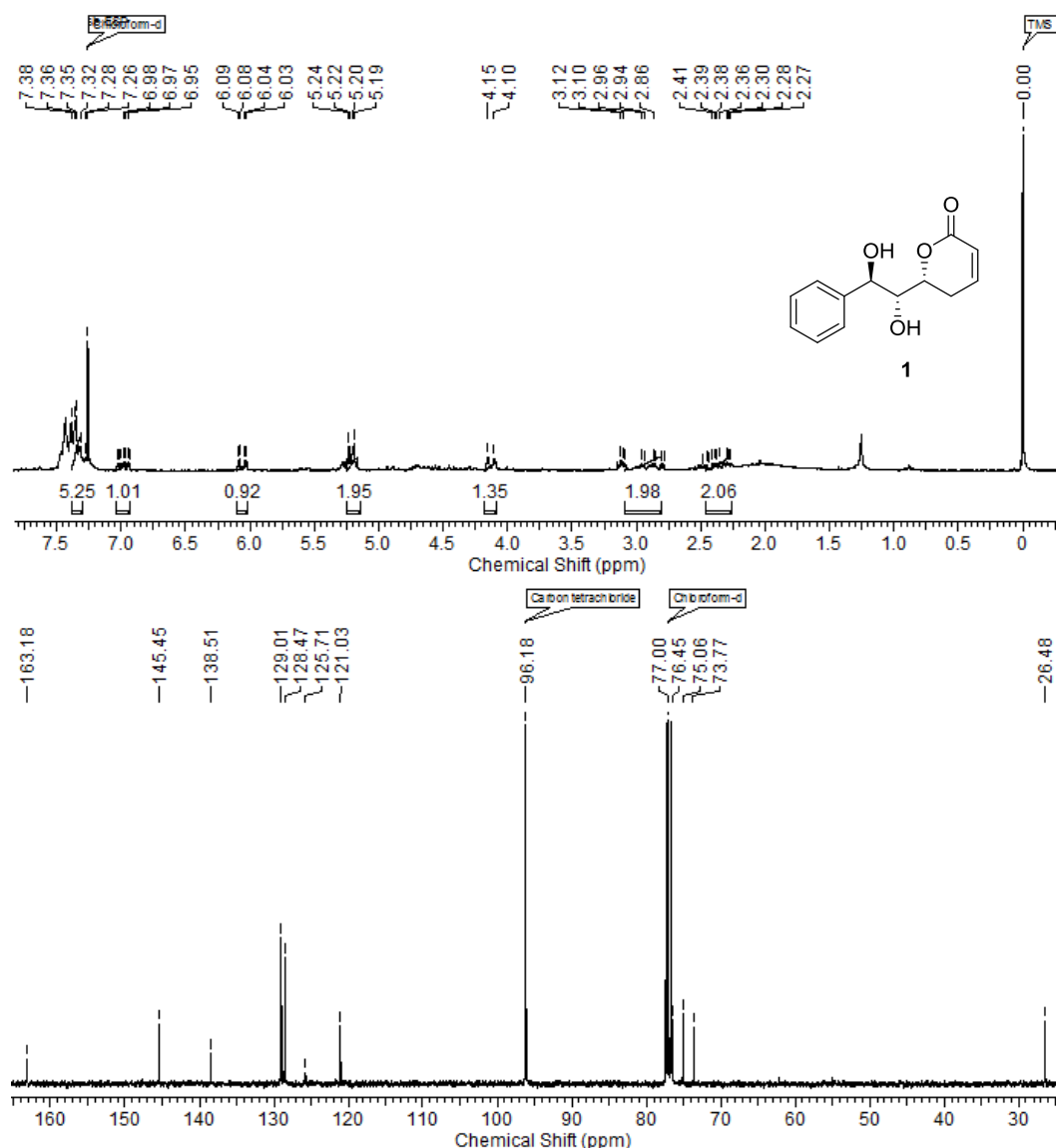


Fig. 13: ^1H and ^{13}C NMR spectra of (+)-goniodiol (**1**)

The ^1H NMR spectrum of (+)-goniodiol (**1**) showed typical proton signals at δ 6.06 (dd, $J = 3.6, 7.2$ Hz, 1H) and 6.98 (ddd, $J = 2.9, 6.4, 8.1$ Hz, 1H), accounting for two olefinic protons. The other signal at δ 2.27-2.40 (m, 2H) was attributed to methylene protons $\text{CH}_2\text{CH}=\text{CH}$. Its ^{13}C NMR spectrum showed characteristic carbonyl resonance at δ 163.2, while two olefinic carbons displayed signals at δ 121.0 and 145.5 (**Fig. 13**). The ee of (+)-goniodiol (**1**) was found to be 98% based on comparison of its optical rotation with the reported value $\{[\alpha]_{25}^{\text{D}} + 73.77$ (c 0.24, CHCl_3); lit.⁶ $[\alpha]_{25}^{\text{D}} + 74.4$ (c 0.3, CHCl_3)}. The spectral data of (+)-goniodiol (**1**) were in complete agreement with the reported values.^{6, 11}

1.1.6 Conclusion

In conclusion, an efficient and straight-forward enantioselective synthesis of (+)-goniodiol (**1**) has been achieved with an overall yield of 5.9% and 98% ee. The approach involved a two-stereocentred Co-catalyzed HKR of racemic epoxide and Lewis acid-mediated diastereoselective allylation of aldehyde as the key chiral inducing steps. This methodology is also amenable for a viable synthesis of other diastereomers (**3-5**) of styryllactone family by suitably employing (*R, R*)-salen Co(OAc) complex for HKR and $\text{BF}_3 \cdot \text{OEt}_2$ for Lewis acid-mediated diastereoselective allylation respectively.

1.1.7 Experimental section

(3-Phenyloxiran-2-yl)methanol (**95**)

To a stirred solution of cinnamyl alcohol **62** (1.8 g, 13.415 mmol) in CH_2Cl_2 at 0 °C was added *m*-chloroperoxybenzoic acid (5.0 g, 28.973 mmol) in portions. After being stirred for 14 h at 0 °C, the mixture was quenched with NaHSO_3 . The aqueous layer

was extracted with CH₂Cl₂ (3x75 mL) and washed with NaHCO₃. The combined organic extracts were dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude epoxy alcohol, which was purified by column chromatography on silica gel with pet. ether:EtOAc (85:15) gave the pure epoxy alcohol **95** (1.77 g, 88%).

Yield: 88%; colorless oil; **IR** (CHCl₃, cm⁻¹): ν_{\max} 669, 757, 929, 1025, 1069, 1216, 1384, 2927, 3019, 3435; **¹H NMR** (200 MHz, CDCl₃): δ 1.98 (dd, $J = 5.3, 7.8$ Hz, 1H), 3.19-3.23 (m, 1H), 3.74-3.86 (m, 1H), 3.92 (d, $J = 2.2$ Hz, 1H), 4.00-4.10 (m, 1H), 7.24-7.36 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.6, 61.2, 62.5, 125.7, 128.3, 128.4, 136.6; **Analysis** for C₉H₁₀O₂ requires: C, 71.98; H, 6.71%; found: C, 71.94; H, 6.69%.

3-Methoxy-3-phenylpropane-1,2-diol (96)

To a stirred solution of epoxy alcohol **95** (2.1 g, 13.984 mmol) in MeOH (75 mL) at 25 °C was added camphor sulfonic acid (0.32 g, 1.39 mmol) and the mixture was stirred for 1 h. After completion of the reaction as monitored by TLC, solvent was distilled off. The crude residue was dissolved in EtOAc, washed with water, dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude diol. The crude diol was then purified by column chromatography over silica gel with pet. ether:EtOAc (60:40) to give methoxy diol **96** (2.45 g, 96%).

Yield: 96%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 720, 845, 1065, 1125, 1654, 2985, 3085, 3465; **¹H NMR** (200 MHz, CDCl₃): δ 2.32 (br s, 1H), 3.30 (s, 3H), 3.63-3.75 (m, 3H), 4.32 (d, $J = 5.1$ Hz, 1H), 7.33-7.36 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 56.6, 62.2, 75.7, 84.5, 127.5, 128.2, 128.4, 137.9; **Analysis** for C₁₀H₁₄O₃ requires: C, 65.91; H, 7.74; found: C, 65.88; H, 7.64%.

(2*SR*, 3*RS*)-3-Methoxy-3-phenyl-1,2-epoxypropane (97)

To a stirred solution of diol **96** (4 g, 21.951 mmol) in dry CH₂Cl₂ (100 mL) were added Bu₂SnO (107 mg, 0.43 mmol) and *p*-toluenesulfonyl chloride (4.61 g, 24.2 mmol) followed by triethylamine (6.11 mL, 43.902 mmol) and 4-dimethylaminopyridine (0.268 g, 2.19 mmol) at 0 °C. The reaction mixture was slowly warmed to 25 °C and stirred for 1.5 h. After completion of the reaction as monitored by TLC, it was diluted with CH₂Cl₂ (3x75 mL) and extracted. The combined organic phases were washed with water, brine, dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude product. The crude tosylate was dissolved in methanol (100 mL) and anhyd. K₂CO₃ (4 g, 28.941 mmol) was added to it and the entire contents were stirred for 30 min at 0 °C. The reaction mass was concentrated and the crude residue dissolved in EtOAc, washed with water, brine, dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude epoxide, which was then purified by column chromatography over silica gel with pet. ether:EtOAc (85:15) to give the racemic methoxy epoxide **97** (2.739 g, 76% over two steps).

Yield: 76% for two steps; colorless liquid; **IR** (CHCl₃, cm⁻¹): ν_{\max} 685, 757, 1035, 1215, 1620, 2978, 3018, 3069; **¹H NMR** (200 MHz, CHCl₃): δ 2.59 (dd, *J* = 2.6, 4.8 Hz, 1H), 2.73 (t, *J* = 4.2 Hz, 1H), 3.14-3.21 (m, 1H), 3.37 (s, 3H), 3.86 (d, *J* = 6.6 Hz, 1H), 7.31-7.42 (m, 5H); **¹³C NMR** (50 MHz, CHCl₃): δ 43.8, 55.0, 55.8, 85.0, 127.4, 128.1, 128.4, 137.8; **Analysis** for C₁₀H₁₂O₂ requires: C, 73.15; H, 7.37; found: C, 73.09; H, 7.31%.

(2*R*, 3*S*)-3-Methoxy-3-phenyl-1,2-epoxypropane (98)

To a solution of (*S*, *S*)-salenCo(II) complex (purchased from Aldrich, USA.) (0.043 g, 0.07 mmol) in toluene (2.0 mL) was added gl. AcOH (0.04 g, 7.3 mmol). It was

allowed to stir at 25 °C in air for 30 min over which time the color was changed from orange-red to a dark brown. It was then concentrated *in vacuo* to get (salen)Co(III)(OAc) complex as brown colored solid. To a stirred solution of (salen)Co(III)(OAc) complex (0.004 g, 0.5 mol%) and racemic methoxy epoxide **97** (1.41 mmol) in THF (0.5 mL) at 0 °C was added H₂O (0.012 g, 0.7 mmol) drop-wise over 5 min. The reaction mixture was allowed to warm to 25 °C and stirred for 14 h. After completion of reaction (monitored by TLC), solvent was removed *in vacuo*. The crude product was purified by column chromatography over silica gel to give (2*R*, 3*S*)-3-methoxy-3-phenyl-1,2-epoxypropane (**98**) on eluting with pet. ether:EtOAc (85:15) and (2*S*, 3*R*)-3-methoxy-3-phenylpropane-1,2-diol (**99**) with pet. ether:EtOAc (60:40) in high enantiomeric excess.

The chiral methoxy epoxide **98** was obtained as a colorless liquid (48% and 96% ee); [%ee was determined by transforming epoxide **98** to the corresponding diol, (0.5M NaOH, *tert*-BuOH-H₂O (3:1), 75 °C, 8 h) and comparing its specific rotation with its antipode **99**]; **Yield**: 48%; colorless liquid; $[\alpha]_{25}^D - 111.34$ (*c* 1.02, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 685, 757, 1035, 1215, 1620, 2978, 3018, 3069; **¹H NMR** (200 MHz, CHCl₃): δ 2.59 (dd, *J* = 2.6, 4.8 Hz, 1H), 2.73 (t, *J* = 4.2 Hz, 1H), 3.14-3.21 (m, 1H), 3.37 (s, 3H), 3.86 (d, *J* = 6.6 Hz, 1H), 7.31-7.42 (m, 5H); **¹³C NMR** (50 MHz, CHCl₃): δ 43.8, 55.0, 55.8, 85.0, 127.4, 128.1, 128.4, 137.8; **Analysis** for C₁₀H₁₂O₂ requires: C, 73.15; H, 7.37; found: C, 73.09; H, 7.31%.

(2*S*, 3*R*)-3-Methoxy-3-phenylpropane-1,2-diol (**99**)

Yield: 47%; colorless gum; $[\alpha]_{25}^D + 113.95$ (*c* 1.16, CHCl₃); lit.³⁵ $[\alpha]_{25}^D + 113.97$ (*c* 1.016, CHCl₃); 98% ee by chiral HPLC analysis (Chiralcel OJ-H, *n*-hexane/ *i*PrOH,

86:14, 0.5 mL/min) retention time 24.16 (1.08%) and 25.50 (98.92%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 720, 845, 1065, 1125, 1654, 2985, 3085, 3465; **^1H NMR** (200 MHz, CDCl_3): δ 2.32 (br s, 1H), 3.30 (s, 3H), 3.63-3.75 (m, 3H), 4.32 (d, $J = 5.1$ Hz, 1H), 7.33-7.36 (m, 5H); **^{13}C NMR** (50 MHz, CDCl_3): δ 56.6, 62.2, 75.7, 84.5, 127.5, 128.2, 128.4, 137.9; **HRMS** (ESI) m/z calcd for $\text{C}_{10}\text{H}_{14}\text{O}_3$ $[\text{M} + \text{Na}]^+$: 205.8811, found: 205.8670; **Analysis** for $\text{C}_{10}\text{H}_{14}\text{O}_3$ requires: C, 65.91; H, 7.74; found: C, 65.88; H, 7.64%.

(1R, 2S)- 3-tert-Butyldiphenylsilyloxy-1-methoxyphenylpropan-2-ol (100)

To a stirred solution of diol **99** (4 g, 21.951 mmol) in dry CH_2Cl_2 (110 mL) was added imidazole (2.24 g, 32.926 mmol) at 0 °C. After stirring for 10 min, TBSCl (3.97 g, 30.91 mmol) was added and the reaction mixture was stirred at 25 °C for 1 h. After completion of the reaction as monitored by TLC, it was poured into water and extracted with CH_2Cl_2 (3x40 mL). The combined organic phases were washed with brine, dried over anhyd. Na_2SO_4 and concentrated under reduced pressure to get the crude product. The crude product was then purified by column chromatography over silica gel with pet. ether:EtOAc (95:5) to give the silyl ether **100** (5.86 g, 90%).

Yield: 90%; viscous liquid; $[\alpha]_{25}^{\text{D}} +59.62$ (c 1.88, CHCl_3); **IR** (CHCl_3 , cm^{-1}): ν_{max} 710, 815, 1042, 1145, 1257, 2985, 3077, 3455; **^1H NMR** (200 MHz, CDCl_3): δ 0.07 (d, $J = 2.5$ Hz, 6H), 0.9 (s, 9H), 2.30 (d, $J = 4.4$ Hz, 1H), 3.22 (s, 3H), 3.69-3.76 (m, 3H), 4.13 (d, $J = 6.1$ Hz, 1H), 7.33-7.36 (m, 5H); **^{13}C NMR** (50 MHz, CDCl_3): δ -5.4, 18.3, 25.9, 56.8, 63.5, 74.5, 83.8, 127.7, 128.0, 128.2, 138.6; **LCMS** calcd for $\text{C}_{16}\text{H}_{28}\text{O}_3\text{Si}$ $[\text{M} + \text{Na}]^+$: 319.1709, found: 319.1713; **Analysis** for $\text{C}_{16}\text{H}_{28}\text{O}_3\text{Si}$ requires: C, 64.82; H, 9.52; found: C, 64.79; H, 9.44%.

(1R, 2S)-2-Benzoyloxy-3-tert-butyldiphenylsilyloxy-1-methoxy-1-phenylpropane (101)

To a stirred solution of alcohol **100** (6 g, 20.237 mmol) in dry DMF was slowly added 60% NaH in oil suspension (1.13 g, 28.331 mmol) followed by the addition of benzyl bromide (3.46 g, 20.237 mmol). The reaction mixture was stirred at 25 °C for 2 h, quenched with cold water and extracted with ether (3x100 mL). The combined organic layers were washed with brine, dried over anhyd. Na₂SO₄ and concentrated to give the crude material, which was then purified by column chromatography over silica gel with pet. ether:EtOAc (97:3) to give the protected ether **101** (7.43 g, 95%).

Yield: 95%; colorless viscous liquid; $[\alpha]_{25}^D +27.4$ (*c* 1.6, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 695, 990, 1045, 1210, 1268, 1343, 1425, 2860, 3048; **¹H NMR** (200 MHz, CDCl₃): δ 0.02 (d, *J* = 3.5 Hz, 6H), 0.90 (s, 9H), 3.28 (s, 3H), 3.46-3.55 (m, 2H), 3.67-3.74 (m, 1H), 4.36 (d, *J* = 4.1 Hz, 1H), 4.43, (s, 2H), 7.33-7.38 (m, 10H); **¹³C NMR** (50 MHz, CDCl₃): δ -5.4, 18.3, 26.0, 57.3, 62.5, 73.7, 82.9, 83.2, 127.3, 128.7, 138.6, 139.4; **Analysis** for C₂₃H₃₄O₃Si requires: C, 71.46; H, 8.86; found: C, 71.42; H, 8.83%.

(1R, 2S)-2-(Benzoyloxy)-1-methoxy-1-phenylpropan-1-ol (102)

To a stirred solution of protected ether **101** (6 g, 15.519 mmol) in dry THF (100 mL) was added tetra-*n*-butylammonium fluoride (6.08 g, 23.28 mmol) at 25 °C. The reaction mixture was stirred for 8 h and then diluted with water and extracted with EtOAc (3x100 mL). The organic layer was washed with brine, dried over anhyd. Na₂SO₄ and concentrated to give the crude material, which was then purified by column chromatography over silica gel with pet. ether:EtOAc (8:2) to give the alcohol **102** (3.85 g, 91%).

Yield: 91%; colorless liquid; $[\alpha]_{25}^D +42.32$ (*c* 1.62, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 690, 773, 837, 1101, 1255, 1471, 2359, 2823, 2856, 2930, 2955, 3495; **¹H NMR** (200

MHz, CDCl₃): δ 2.32 (t, J = 6.2 Hz, 1H), 3.26 (s, 3H), 3.46-3.55 (m, 1H), 3.78 (t, J = 6.1 Hz, 2H), 4.13-4.35 (m, 3H), 7.21-7.24 (m, 3H), 7.30-7.35 (m, 7H); ¹³C NMR (50 MHz, CDCl₃): δ 56.9, 62.1, 72.7, 82.2, 84.2, 127.6, 127.8, 128.2, 137.8, 139.1; **Analysis** for C₁₇H₂₀O₃ requires: C, 74.97; H, 7.40; found: C, 74.93; H, 7.38%.

(1R, 2S, 3R)-2-(Benzyloxy)-1-methoxy-1-phenyl-hex-5-en-3-ol (103)

To a stirred solution of alcohol **102** (3.5 g, 12.851 mmol) in dry DMSO (100 mL) was slowly added 2-iodoxybenzoic acid (4.32 g, 15.421 mmol). The reaction mixture was stirred for 2 h at 25 °C and quenched with cold water. It was filtered and the filtrate was then extracted with ether (3x100 mL). The combined organic layers were washed with brine, dried over anhyd. Na₂SO₄ and concentrated to give the crude aldehyde, which was directly used for the next step without purification. To a stirred crude aldehyde solution in dry CH₂Cl₂ (100 mL) was added TiCl₄ (4.01 mL, 1M solution in CH₂Cl₂) at -78 °C. After stirring for 10 min, tributyl allyl stannane (8.15 g) was added. The reaction mixture was stirred at -78 °C for 30 min, quenched with NaHCO₃, extracted with CH₂Cl₂ (3x100 mL). The combined organic layers were washed with brine, dried over anhyd. Na₂SO₄ and concentrated to give the crude material, which was then purified by column chromatography over silica gel with pet. ether:EtOAc (85:15) to give the alcohol **103** (2.93 g, 73%).

Yield: 73%; colorless liquid; $[\alpha]_{25}^D +18.21$ (c 1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 756, 1109, 1209, 1396, 1496, 1650, 1980, 3020, 3063, 3479; **¹H NMR** (200 MHz, CDCl₃): δ 2.34 (t, J = 6.1 Hz, 2H), 2.60 (d, J = 8.7 Hz, 1H), 3.25 (s, 3H), 3.37-3.42 (m, 1H), 3.84-3.95 (m, 2H), 4.15 (d, J = 10.9 Hz, 1H), 4.33 (d, J = 7.8 Hz, 1H), 5.03-5.10 (m, 2H), 5.76-5.93 (m, 1H), 7.23-7.37 (m, 10H); **¹³C NMR** (50 MHz, CDCl₃): δ 38.7, 56.7, 70.5, 74.0, 77.1, 83.0, 117.0, 127.7, 128.0, 135.3, 137.6, 139.4; **Analysis** for C₂₀H₂₄O₃ requires: C, 76.89; H, 7.74; found: C, 76.86; H, 7.71%.

[(1R, 2S, 3R)-2-(Benzyloxy)-3-(methoxy)-3-phenyl-1-(prop-2-enyl)propyl] prop-2 enoate (104)

A stirred solution of alcohol **103** (1 g, 3.2 mmol) in dry CH₂Cl₂ (20 mL) was cooled to 0 °C followed by addition of cinnamoyl chloride (2.13 g, 12.8 mmol), dry pyridine (1.03 mL, 12.80 mmol) and 4-dimethylaminopyridine (0.12 g, 1.0 mmol). The entire reaction mixture was then warmed to 25 °C and stirred for 12 h. The resulting mixture was filtered through a short pad of silica to remove solid pyridinium hydrochloride and washed thoroughly with ether. Solvent was removed under vacuum to give the crude product, which was then purified by column chromatography over silica gel with pet. ether:EtOAc (95:5) to give cinnamate ester **104** (0.906 g, 64%).

Yield: 64%; colorless liquid; $[\alpha]_{25}^D +72.39$ (*c* 1.2, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 767, 1118, 1203, 1309, 1454, 1637, 1714, 2885, 3030, 3063; **¹H NMR** (200 MHz, CDCl₃): δ 2.35- 2.55 (m, 2H), 3.24 (s, 3H), 3.68-3.73 (m, 1H), 4.39 (d, *J* = 6.6 Hz, 2H), 4.58-4.84 (m, 3H), 4.96 (d, *J* = 6.6 Hz, 2H) 5.03 (br s, 1H), 5.52-.5.71 (m, 1H), 6.43 (d, *J* = 16.0 Hz, 1H), 7.30-7.42 (m, 15H), 7.63 (d, *J* = 16.0 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 35.9, 56.7, 72.5, 74.7, 82.4, 82.6, 118.0, 118.4, 127.7, 128.3, 128.9, 130.1, 133.9, 137.7, 139.8, 144.9, 166.3; **LCMS** calcd for C₂₉H₃₀O₄ [M + Na]⁺: 465.2042, found: 465.2961; **Analysis** for C₂₉H₃₀O₄ requires: C, 78.71; H, 6.83; found: C, 78.62; H, 6.80%.

(6R)-6-[(1S, 2R)-1-(Benzyloxy)-2-(methoxy)-2-phenylethyl]-5,6-dihydropyran-2-one (105)

A solution of Grubbs' II generation catalyst [RuCl₂(=CHPh)(PCy₃)(*bismesitylimidazo*-lidinylidene)] (0.115 g, 10 mol%) in dry CH₂Cl₂ (10 mL) was added dropwise to a refluxing solution of cinnamoyl ester **104** (0.6 g, 1.356 mmol) in CH₂Cl₂ (30 mL). Heating to reflux was continued for 8 h till starting material was consumed completely (TLC). Solvent was distilled off under reduced pressure and the crude

product was then purified by column chromatography over silica gel with pet. ether:EtOAc (75:25) to give lactone **105** (0.348 g, 76%).

Yield: 76%; colorless liquid; $[\alpha]_{25}^D +84.31$ (*c* 0.8, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 754, 815, 960, 1066, 1120, 1240, 1382, 1456, 1720, 2330, 2823, 2899, 2933, 3030, 3063; **¹H NMR** (200 MHz, CDCl₃): δ 1.93-2.07 (m, 1H), 2.47-2.65 (m, 1H), 3.25 (s, 3H), 3.40 (dd, *J* = 2.2, 8.9 Hz, 1H), 3.78 (s, 2H), 4.55 (d, *J* = 8.9 Hz, 1H), 4.85-4.94 (m, 1H), 5.96-6.02 (m, 1H), 6.79-6.88 (m, 1H), 7.24-7.45 (m, 10H); **¹³C NMR** (50 MHz, CDCl₃): δ 25.9, 56.5, 74.4, 80.7, 82.3, 121.3, 127.7, 128.1, 137.2, 139.4, 163.7; **Analysis** for C₂₁H₂₂O₄ requires: C, 74.54; H, 6.55; found: C, 74.48; H, 6.52%.

(6R)-6-[(1S, 2R)-1,2-Dihydroxy-2-phenylethyl]-5,6-dihydropyran-2-one:
[(+)-Goniodiol] (1)

To a stirred and cooled (-78 °C) solution of α -pyrone **105** (68 mg, 0.201 mmol) in dry CH₂Cl₂ (3 mL), was added BBr₃ (0.6 mL, 1M solution in CH₂Cl₂). After stirring for 6 h at -78 °C, it was quenched with an aq. solution of NaHCO₃. The reaction mixture was allowed to warm to 20 °C and was extracted with EtOAc (2x30 mL). The combined organic layers were dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude product. The crude product was then purified by column chromatography over silica gel with pet. ether:EtOAc (20:80) to give (+)-goniodiol (**1**) (0.034 g, 71%).

Yield: 71%; colorless liquid; $[\alpha]_{25}^D +73.77$ (*c* 0.24, CHCl₃); lit.¹⁵ $[\alpha]_{25}^D +74.4$ (*c* 0.3, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 1035, 1259, 1380, 1690, 3390; **¹H NMR** (200 MHz, CDCl₃): δ 2.27-2.40 (m, 2H) 2.78 - 3.12 (m, 2H), 4.10 (t, *J* = 7.8 Hz, 1H), 5.18 - 5.24 (m, 2H), 6.06 (dd, *J* = 3.6, 7.2 Hz, 1H), 6.98 (ddd, *J* = 2.9, 6.4, 8.1 Hz, 1H) 7.27-7.38 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 26.5, 73.8, 75.1, 76.4, 121.03, 125.8, 128.4,

129.0, 138.5, 145.4, 163.2; **Analysis** for C₁₃H₁₄O₄ requires: C, 66.66; H, 6.02; found: C, 66.62; H, 6.01%.

Section II:

Enantioselective Synthesis of (-)-Polysphorin Analog *via* Co-Catalyzed Hydrolytic Kinetic Resolution of *syn*-(2*SR*, 3*SR*)-3-(Methoxymethyl)oxy-3-(3,4,5-trimethoxyphenyl)-1,2-epoxy propane

1.2.1 Introduction

Malaria is by far the most important tropical parasitic disease that kills more people than any other communicable diseases except for tuberculosis. In many developing countries, especially in Africa and Asia, malaria exacts an enormous toll in lives, in medical costs, and in days of labor lost. *Plasmodium falciparum* accounts for the majority of infections and is found to be the most lethal.³⁶ However, malaria is a curable disease if promptly diagnosed and adequately treated. Although several research programs are focused on various strategies to control malaria, drug discovery is one of the main areas of concentrated effort.³⁷ In humans, malaria develops *via* two phases: an hepatic phase and an erythrocytic phase. Most of the research programs were focused on the erythrocytic stage of *Plasmodium falciparum*. However, the development of drugs able to inhibit the parasite during the hepatic phase is of very high importance as they could act against relapse and constitute a radical cure of the parasite. Today, only two drugs, primaquine and tafenoquine, are active during the hepatic phase of parasite development. However, these drugs have a hematologic toxicity. Accordingly, there is a strong need of antimalarial drugs active during the hepatic phase of parasite development, and without any toxicity.

Polysphorin (**106**), a neolignan was isolated from *Piper polysporum* C in China,³⁸ and from the leaves and stems of *Rhaphidopora decursiva* in Vietnam (**Fig. 14**).³⁹ The structure and relative configuration of polysphorin (**106**) were established by synthetic

studies and X-ray crystallography.⁴⁰ Biological screening has since shown polysphorin (**106**) to possess *in vitro* antimalarial activity (IC₅₀ 400ng/ml).³⁹ Several other members of this family of neolignans have also been shown to display interesting biological properties: raphidecursinol B (**107**) shows activity against *Plasmodium falciparum*, the parasite responsible for the most severe forms of malaria,⁴¹ but is ten times less active than polysphorin (**106**);³⁹ neolignans virolin (**108**) and surinamensin (**109**) both displayed interesting and varied biological properties spanning from antimalarial to antifungal,⁴² antileishmanial, antioxidant, and antischistosomal activities.⁴³

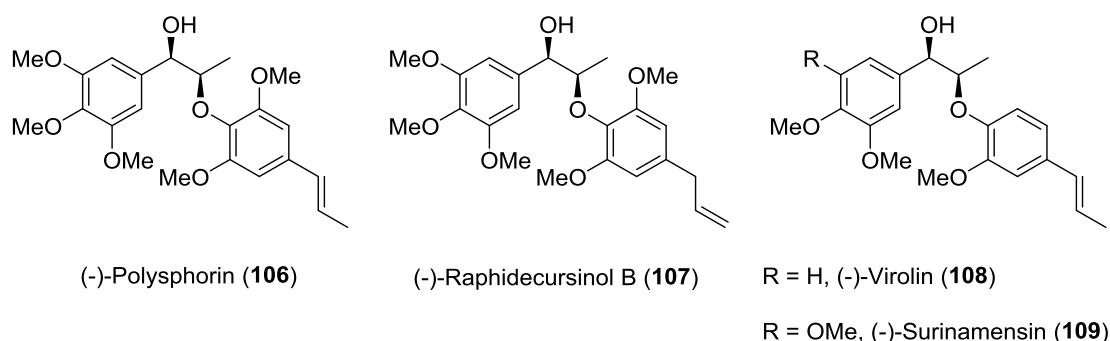
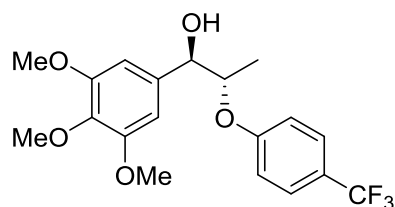
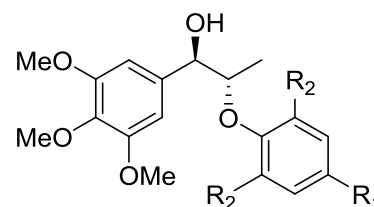


Fig. 14: Structures of bioactive neolignans

The pharmaceutically important biological profiles coupled with the scarce availability from natural sources make them ideal targets for testing new synthetic methodology and prompted many groups to attempt the laboratory synthesis of this class of compounds.^{40, 44-46} Synthetic methods also provide the opportunity to expand the diversity by generating non natural analogs. Recently, Choppin *et al*, synthesized new polysphorin analogs (**110a-x**), and it was found that both the *syn*- and *anti*-diastereomers are highly effective in the hepatic phase (**Fig. 15**).⁴⁵

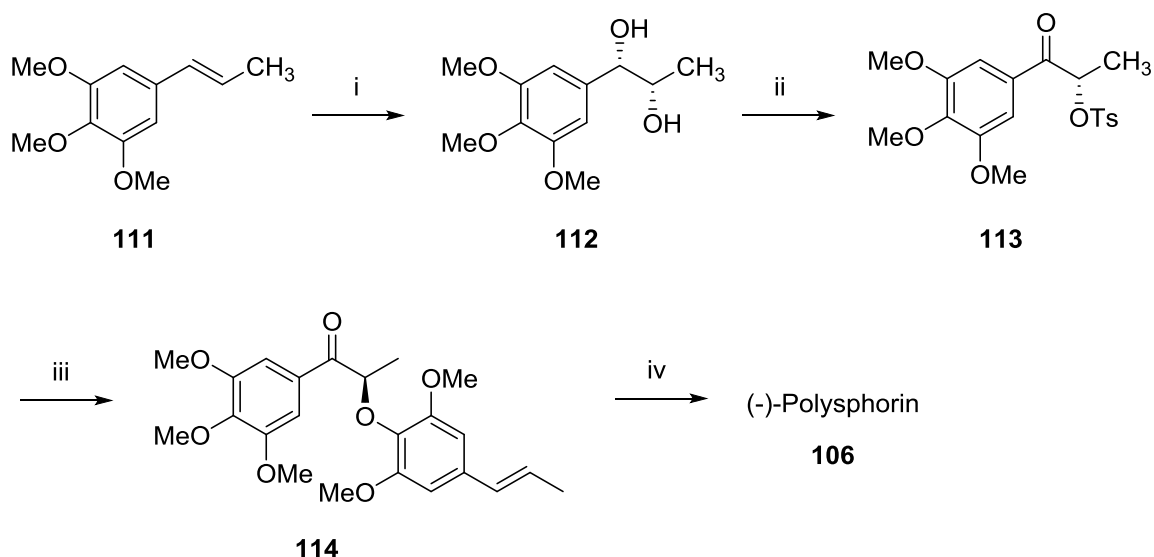
Polysphorin analog (**110a**)Polysphorin analogs (**110b-x**)R₁ = OMe, Cl, CN, NH₂, etcR₂ = OMe, OCF₃, CH₃, H, etc**Fig. 15:** Structures of bioactive polysphorin analogs

1.2.2 Review of Literature

Literature search reveals that there are only three reports on the asymmetric synthesis and one report on the racemic synthesis of polysphorin (**106**) and its analogs (**110a-x**) available. A short description of all the four reported methods is presented below.

Ley's approach (2003)⁴⁰

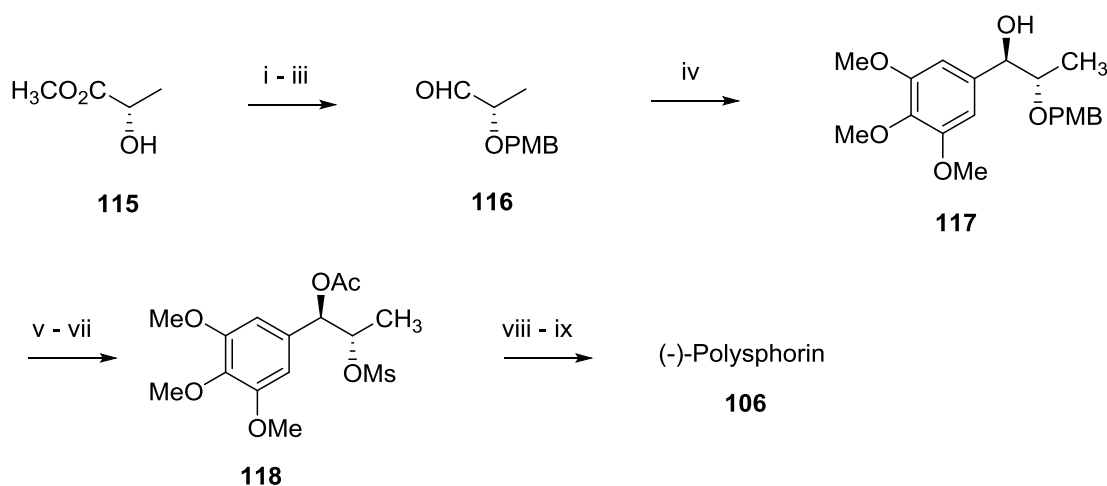
Ley *et al.* have reported the synthesis of polysphorin (**106**) by employing Sharpless asymmetric dihydroxylation as the key reaction. Thus, 1,2,3-trimethoxy-5-(*E*)-(propenyl)-benzene **111** was subjected to Sharpless asymmetric dihydroxylation using AD-mix- α , followed by purification under Frechet's protocol (polymer supported boronic acid) resulted in diol **112** in 96% ee and 92% yield. Tosylated α -hydroxy ketone **113** was prepared efficiently in two steps *via* selective benzylic oxidation of diol **112** using DDQ, followed by tosylation of α -hydroxy ketone. The S_N2 coupling of tosylate **113** with (*E*)-2,6-dimethoxy-4-(propenyl)-phenol using polymer-supported phosphazene base gave phenolic ether **114**. Finally, polymer-supported diastereoselective borohydride reduction of ketone **114** afforded (-)-polysphorin (**106**) (dr = 32 : 1) (**Scheme 18**).



Scheme 18: (i) AD-mix- α , $\text{CH}_3\text{SO}_2\text{NH}_2$, *tert*-BuOH:H₂O (1:1), 0 °C to 25 °C, 16 h, polymer-supported boronic acid, toluene reflux, 24 h, 83%; (ii) (a) DDQ, dioxane, 60 °C, 12 h, polymer-supported ascorbate, 25 °C, 16 h, 89%; (b) *p*-toluene sulfonic anhydride, pyridine, 0 °C, 1 h, 94%; (iii) (*E*)-2,6-dimethoxy-4-(propenyl)phenol, polymer-supported phosphazene base, CH_3CN , 25 °C, 15 h, 100%; (iv) polymer-supported borohydride, CH_3OH , 25 °C, 16 h, 96%, dr = 32:1.

Casiraghi's approach (2006)⁴⁴

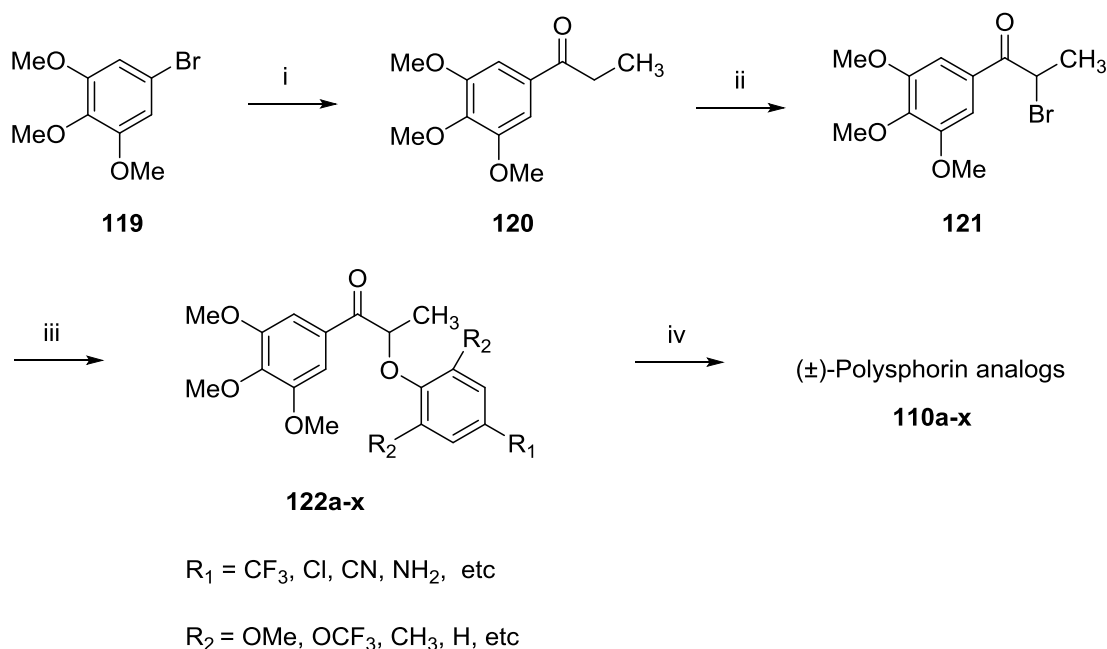
Casiraghi *et al.* have achieved the synthesis of (-)-polysphorin (**106**) using a chiral pool approach commencing from (*S*)-methyl lactate **115**. The hydroxy group of **115** was protected as its PMB ether, followed by reduction of ester group and oxidation of the resulting alcohol under Swern condition gave aldehyde **116** in 91% yield. Arylation of aldehyde **116** with lithium derivative of 5-bromo-1,2,3-trimethoxy benzene afforded the *anti*-configured compound **117** in 80% yield (*anti:syn* = 83:17). Benzylic alcohol **117** was converted to mesylate **118** using simple standard transformations. The $\text{S}_{\text{N}}2$ displacement of mesylate **118** with (*E*)-2,6-dimethoxy-4-(propenyl)-phenol under microwave irradiation and subsequent deacetylation afforded (-)-polysphorin (**106**) in 40% overall yield and 97% ee (**Scheme 19**).



Scheme 19: (i) PMBOC(CCl₃)=NH, Sc(OTf)₃, toluene; (ii) LiAlH₄, THF; (iii) (COCl)₂, DMSO, CH₂Cl₂, -80 °C, then Et₃N, 91%, (over three steps); (iv) 5-bromo-1,2,3 trimethoxy benzene, *tert*-BuLi, THF, -85 °C, 1 h, 80%, dr = 83:17; (v) Ac₂O, Et₃N, DMAP, CH₃CN, 25 °C, 1 h; (vi) DDQ, CH₂Cl₂, 25 °C, 3 h; (vii) MsCl, Et₃N, 25 °C, 3 h, 80% (over three steps); (viii) Cs₂CO₃, 18-crown-6, (*E*)-2,6-dimethoxy-4-(propenyl) phenol, DMF, MW 200 W, 2 bar, 120 °C, 10 min, 70%; (ix) NaOMe, CH₃OH, 25 °C, 1 h, 98%.

Choppin's approach (2010)⁴⁵

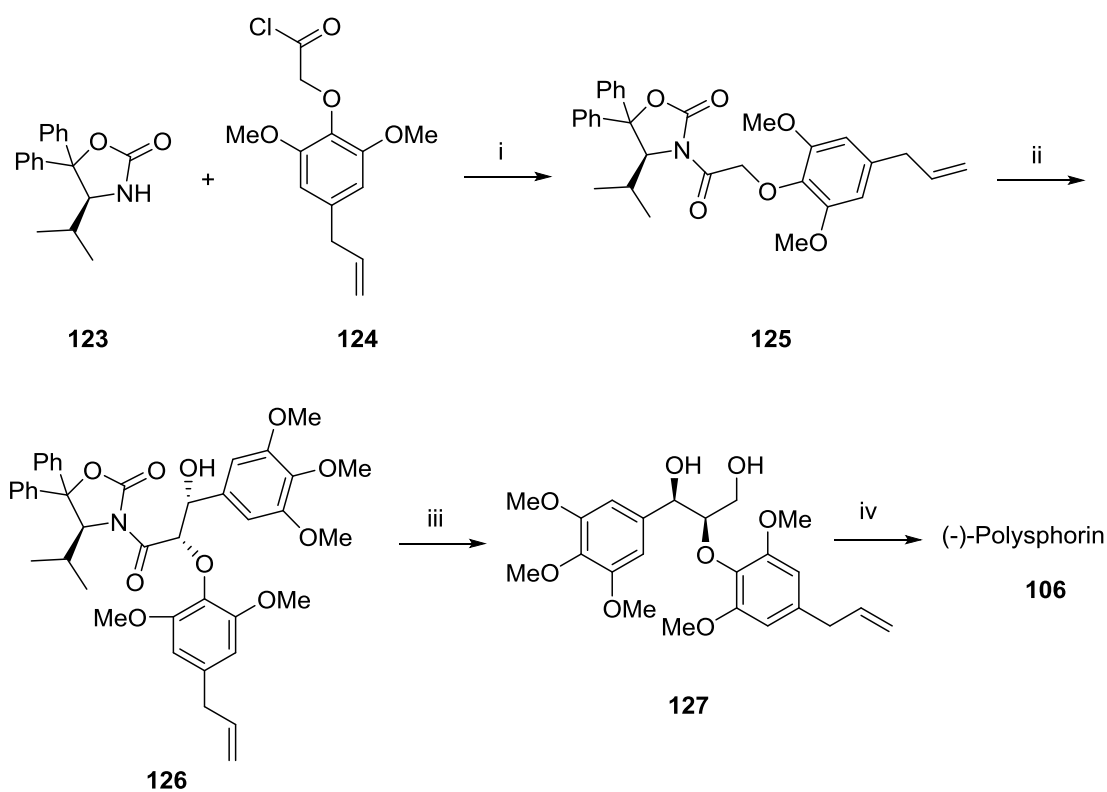
Choppin *et al.* have developed a useful synthetic method for the racemic synthesis of *syn*-(±)-polysphorin analogs (**110a-x**). Reaction of 5-bromo-1,2,3-trimethoxy benzene **119** with *N,N*-dimethylpropionamide in the presence of *tert*-BuLi afforded aryl ketone **120** in 49% yield. α-Bromination of ketone **120** gave bromo compound **121**, which on substitution with different phenols resulted in aryl keto ethers **122a-x**. Finally, diastereoselective reduction of ketones **122a-x** using ZnCl₂ and NaBH₄ afforded (±)-polysphorin analogs (**110a-x**) with diastereomeric ratio of *anti:syn* in 69:31 to 85:15 (**Scheme 20**).



Scheme 20: (i) *tert*-BuLi, THF, -78 °C, *N,N*-dimethylpropionamide, 1 h, 49%; (ii) Br₂, toluene, 25 °C, 1 h, 98%; (iii) substituted phenol, K₂CO₃, butan-2-one, reflux, 16 h, 59-90%; (iv) ZnCl₂, NaBH₄, Et₂O, 0 °C, 2 h, 84-90%, *anti:syn* = 69:31 to 85:15.

Gawali's approach (2012)⁴⁶

Gawali *et al.* have utilized valine-based chiral oxazolidinone **123** for the introduction of chirality in (-)-polysphorin (**106**) synthesis. Chiral oxazolidinone **123** was thus attached to aryloxyacid chloride derivative **124** under basic conditions to afford *N*-acylimidazolidinone derivative **125**. Aldol reaction between boron enolate of imidazolidinone **125** with 3,4,5-trimethoxybenzaldehyde afforded *syn*-aldol product **126** in 86% yield and as a single diastereomer. The reductive removal of the chiral auxiliary afforded the diol **127**. Further, deoxygenation of the primary alcohol in **127** followed by olefin isomerization delivered (-)-polysphorin (**106**) (**Scheme 21**).



Scheme 21: (i) *n*-BuLi, THF, 0 °C to 25 °C, 92%; (ii) *n*-Bu₂BOTf, ^tPr₂NEt, CH₂Cl₂, 0 °C to 25 °C, 3,4,5-trimethoxybenzaldehyde, -78 °C to 25 °C, 86%; (iii) LiAlH₄, Et₂O, 25 °C, 4 h, 87%; (iv) (a) *p*-TsCl, cat. DMAP, Et₃N, CH₂Cl₂, 0 °C to 25 °C, 3 h; (b) LiAlH₄, Et₂O, 25 °C, 2 h, 98%; (c) PdCl₂, CH₃OH, 25 °C, 6 h, 97%.

1.2.3 Present Work

1.2.3.1 Objective

From the above discussions, it is obvious that only a few reports are available for the synthesis of polysphorin (**106**) and its analogs (**110a-x**); Notably, some of the reported methods suffer from certain limitations such as use of chiral starting material, chiral auxiliary, expensive reagents, lack of selectivity, etc. Hence, a catalytic method that introduces chirality into the molecule, along with a flexibility to obtain all the four diastereoisomers is desirable. This section describes an elegant

synthetic route to the synthesis of (-)-polysphorin analog (**110a**) using a late-stage two-stereocentered Co-catalyzed HKR of racemic *syn*-epoxide **136** as the key step.

Retrosynthetic analysis of (-)-polysphorin analog (**110a**) reveals that alcohol **139** could be visualized as the key intermediate. The alcohol **139** could be obtained by means of the regioselective opening of chiral epoxide **138**. The chiral epoxide **138** could in turn be obtained by performing Co-catalyzed two-stereocentered HKR of the corresponding racemic *syn*-alkoxy epoxide **136**. The required racemic *syn*-alkoxy epoxide **136** could be prepared from *syn*-diol **130**, which in turn could be readily prepared from aldehyde **128** (Fig. 16).

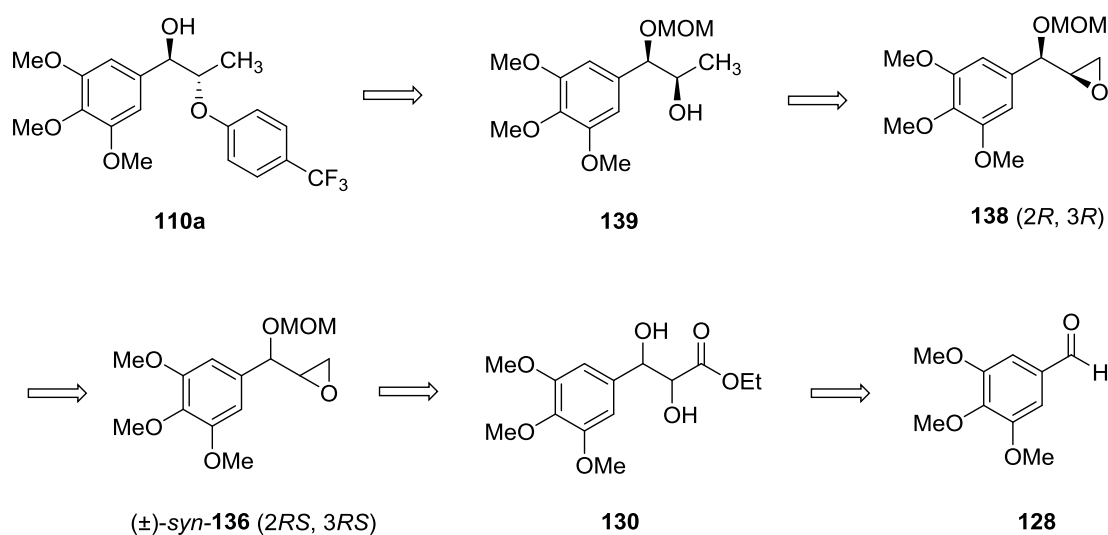
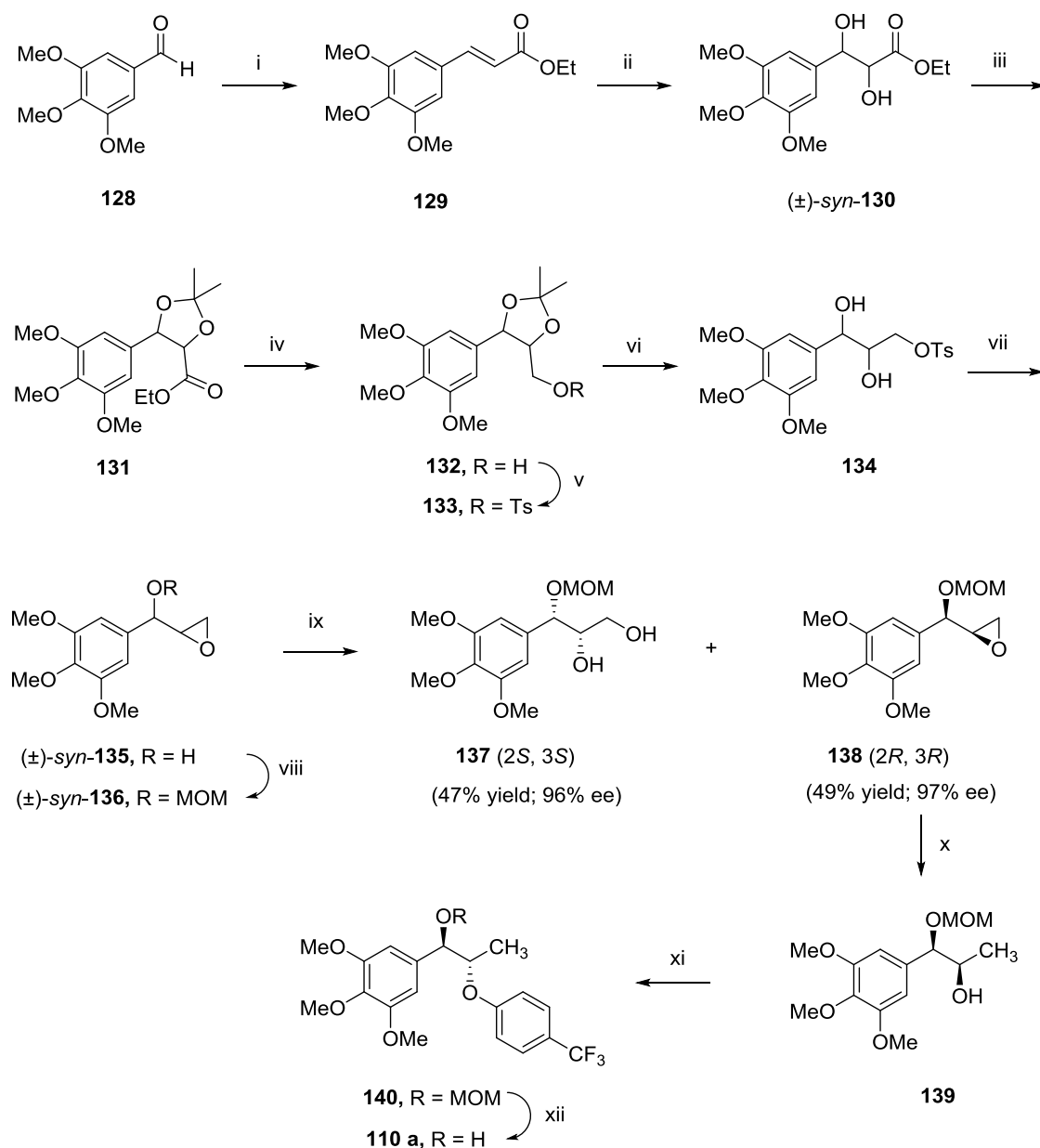


Fig. 16: Retrosynthetic analysis of (-)-polysphorin analog (**110 a**)

1.2.4 Results and Discussion

The complete synthetic sequence for (-)-polysphorin analog (**110a**) is shown in **Scheme 22**, wherein late-stage two-stereocentered Co-catalyzed HKR of racemic *syn*-epoxide constitutes a key step for the introduction of chirality in the molecule.



Scheme 22: (i) $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$, CH_2Cl_2 , 25 °C, 5 h, 96%; (ii) $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$ (2 mol%), NMO, acetone: H_2O (4:1), 0 to 25 °C, 20 h, 93%; (iii) 2,2-dimethoxypropane, camphor sulfonic acid (10 mol %), CH_2Cl_2 , 25 °C, 6 h, 98%; (iv) LiAlH_4 , THF, 0 to 25 °C, 1 h, 92%; (v) *p*-TsCl, Et_3N , DMAP (10 mol%), CH_2Cl_2 , 25 °C, 4 h, 94%; (vi) camphor sulfonic acid (10 mol %), CH_3OH , 40 °C, 1 h, 96%; (vii) K_2CO_3 , CH_3OH , 25 °C, 3 h, 88%; (viii) MOMCl, $i\text{Pr}_2\text{NEt}$, CH_2Cl_2 , 25 °C, 8 h, 92%; (ix) (*S,S*)-Co(salen)OAc (0.5 mol%), H_2O (0.5 equiv), 0 °C, 14 h; (x) LiAlH_4 , THF, 0 to 25 °C, 4 h, 96%; (xi) PPh_3 , $i\text{PrO}_2\text{C-N=N-CO}_2\text{Pr}^i$, 4-(trifluoromethyl)phenol, THF, 70 °C, 12 h, 74%; (xii) conc. HCl, CH_3OH , 25 °C, 4 h, 86%.

The present synthesis of (-)-polysphorin analog (**110a**) started from commercially available 3,4,5-trimethoxybenzaldehyde **128**, which on Wittig olefination (CH_2Cl_2 , $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$) gave the cinnamate ester **129** in 96% yield. The formation of cinnamate ester **129** was confirmed from its ^1H NMR spectrum, which showed two doublets at δ 6.31 (d, $J = 16$ Hz, 1H) and 7.57 (d, $J = 16$ Hz, 1H) establishing the presence of α - and β - CH olefin protons of $-\text{CH}=\text{CHCO}_2\text{Et}$. This was further ascertained by the presence of characteristic carbon signals at δ 117.3 and 144.3 corresponding to α - and β - carbon signal of $-\text{CH}=\text{CHCO}_2\text{Et}$ in its ^{13}C NMR spectrum (**Fig. 17**).

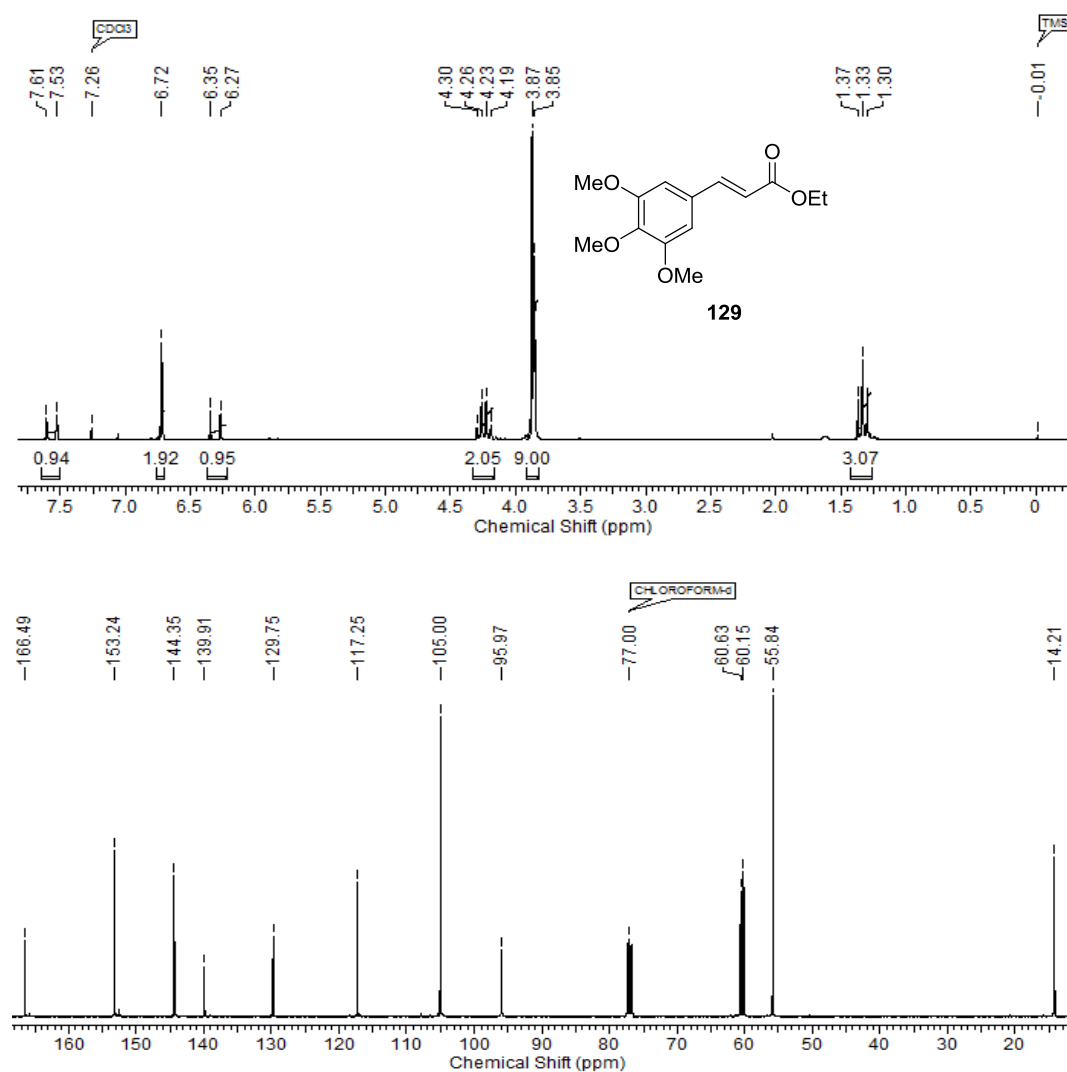


Fig. 17: ^1H and ^{13}C NMR spectra of cinnamate ester **129**

The Upjohn dihydroxylation of cinnamate ester **129** with catalytic amount of $K_2OsO_4 \cdot 2H_2O$ and NMO gave racemic *syn*-diol **130** in 93% yield.⁴⁷ The formation of diol **130** was confirmed from its 1H NMR spectrum, which showed characteristic multiplets at δ 4.34-4.36 (m, 1H) and 4.93-4.96 (m, 1H), corresponding to methine (-CH(OH)-CH(OH)-) protons, while its ^{13}C NMR spectrum showed characteristic carbon signals at δ 74.8 and 77.2 due to methine carbons attached to hydroxyl groups (Fig. 18).

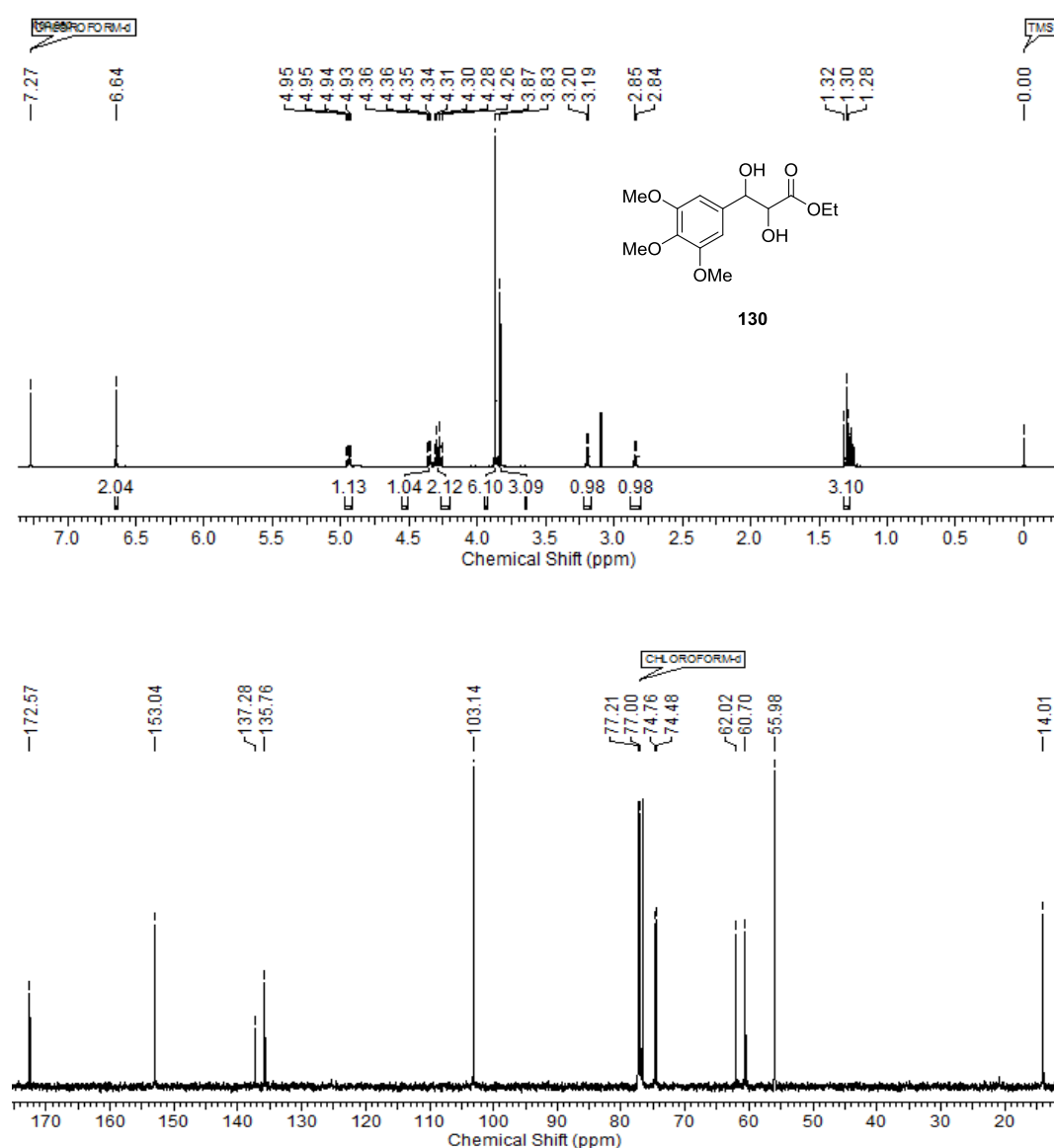


Fig. 18: 1H and ^{13}C NMR spectra of racemic *syn*-diol **130**

The racemic *syn*-diol **130** was then protected as its acetonide **131** on treatment with 2, 2-dimethoxypropane in the presence of catalytic amount of camphor sulfonic acid.⁴⁸ Acetonide **131** was confirmed from its ¹H NMR spectrum, which showed two characteristic singlets at δ 1.55 (s, 3H) and 1.61 (s, 3H) corresponding to the methyl protons of isopropylidene group, while its ¹³C NMR spectrum showed characteristic carbon signals at δ 25.7 and 26.9 due to methyl carbon of isopropylidene group (Fig. 19).

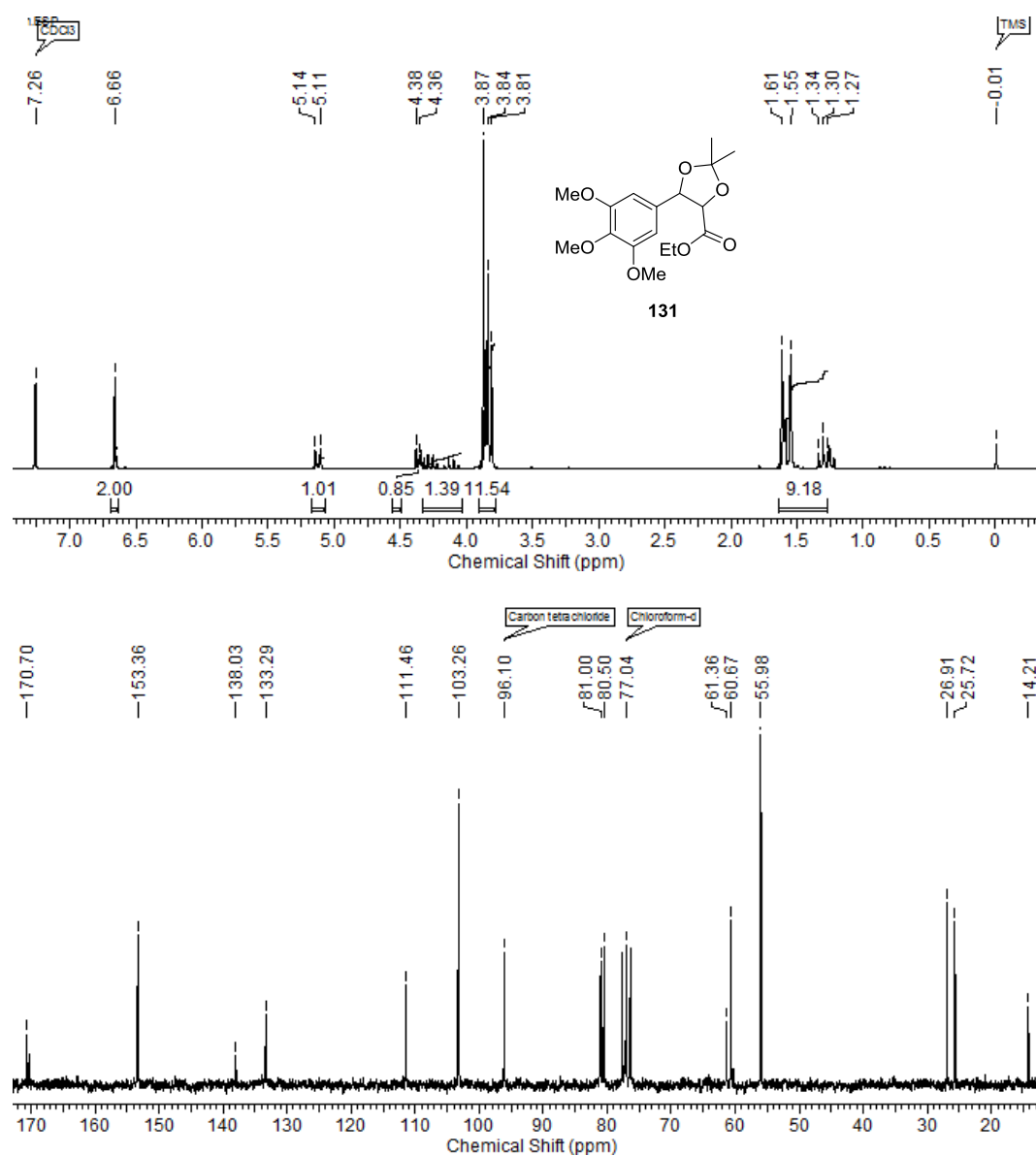


Fig. 19: ¹H and ¹³C NMR spectra of acetonide **131**

The reduction of the ester function in **131** was achieved on its treatment with LiAlH_4 in THF at room temperature to afford the corresponding alcohol **132** in 92% yield. Its ^1H NMR spectrum showed a characteristic multiplet at δ 3.79-3.82 (m, 2H) for methylene ($-\text{CH}_2\text{-OH}$) protons, while its ^{13}C NMR spectrum showed a typical carbon signal at δ 60.1 due to methylene carbon ($-\text{CH}_2\text{-OH}$) attached to hydroxyl group, which confirmed the formation of alcohol **132** (Fig. 20).

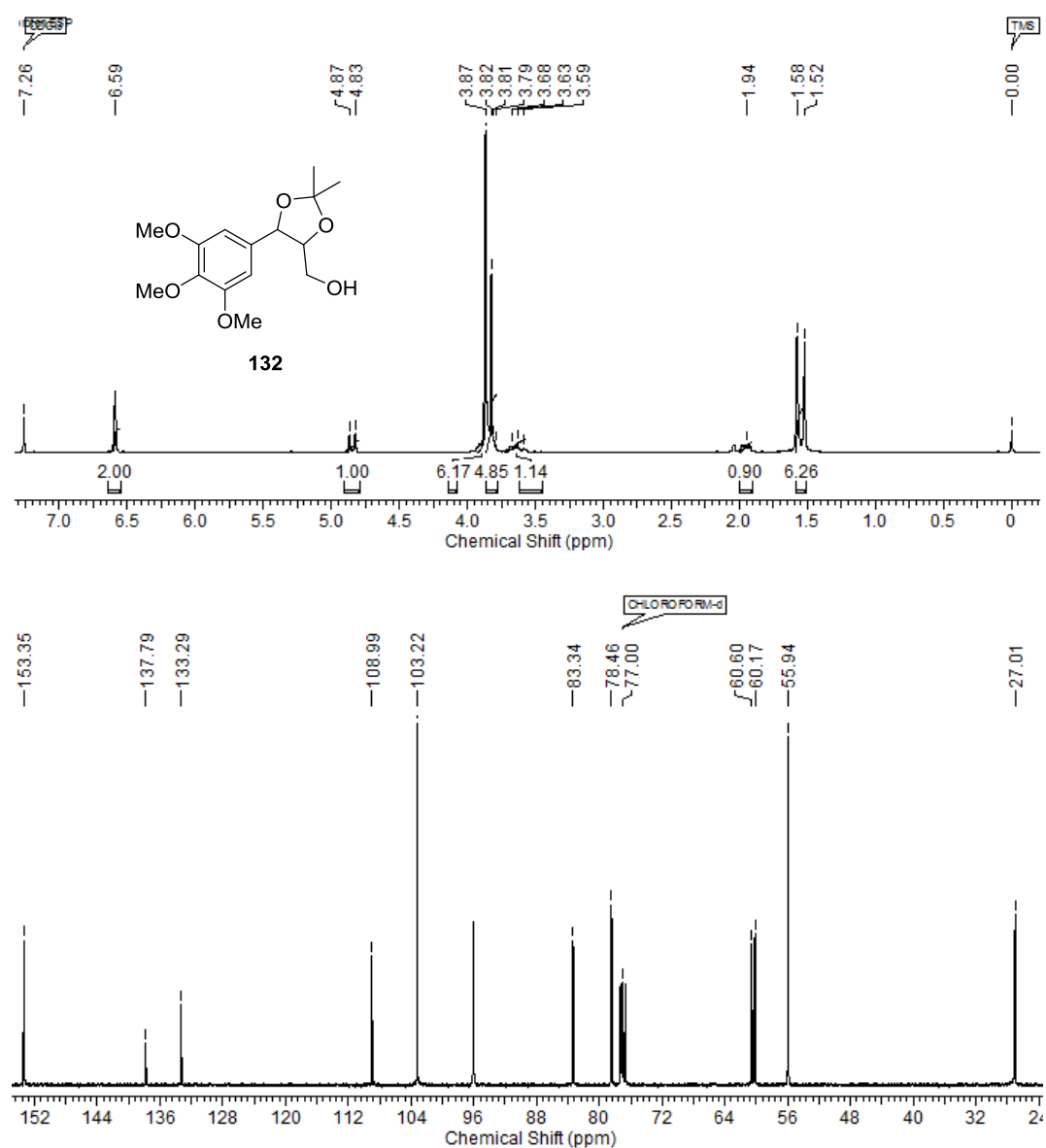


Fig. 20: ^1H and ^{13}C NMR spectra of alcohol **132**

The primary hydroxyl function in **132** was duly protected [*p*-TsCl, Et₃N, DMAP (10 mol%), CH₂Cl₂] as the tosylate **133**. Formation of tosylate **133** was confirmed by its ¹H NMR spectrum, which showed a characteristic singlet at δ 2.45 (s, 3H) and two doublets at δ 7.33 (d, *J* = 8.2 Hz, 2H) and 7.76 (d, *J* = 8.3 Hz, 2H) due to methyl and aromatic protons of tosyl group respectively, while its ¹³C NMR spectrum showed a typical carbon signal at δ 21.6 for methyl carbon of tosyl group (**Fig. 21**).

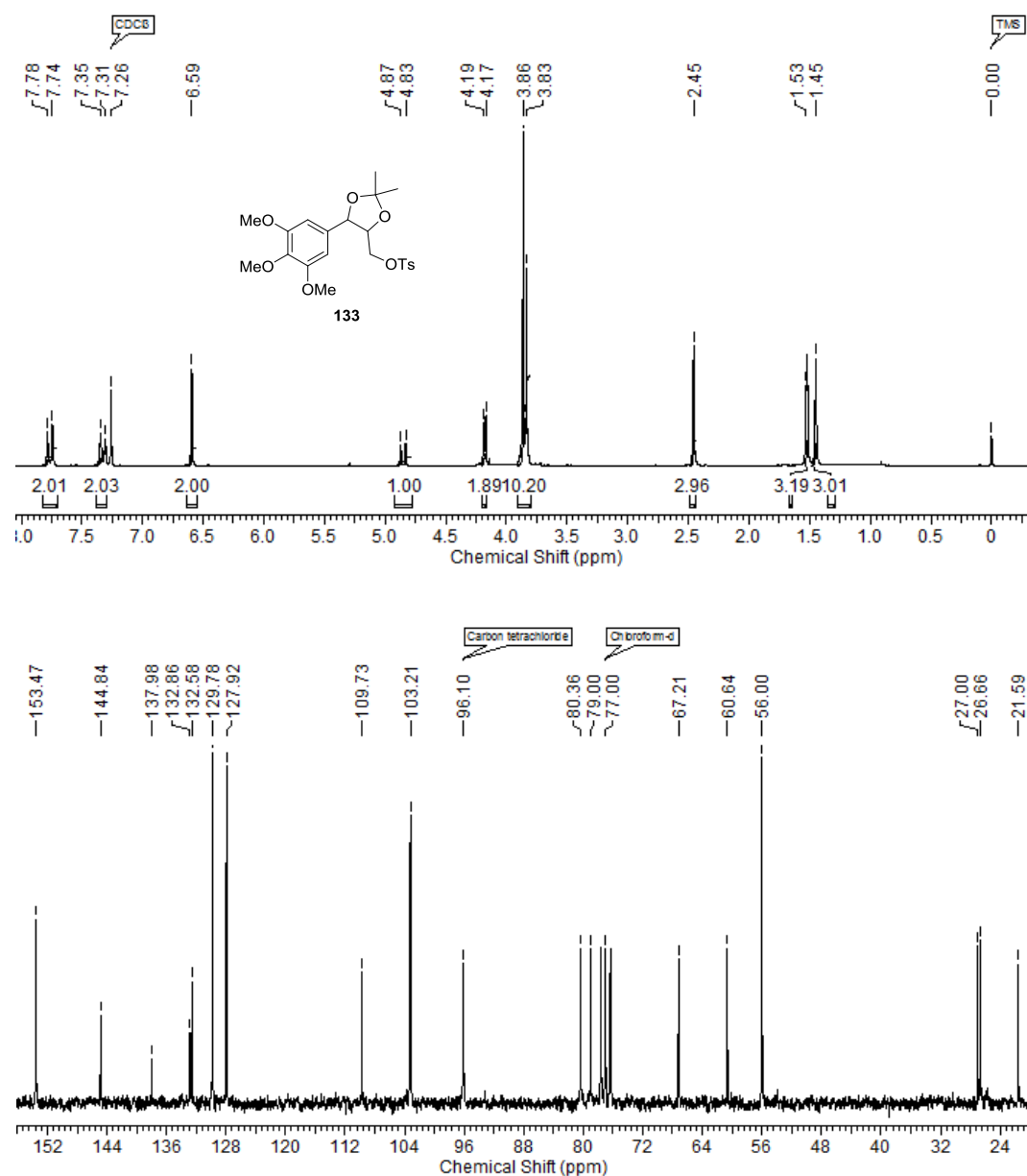


Fig. 21: ¹H and ¹³C NMR spectra of tosylate **133**

The selective deprotection of acetonide group was achieved using catalytic amount of camphor sulfonic acid in methanol to afford diol **134** in 96% yield.⁴⁹ The disappearance of proton singlets at δ 1.45 (s, 3H) and 1.53 (s, 3H) in the ^1H NMR spectrum of **134** corresponding to the methyl protons of isopropylidene group confirmed the removal of acetonide group (**Fig. 22**).

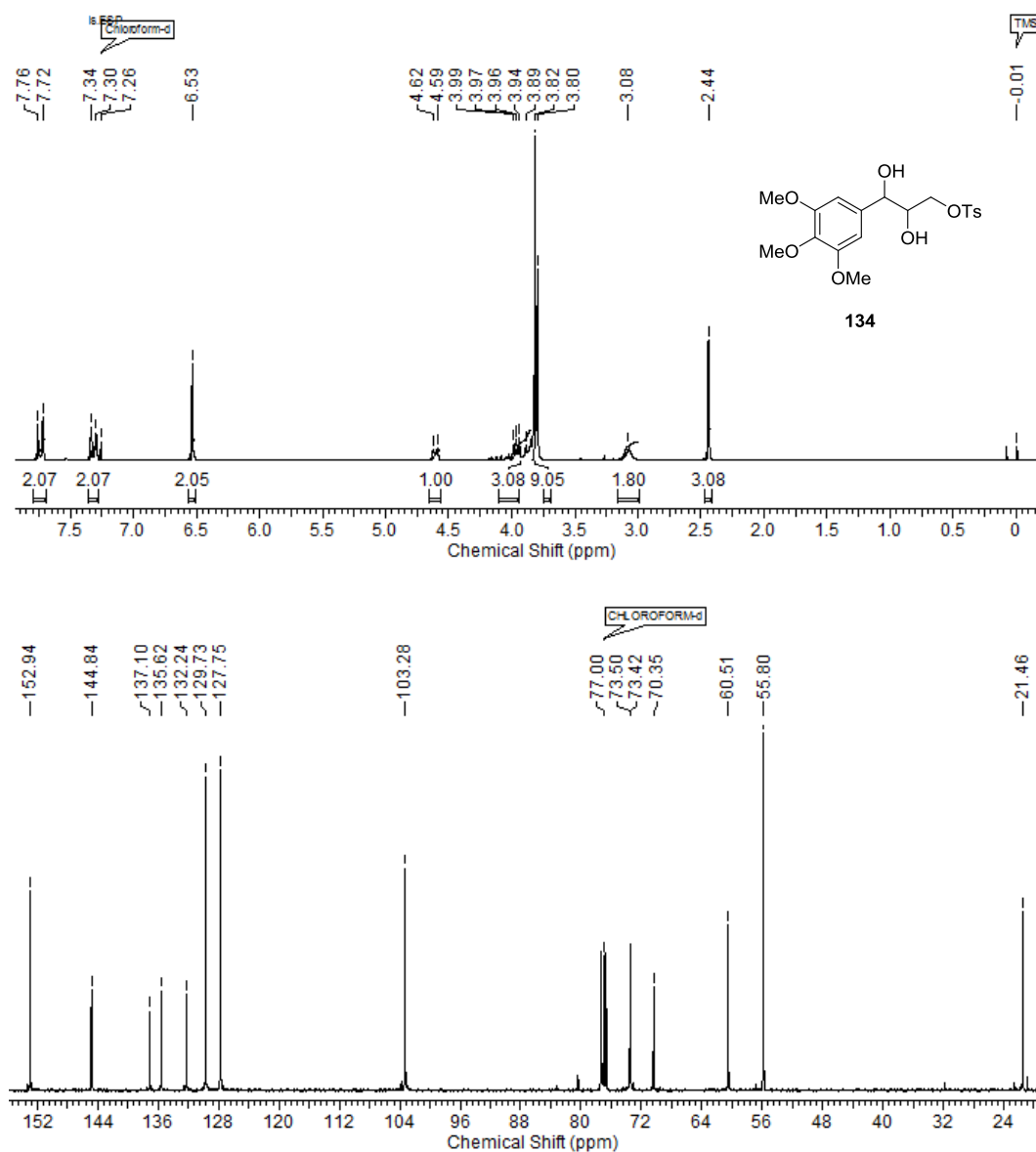


Fig. 22: ^1H and ^{13}C NMR spectra of diol **134**

Racemic *syn*-diol **134** was further converted to racemic *syn*-epoxide **135** in 88% yield, when carried out under basic conditions (K_2CO_3 , CH_3OH). The formation of racemic

syn-epoxide **135** was confirmed from its ^1H NMR spectrum, which showed typical proton signals at δ 2.48-2.52 (m, 1H) and 2.84 (t, $J = 4.4$ Hz, 2H), corresponding to methylene and methine protons of epoxide respectively. Its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 45.1 and 55.9 corresponding to methylene and methine carbons of the epoxide moiety respectively (**Fig. 23**).

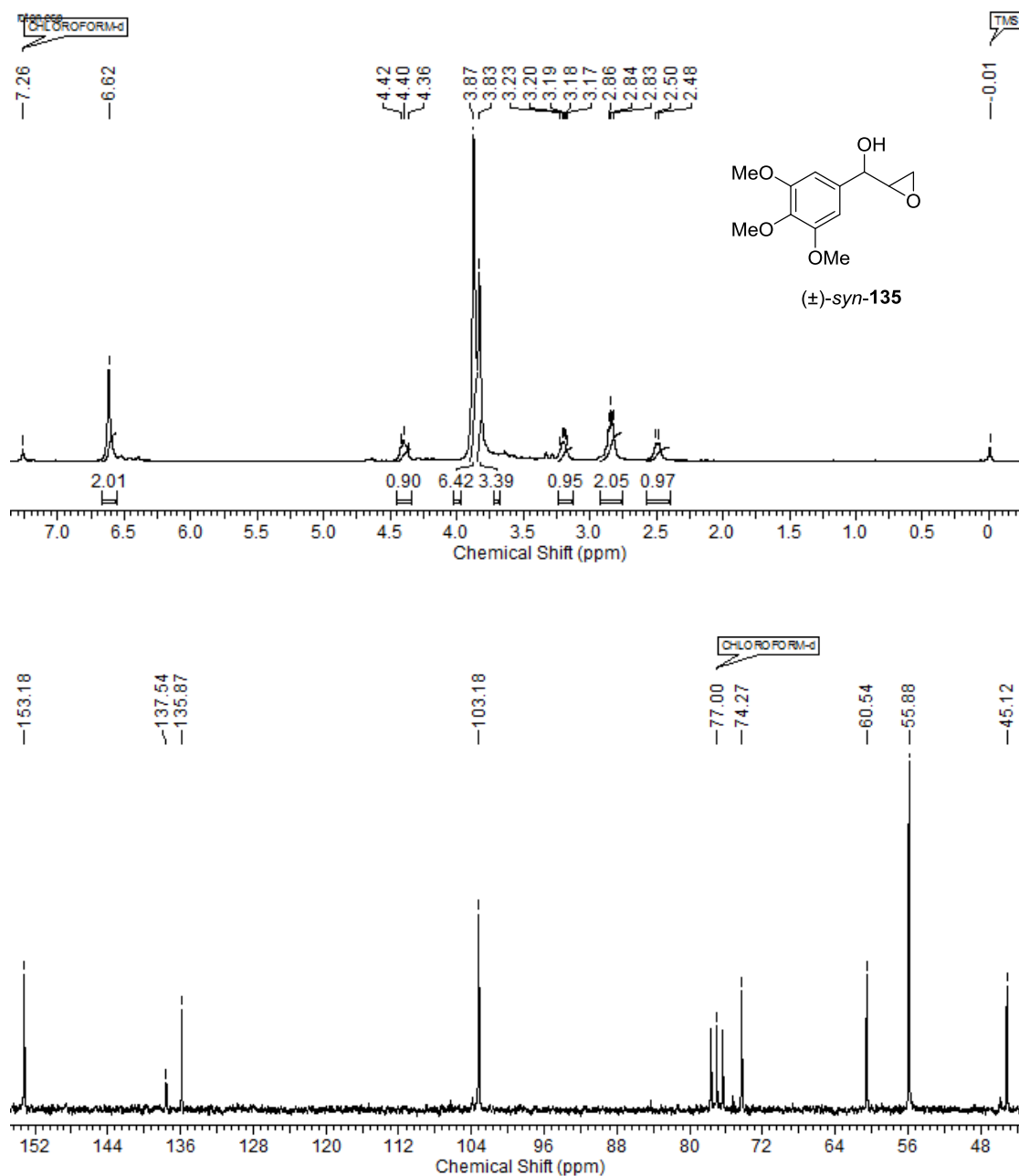


Fig. 23: ^1H and ^{13}C NMR spectra of (\pm)-*syn*-epoxide **135**

The benzylic hydroxyl group in **135** was then protected as its MOM ether **136** in 92% yield (MOMCl and diisopropylethylamine).⁵⁰ MOM protection in racemic *syn*-epoxide **136** was confirmed from its ¹H NMR spectrum, which showed typical proton signals at δ 4.59-4.71 (m, 2H) corresponding to the methylene protons of the MOM group, while its methyl protons appeared as a singlet at δ 3.39 (s, 3H). It was further confirmed by the appearance of carbon signals at δ 55.5 and 94.2 corresponding to the methyl and methylene carbons respectively of the MOM group in its ¹³C NMR spectrum (Fig. 24).

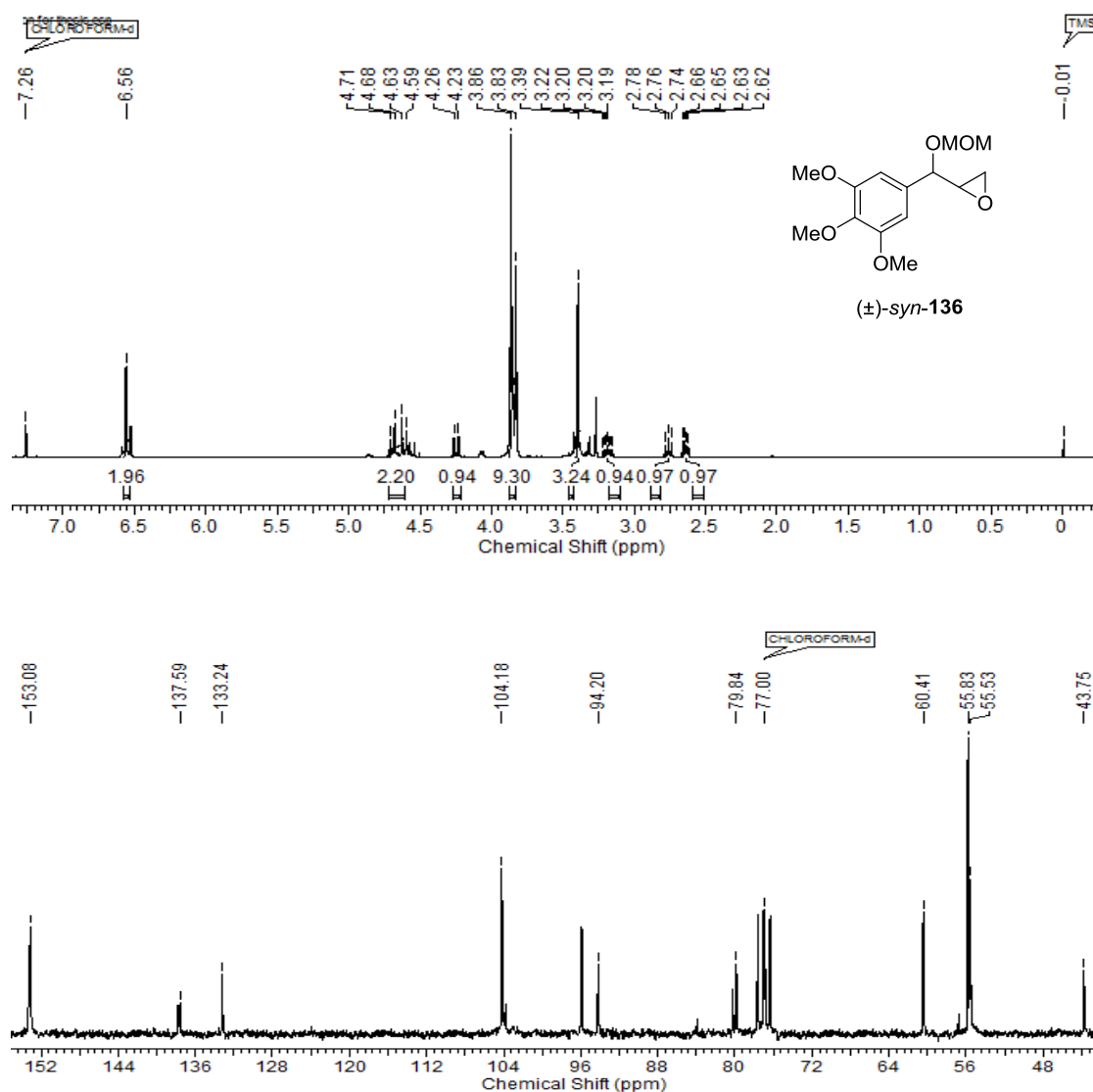
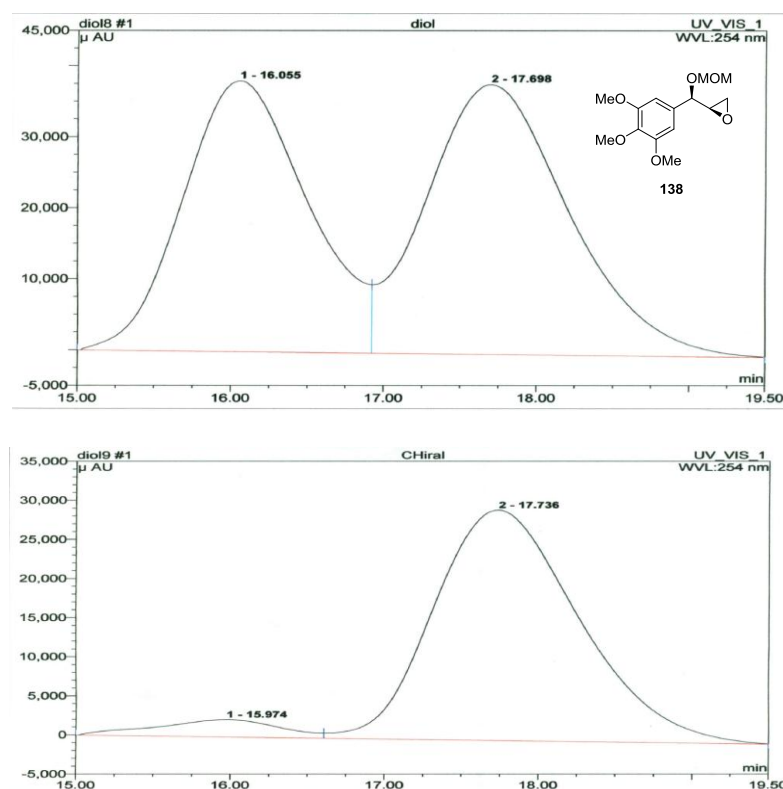


Fig. 24: ¹H and ¹³C NMR spectra of (\pm) -*syn*-epoxide **136**

Then, *syn*-3-(methoxymethyl)oxy-3-(3,4,5-trimethoxyphenyl)-1,2-epoxypropane **136** was subjected to two-stereocentered HKR with (*S,S*)-salen Co(OAc) complex^{20a,b} (0.5 mol%) and H₂O (0.5 equiv), which produced the corresponding chiral diol **137** (47%, 96% ee) and chiral *syn*-epoxide **138** (49%, 97% ee) in high optical purity. The chiral *syn*-epoxide **138** was readily separated from diol **137** by the column chromatographic purification. The optical purity of chiral *syn*-epoxide **138** was determined to be 97% ee from chiral HPLC analysis (Chiralcel OJ-H, *n*-hexane/ *i*PrOH, 90:10, 0.5 mL/min) retention time 15.97 (1.50%) and 17.73 (98.50%) (**Fig. 25**).



No	Ret. Time min	Height μ AU	Area μ AU* min	Rel. Area %
1	15.97	560.800	512.360	1.50
2	17.73	36825.916	33644.986	98.50

Fig. 25: HPLC chromatogram of *syn*-epoxide **138**

The regioselective reductive ring opening of chiral *syn*-epoxide **138** with LiAlH_4 ⁵¹ in THF furnished the secondary alcohol **139** in 96% yield. The ^1H and ^{13}C NMR spectra were in accordance with the proposed structure of **139**. In its ^1H NMR spectrum the newly generated methyl protons resonated at δ 1.54 as a doublet, while its typical carbon signal appeared at δ 18.4 in its ^{13}C NMR spectrum (Fig. 26). The IR spectrum of **139** displayed a characteristic strong absorption band at 3452 cm^{-1} indicating the presence of hydroxyl group.

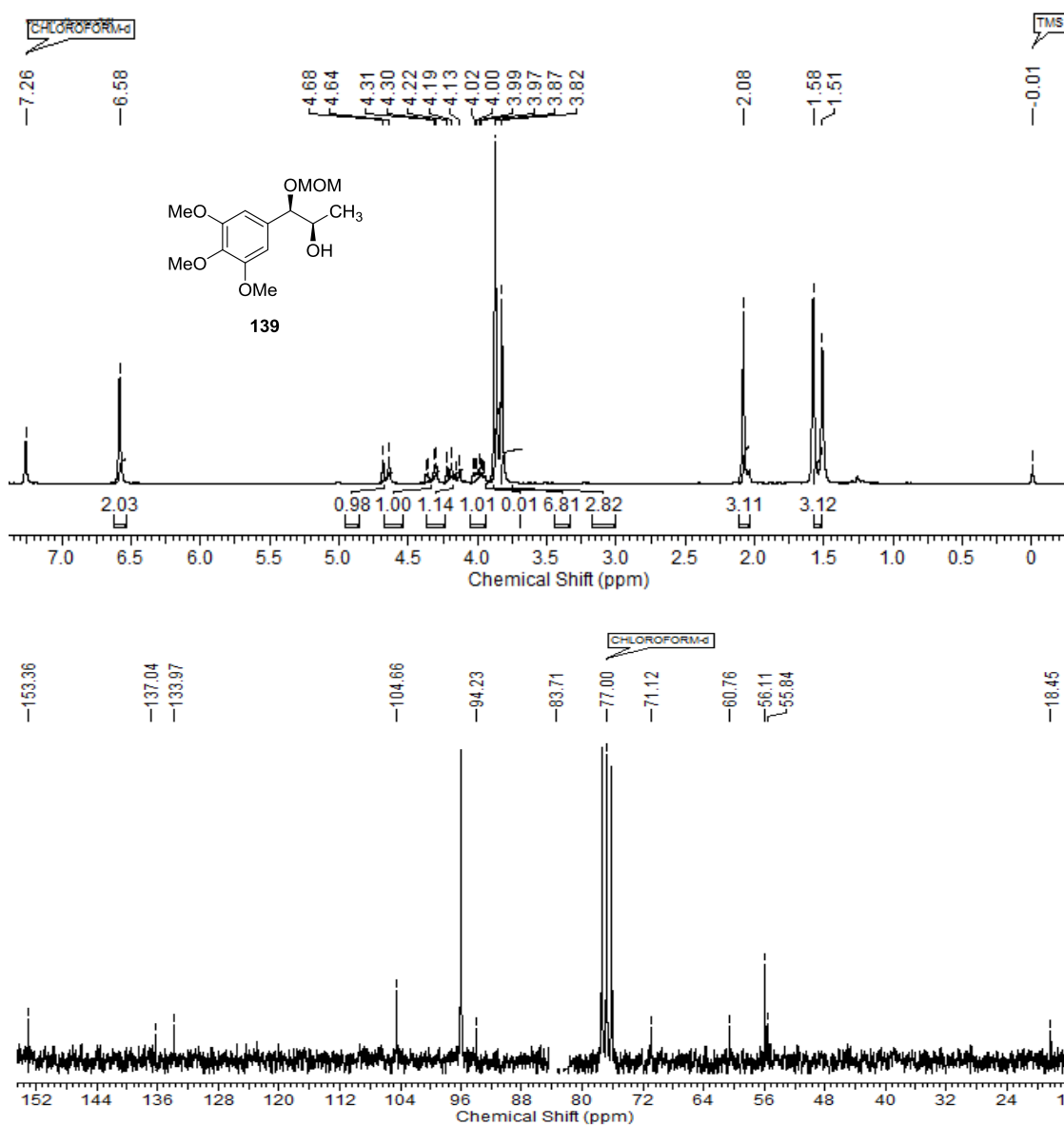
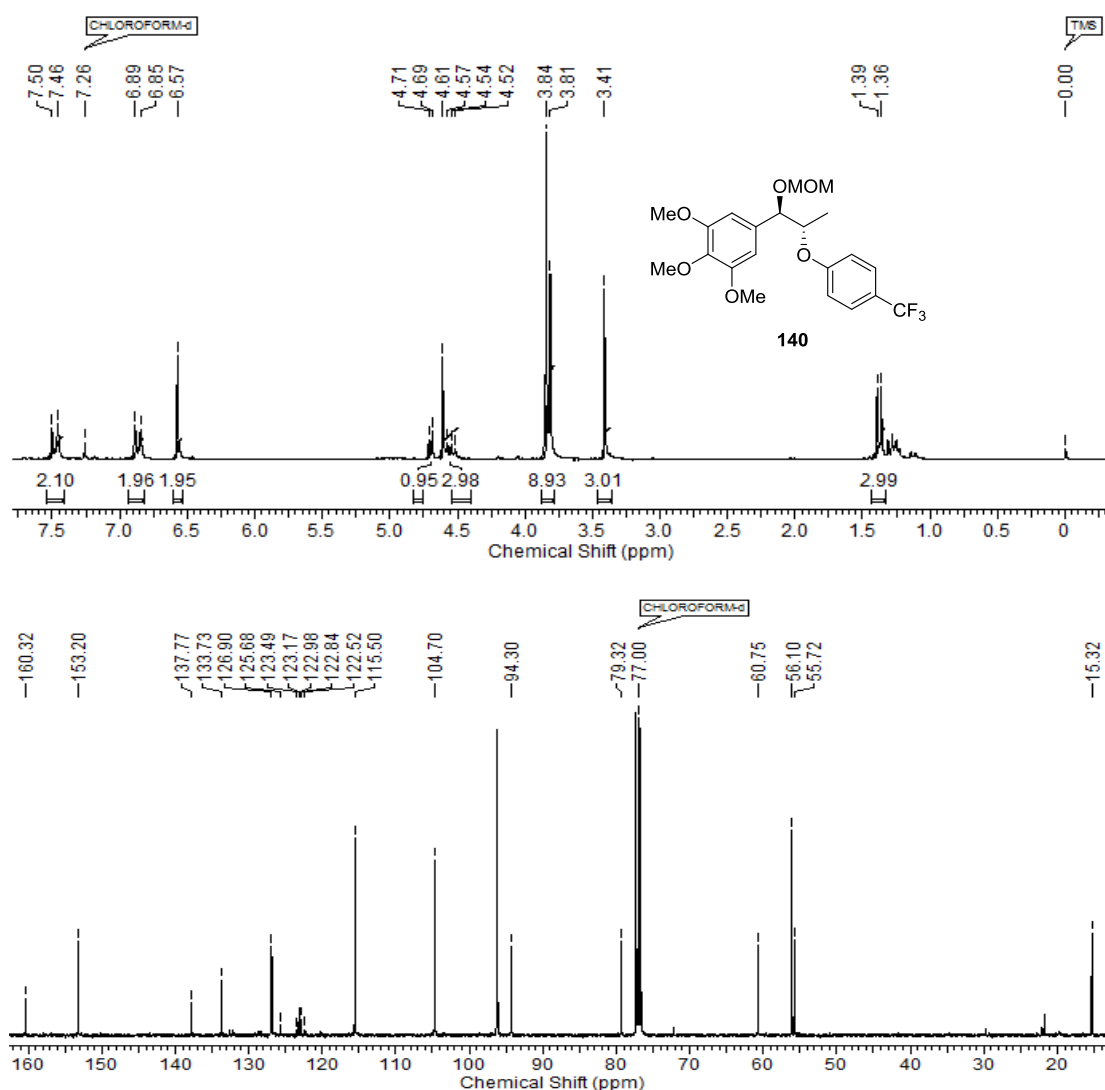


Fig. 26: ^1H and ^{13}C NMR spectra of alcohol **139**

Mitsunobu reaction of alcohol **139** with 4-(trifluoromethyl)phenol in the presence of PPh_3 and diisopropyl azodicarboxylate (DIAD) in THF at 70 °C afforded aryloxy ether **140** in 74% yield; $[\alpha]_D^{25} -46.20$ (c 1.0, CHCl_3). The ^1H NMR spectrum of the aryloxy ether **140** showed characteristic proton signal as doublets at δ 6.87 (d, $J = 8.6$ Hz, 2H) and 7.48 (d, $J = 8.6$ Hz, 2H) corresponding to the aromatic protons of 4-(trifluoromethyl)ether moiety. Its ^{13}C NMR spectrum showed characteristic carbon signals at δ 122.9 (q, $J = 32.3$ Hz) and 124.3 (d, $J = 271.2$ Hz) corresponding to the -C-CF₃ and -C-CF₃ carbons respectively (**Fig. 27**).



Finally, deprotection of the MOM protecting group with con. HCl in methanol furnished (-)-polysphorin analog (**110a**) in 86% yield and 97% ee; $[\alpha]_D^{25}$ -126.30 (c 1.0, CHCl_3). The ^1H NMR spectrum of **110a** showed typical proton signals at δ 1.25 (d, $J = 6.3$ Hz, 3H) and 2.64 (br s, 1H), which accounted for methyl and hydroxyl group protons respectively.

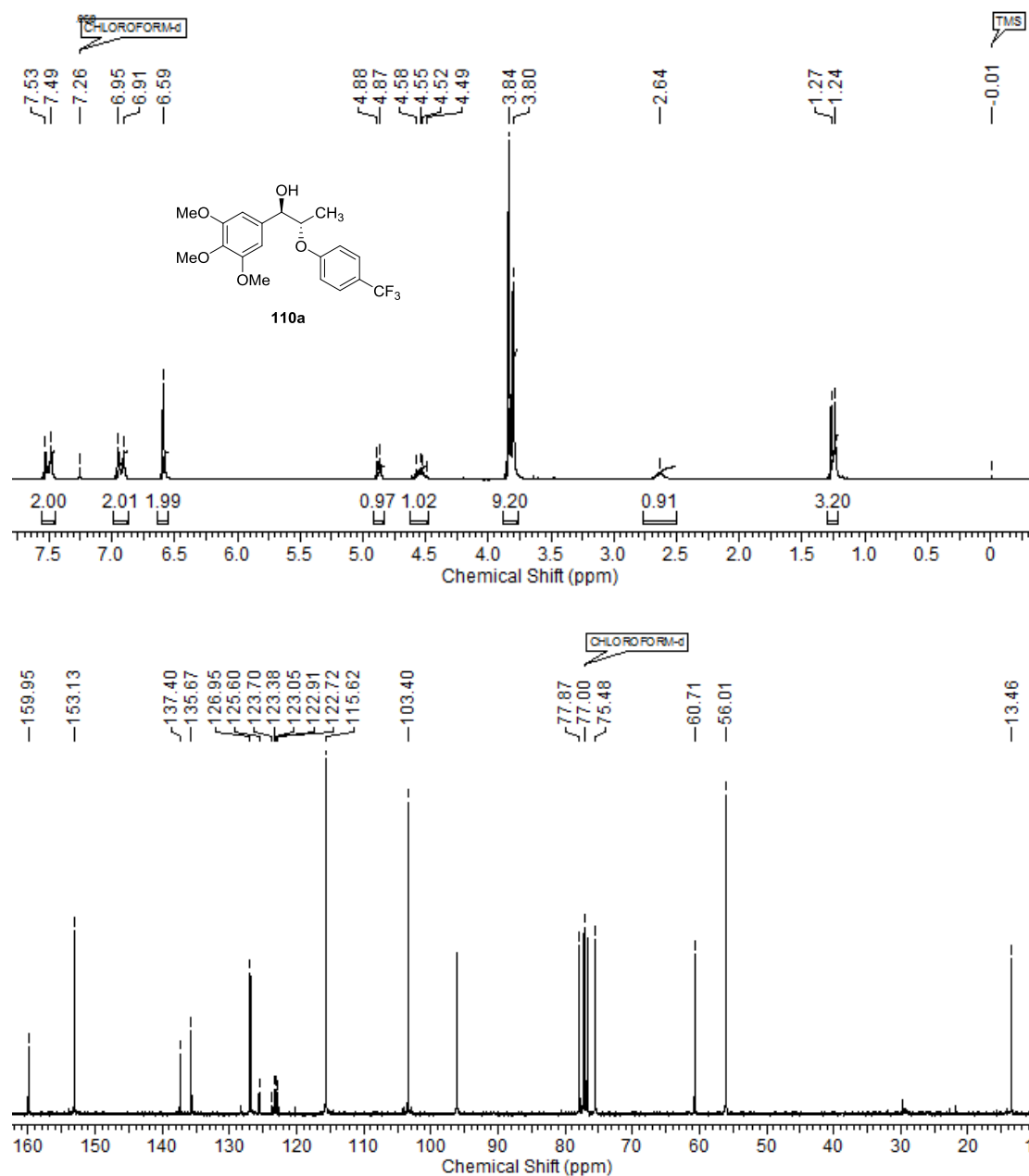


Fig. 28: ^1H and ^{13}C NMR spectra of (-)-polysphorin analog (**110a**)

The other signals at δ 6.93 (d, $J = 8.6$ Hz, 2H) and 7.51 (d, $J = 8.6$ Hz, 2H) were attributed to aromatic protons of 4-(trifluoromethyl)ether moiety. Its ^{13}C NMR spectrum showed characteristic carbon signals at δ 56.0, 60.7, 75.5, 77.8, 137.4, 153.1 and 159.9 for the carbons attached to oxygen atom (**Fig. 28**). The spectral data of (-)-polysphorin analog (**110a**) were in complete agreement with the reported values.⁴⁵

1.2.5 Conclusion

In conclusion, we have achieved the asymmetric synthesis of (-)-polysphorin analog (**110a**) (overall yield 18%, 97% ee) *via* late-stage two-stereocentered HKR of racemic *syn*-epoxide as the key step. The high enantiomeric excess obtained in this method render our approach a good alternative to the known methods. The present protocol provides for all four diastereomers of polysphorin analogs with a large diversity of aromatic units by suitably employing (*R, R*)-salen Co(OAc) complex for HKR and with various aromatic phenols for Mitsunobu reaction.

1.2.6 Experimental Section

Ethyl (*E*)-3-(3,4,5-trimethoxyphenyl)acrylate (**129**)

To a stirred solution of 3,4,5-trimethoxybenzaldehyde **128** (9.81 g, 50 mmol) in CH_2Cl_2 (100 mL), $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$ (19.25 g, 55 mmol) was added. It was then allowed to stir for 5 h under N_2 atmosphere. After the completion of reaction, CH_2Cl_2 was distilled out to give the crude product. Chromatographic purification of the crude product over silica gel with pet. ether:EtOAc (90:10) as eluent, afforded cinnamate ester **129** (12.78 g).

Yield: 96%, colorless solid; **mp:** 68-70 °C, (lit.⁵² **mp:** 67 °C); **IR** (CHCl_3 , cm^{-1}): ν_{max} 768, 980, 1039, 1176, 1203, 1270, 1312, 1367, 1450, 1639, 1705, 3063; **^1H NMR**

(200 MHz, CDCl₃): δ 1.33 (t, $J = 7.2$ Hz, 3H), 3.85 (s, 3H), 3.87 (s, 6H), 4.25 (q, $J = 7.2$ Hz, 2H), 6.31 (d, $J = 16$ Hz, 1H), 6.72 (s, 2H), 7.57 (d, $J = 16$ Hz, 1H); ¹³C NMR (50 MHz, CDCl₃): δ 14.2, 55.8, 60.1, 60.6, 105.0, 117.3, 129.8, 139.9, 144.3, 153.2, 166.5; **Analysis:** C₁₄H₁₈O₅ requires C, 63.15; H, 6.81; found: C, 63.02; H, 6.71%.

Ethyl 2,3-dihydroxy-3-(3,4,5-trimethoxyphenyl)propanoate (130)

Potassium osmate (0.16g, 0.45 mmol) and 50% aqueous *N*-methylmorpholine *N*-oxide (10.4 mL, 44.5 mmol) were added to a solution of cinnamate ester **129** (6 g, 22.5 mmol) in acetone (41.6 mL) at 0 °C. After continuous stirring for 20 h at 25 °C, the reaction mixture was diluted with CH₂Cl₂ (40 mL), and dried over anhyd. Na₂SO₄ and concentrated to give the crude diol, which was purified by column chromatography over silica gel using pet. ether:EtOAc (40:60) as eluent, afforded diol **130** (6.34 g).

Yield: 93%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 670, 762, 1039, 1182, 1217, 1326, 1732, 2362, 3020, 3451; **¹H NMR** (200 MHz, CDCl₃): δ 1.30 (t, $J = 7.3$ Hz, 3H), 2.84 (br s, 1H), 3.18-3.20 (m, 1H), 3.83 (s, 3H), 3.87 (s, 6H), 4.29 (q, $J = 7.3$ Hz, 2H), 4.34-4.36 (m, 1H), 4.93-4.96 (m, 1H), 6.64 (s, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.0, 56.0, 60.7, 62.0, 74.5, 74.8, 77.2, 103.2, 135.7, 137.3, 153.0, 172.6; **Analysis:** C₁₄H₂₀O₇ requires C, 55.99; H, 6.71; found: C, 55.87; H, 6.60%.

Ethyl 2,2-dimethyl-5-(3,4,5-trimethoxyphenyl)-1,3-dioxolane-4-carboxylate (131)

To a solution of diol **130** (2 g, 6.6 mmol) in dry CH₂Cl₂ was added 2,2-dimethoxy propane (8 mL) and camphor sulfonic acid (0.152 g, 0.3 mmol). The reaction mixture was stirred at 25 °C for 6 h. The reaction mixture was diluted with water, extracted with CH₂Cl₂ (3 × 50 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to afford the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (85:15) to furnish **131** (2.3 g).

Yield: 98%; colorless solid; **mp:** 57-59 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 682, 721, 834, 1006, 1132, 1235, 1381, 1462, 1595, 1753, 2990; **¹H NMR** (200 MHz, CDCl₃): δ 1.30 (t, $J = 7.2$ Hz, 3H), 1.55 (s, 3H), 1.61 (s, 3H), 3.84 (s, 3H), 3.87 (s, 8H), 4.36-4.38 (m, 1H), 5.13 (d, $J = 7.5$ Hz, 1H), 6.66 (s, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.2, 25.7, 26.9, 56.0, 60.7, 61.4, 80.5, 81.0, 103.3, 111.5, 133.3, 138.0, 153.4, 170.7; **Analysis:** C₁₇H₂₄O₇ requires C, 59.99; H, 7.11; found: C, 59.45; H, 6.99%.

(2,2-Dimethyl-5-(3,4,5-trimethoxyphenyl)-1,3-dioxolan-4-yl)methanol (132)

A solution of ester **131** (2 g, 5.8 mmol) in THF (10 mL) was added to a stirred slurry of lithium aluminium hydride (0.23 g, 5.8 mmol) in THF (30 mL). After being stirred for 1 h at 25 °C, the reaction was carefully quenched with EtOAc and water. The reaction mixture was then extracted with EtOAc (2 × 100 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by column chromatography using pet. ether:EtOAc (70:30) to give the alcohol **132** (1.62 g).

Yield: 92%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 670, 763, 1128, 1217, 1461, 1595, 2362, 3019, 3422; **¹H NMR** (200 MHz, CDCl₃): δ 1.52 (s, 3H), 1.58 (s, 3H), 1.94 (br s, 1H), 3.59-3.68 (m, 1H), 3.79-3.82 (m, 5H), 3.87 (s, 6H), 4.85 (d, $J = 8.4$ Hz, 1 H), 6.59 (s, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 27.0, 27.1, 55.9, 60.1, 60.6, 78.4, 83.4, 103.2, 109.0, 133.3, 137.8, 153.3; **Analysis:** C₁₅H₂₂O₆ requires C, 60.39; H, 7.43; found: C, 60.22; H, 7.28%.

(2,2-Dimethyl-5-(3,4,5-trimethoxyphenyl)-1,3-dioxolan-4-yl)methyl 4-methyl benzenesulfonate (133)

A solution of alcohol **132** (2 g, 6.7 mmol) in dry CH₂Cl₂ (50 mL) was treated with *p*-toluene sulfonyl chloride (1.39 g, 7.3 mmol), DMAP (0.081 g, 0.67 mmol) and Et₃N (2.3 mL, 16.7 mmol) at 25 °C. After being stirred for 4 h, the mixture was extracted with CH₂Cl₂ (3 × 50 mL), washed with water and the combined organic phases were

dried over anhyd. Na_2SO_4 and concentrated to give the crude tosylate, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (80:20) to give the tosylate **133** (2.84 g).

Yield: 94%; colorless gum; **IR** (CHCl_3 , cm^{-1}): ν_{max} 690, 762, 1129, 1224, 1631, 2371, 3019; **^1H NMR** (200 MHz, CDCl_3): δ 1.45 (s, 3H), 1.53 (s, 3H), 2.45 (s, 3H), 3.83 (s, 3H), 3.86 (s, 6H), 4.06-4.19 (m, 3H); 4.85 (d, $J = 8.3$ Hz, 1H), 6.59 (s, 2H), 7.33 (d, $J = 8.2$ Hz, 2H), 7.76 (d, $J = 8.3$ Hz, 2H); **^{13}C NMR** (50 MHz, CDCl_3): δ 21.6, 26.7, 27.0, 56.0, 60.6, 67.2, 79.0, 80.4, 103.2, 109.7, 127.9, 129.8, 132.6, 132.9, 138.0, 144.9, 153.5; **Analysis:** $\text{C}_{22}\text{H}_{28}\text{O}_8\text{S}$ requires C, 58.39; H, 6.24; found: C, 58.31; H, 6.04%.

2,3-Dihydroxy-3-(3,4,5-trimethoxyphenyl)propyl-4-methylbenzenesulfonate (134)

To a solution of tosylate **133** (2.84 g, 6.27 mmol) in CH_3OH (40 mL) was added camphor sulfonic acid (0.317 g, 0.62 mmol) and stirred for 1 h at 40 °C. Solvent was evaporated and the residue extracted with EtOAc (3×30 mL) and the combined organic phases were dried over anhyd. Na_2SO_4 and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (50:50) to give diol **134** (2.48 g).

Yield: 96%; colourless gum; **IR** (CHCl_3 , cm^{-1}): ν_{max} 688, 771, 1130, 1216, 1470, 1631, 2368, 3028, 3432; **^1H NMR** (200 MHz, CDCl_3): δ 2.44 (s, 3H), 3.08 (brs, 1H), 3.80 (s, 3H), 3.82 (s, 6H), 3.88-3.89 (m, 3H); 4.61 (d, $J = 6.2$ Hz, 1H), 6.53 (s, 2H), 7.32 (d, $J = 8.3$ Hz, 2H), 7.74 (d, $J = 8.3$ Hz, 2H); **^{13}C NMR** (50 MHz, CDCl_3): δ 21.5, 55.8, 60.5, 70.3, 73.4, 73.5, 103.3, 127.7, 129.7, 132.2, 135.6, 137.1, 144.8, 152.9; **Analysis:** $\text{C}_{19}\text{H}_{24}\text{O}_8\text{S}$ requires C, 55.33; H, 5.87; found: C, 55.16; H, 5.72%.

Oxiran-2-yl (3,4,5-trimethoxyphenyl)methanol (135)

To a stirred solution of **134** (2.48 g, 6.02 mmol) in CH₃OH (30 mL) was added anhydrous K₂CO₃ (1.62 g, 12.04 mmol) at 0 °C. The mixture was then warmed up and stirred at 25 °C. After the reaction was complete (monitored by TLC), 50 mL of aq. NaHCO₃ was added and the reaction mixture was extracted with EtOAc (3 × 60 mL). The combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by flash column chromatography using pet. ether:EtOAc (65:35) to give epoxy alcohol, **135** (1.27 g).

Yield: 88%; colorless oil; **IR** (CHCl₃, cm⁻¹): ν_{\max} 690, 748, 1109, 1690, 2408, 2945, 3040, 3420; **¹H NMR** (200 MHz, CHCl₃): δ 2.48-2.52 (m, 1H), 2.84 (t, $J = 4.4$ Hz, 2H), 3.17-3.23 (m, 1H), 3.83 (s, 3H), 3.86 (s, 6H), 4.36-4.42 (m, 1H), 6.62 (s, 2H); **¹³C NMR** (50 MHz, CHCl₃): δ 45.1, 55.9, 60.5, 74.3, 103.2, 135.9, 137.5, 153.2; **Analysis:** C₁₂H₁₆O₅ requires C, 59.99; H, 6.71; found: C, 59.86; H, 6.64%.

2-[(Methoxymethoxy)(3,4,5-trimethoxyphenyl)methyl]oxirane (136)

To a solution of the epoxy alcohol **135** (1.00 g, 4.1 mmol) and diisopropylethylamine (3.60 mL, 20.8 mmol) in dry CH₂Cl₂ (20 mL) was added methoxymethyl chloride (0.80 mL, 10.2 mmol) under nitrogen over 5 min at 0 °C, and the mixture was allowed to warm to 25 °C and stirred for 8 h. After cooling to 0 °C, the reaction mixture was quenched with water and extracted with CH₂Cl₂ (3 × 20 mL). The combined organic extracts were washed with water (3 × 50 mL) and brine, dried over anhyd. Na₂SO₄ and concentrated under reduced pressure. Silica gel column chromatographic purification of the crude product using pet. ether:EtOAc (70:30) gave *syn*-epoxide **136** (1.15 g).

Yield: 92%; colorless oil; **IR** (CHCl₃, cm⁻¹): ν_{\max} 672, 760, 1102, 1209, 1677, 2361, 2930, 3026, 3633; **¹H NMR** (200 MHz, CHCl₃): δ 2.64 (dd, $J = 2.6, 4.9$ Hz, 1H),

2.76 (t, $J = 4.4$ Hz, 1H), 3.15-3.22 (m, 1H), 3.39 (s, 3H), 3.83 (s, 3H), 3.86 (s, 6H), 4.24 (d, $J = 6.3$ Hz, 1H), 4.59-4.71 (m, 2H), 6.56 (s, 2H); ^{13}C NMR (50 MHz, CHCl_3): δ 43.7, 55.5, 55.8, 60.4, 79.8, 94.2, 104.2, 133.2, 137.6, 153.1; **Analysis:** $\text{C}_{14}\text{H}_{20}\text{O}_6$ requires C, 59.15; H, 7.09; found: C, 59.04; H, 7.16%.

(2*S*, 3*S*)-3-(Methoxymethoxy)-3-(3,4,5-trimethoxyphenyl)propane-1,2-diol (137)

To a solution of (*S*, *S*)-salen Co(II) complex (purchased from Aldrich, USA.) (0.043 g, 0.07 mmol) in toluene (2.0 mL) was added gl. AcOH (0.04 g, 7.3 mmol). It was allowed to stir at 25 °C in air for 30 min over which time the color was changed from orange- red to a dark brown. It was then concentrated *in vacuo* to get (salen)Co(III)(OAc) complex as brown colored solid. To a stirred solution of (salen)Co(III)(OAc) complex (0.004 g, 0.5 mol%) and racemic *syn*-epoxide **136** (1.41 mmol) in THF (0.5 mL) at 0 °C was added H_2O (0.012 g, 0.7 mmol) drop-wise over 5 min. The reaction mixture was allowed to warm to 25 °C and stirred for 14 h. After completion of reaction (monitored by TLC), solvent was removed *in vacuo*. The crude product was purified by column chromatography over silica gel to (2*R*, 3*R*)-3-(methoxymethoxy)-3-(3,4,5-trimethoxy phenyl)-1,2 epoxy propane (**138**) on eluting with pet. ether:EtOAc (70:30) and (2*S*, 3*S*)-3-(methoxymethoxy)-3-(3,4,5-trimethoxyphenyl)propane-1,2-diol (**137**) with pet. ether:EtOAc (55:45) in high enantiomeric excess.

The chiral diol **137** was obtained as a colorless liquid (47% and 96% ee); [%ee was determined by transforming diol **137** to the corresponding *syn*-epoxide, [(i) TsCl , Bu_2SnO (2 mol%), Et_3N , DMAP (10 mol%), CH_2Cl_2 , 3 h; (ii) K_2CO_3 , MeOH, 1 h, 83% (over two steps) and comparing its specific rotation with its antipode **138**];

Yield: 47%; colorless liquid; $[\alpha]_{25}^{\text{D}} +71.55$ (c 1, CH_3OH); **IR** (CHCl_3 , cm^{-1}): ν_{max} 670, 748, 840, 1075, 1275, 1278, 1385, 1494, 1644, 2920, 3018, 3088, 3458; ^1H

NMR (200 MHz, CHCl₃): δ 2.64 (dd, $J = 2.6, 4.9$ Hz, 1H), 2.76 (t, $J = 4.4$ Hz, 1H), 3.15-3.22 (m, 1H), 3.39 (s, 3H), 3.83 (s, 3H), 3.86(s, 6H), 4.24 (d, $J = 6.3$ Hz, 1H), 4.59-4.71 (m, 2H), 6.56 (s, 2H); **¹³C NMR** (50 MHz, CHCl₃): δ 55.5, 55.8, 60.4, 64.0, 65.1, 85.2, 94.4, 103.8, 133.6, 137.2, 153.3; **Analysis** for C₁₄H₂₂O₇ requires: C, 55.62; H, 7.34; found: C, 55.51; H, 7.21%.

(2R, 3R)-3-(Methoxymethoxy)-3-(3,4,5-trimethoxyphenyl-1,2-epoxypropane (138)

Yield: 49%; colorless oil; $[\alpha]_{25}^D -72.30$ (c 1.16, CH₃OH); 97% ee by chiral HPLC analysis (Chiralcel OJ-H, *n*-hexane/ *i*PrOH, 90:10, 0.5 mL/min) retention time 15.97 (1.50%) and 17.73 (98.50%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 672, 760, 1102, 1209, 1677, 2361, 2930, 3026, 3633; **¹H NMR** (200 MHz, CHCl₃): δ 2.64 (dd, $J = 2.6, 4.9$ Hz, 1H), 2.76 (t, $J = 4.4$ Hz, 1H), 3.15-3.22 (m, 1H), 3.39 (s, 3H), 3.83 (s, 3H), 3.86 (s, 6H), 4.24 (d, $J = 6.3$ Hz, 1H), 4.59-4.71 (m, 2H), 6.56 (s, 2H); **¹³C NMR** (50 MHz, CHCl₃): δ 43.7, 55.5, 55.8, 60.4, 79.8, 94.2, 104.2, 133.2, 137.6, 153.1; **Analysis:** C₁₄H₂₀O₆ requires C, 59.15; H, 7.09; found: C, 59.06; H, 7.02%.

(1R, 2R)-1-(Methoxymethoxy)-1-(3,4,5-trimethoxyphenyl)propan-2-ol (139)

A solution of epoxide **138** (1.00 g, 3.5 mmol) in THF (30 mL) was added to a stirred slurry of lithium aluminium hydride (0.167 g, 4.4 mmol). After being stirred for 4 h at 25 °C, the reaction was carefully quenched with water. It was then extracted with EtOAc (3 × 50 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by flash column chromatography using pet. ether:EtOAc (60:40) to give the alcohol **139** (0.967 g).

Yield: 96%; colorless oil; $[\alpha]_{25}^D -116.42$ (c 1.1, CH₃OH); **IR** (CHCl₃, cm⁻¹): ν_{\max} 690, 762, 1031, 1129, 1217, 1461, 1593, 2363, 3019, 3452; **¹H NMR** (200 MHz, CDCl₃): δ 1.54 (d, $J = 13.2$ Hz, 3H), 2.08 (s, 3H), 3.82 (s, 3H), 3.87 (s, 7H), 3.97-4.03 (m, 1H), 4.13-4.22 (m, 1H), 4.30-4.37 (m, 1H), 4.66 (d, $J = 8.6$ Hz, 1H), 6.58 (s, 2H); **¹³C**

NMR (50 MHz, CDCl₃): δ 18.4, 55.8, 56.1, 60.8, 71.1, 83.7, 94.2, 104.7, 134.0, 137.0, 153.4; **Analysis:** C₁₄H₂₂O₆ requires C, 58.73; H, 7.74; found: C, 58.62; H, 7.61%.

1,2,3-Trimethoxy-5-[(1R, 2S)-1-(methoxymethoxy)-2-(4-(trifluoromethyl)phenoxy)propyl]benzene (140)

To a stirred solution of chiral alcohol **139** (0.100 g, 0.35 mmol), PPh₃ (0.183 g, 0.7 mmol) and diisopropylazodicarboxylate (0.120 g, 0.6 mmol) in THF (5 mL) at 0 °C was added 4-(trifluoromethyl)phenol (0.113 g, 0.7 mmol). The resulting solution was stirred for 12 h at 70 °C and then concentrated *in vacuo*. The crude compound was purified by column chromatography using pet. ether:EtOAc (80:20) to afford the aryloxy ether **140** (0.112 g).

Yield: 74%; gum; $[\alpha]_D^{25}$ -46.20 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 891, 1052, 1097, 1130, 1190, 1325, 1374, 1485, 1629, 1765, 2928, 3015; **¹H NMR** (200 MHz, CDCl₃): δ 1.37 (d, *J* = 6.2 Hz, 3H), 3.41 (s, 3H), 3.81 (s, 3H), 3.84 (s, 6H), 4.52-4.69 (m, 3H), 4.70 (d, *J* = 4.8 Hz, 1H), 6.57 (s, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 7.48 (d, *J* = 8.6 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 15.3, 55.7, 56.1, 60.7, 79.3, 94.3, 104.7, 115.5, 122.9 (q, *J* = 32.3 Hz), 124.3 (d, *J* = 271.2 Hz), 126.9, 133.7, 137.8, 153.2, 160.3; **Analysis:** C₂₁H₂₅F₃O₆ requires C, 58.60; H, 5.85; found: C, 58.48; H, 5.73%.

(1R, 2S)-2-(4-(Trifluoromethyl)phenoxy)-1-(3,4,5-trimethoxyphenyl)propan-1-ol: [(-)-Polysphorin analog] (110a)

To a stirred solution of the aryloxy ether **140** (0.100 g, 0.23 mmol) in CH₃OH (5 mL) was added con. HCl (2 mL) and stirred for 4 h and then extracted with EtOAc (3 × 10 mL), washed with brine, dried over anhyd. Na₂SO₄ and concentrated under reduced

pressure. Silica gel column chromatographic purification of the crude product using petroleum ether:EtOAc (60:40) gave (-)-polysphorin analog **110a** (76 mg).

Yield: 86%; gum; $[\alpha]_D^{25}$ -126.30 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 775, 865, 1019, 1068, 1327, 1413, 1440, 2957, 3471; **¹H NMR** (200 MHz, CDCl₃): δ 1.25 (d, *J* = 6.3 Hz, 3H), 2.64 (br s, 1H), 3.80 (s, 3H), 3.84 (s, 6H), 4.49-4.58 (m, 1H), 4.87 (d, *J* = 3.9 Hz, 1H), 6.59 (s, 2H), 6.93 (d, *J* = 8.6 Hz, 2H), 7.51 (d, *J* = 8.6 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.5, 56.0, 60.7, 75.5, 77.8, 103.4, 115.6, 123.2 (q, *J* = 32.6 Hz), 123.8 (d, *J* = 270.3 Hz), 126.9, 135.7, 137.4, 153.1, 159.9; **Analysis:** C₁₉H₂₁F₃O₅ requires C, 59.07; H, 5.48; found: C, 59.14; H, 5.29%.

1.2.7 References

1. Jewers, K.; Davis, J. B.; Dougan, J.; Manchanda, A. H.; Blunden, G.; Aye, K.; Wetchapinan, S. *Phytochemistry* **1972**, *11*, 2025.
2. Wu, Y. C.; Duh, C. Y.; Chang, F. R.; Chang, G. Y.; Wang, S. K.; Chang, J. J.; McPhail, A. T.; Lee, K. H. *J. Nat. Prod.* **1991**, *54*, 1077.
3. Sam, T.; Sew-Yeu, C.; Matsjeh, S.; Gan, E. K.; Razak, D.; Mohamed, A. L. *Tetrahedron Lett.* **1987**, *28*, 2541.
4. (a) Talapatra, S. K.; Basu, D.; Deb, T.; Goswami, S.; Talapatra, B. *Indian J. Chem. Sect. B*, **1985**, *24B*, 29; (b) Lan, Y. H.; Chang, F. R.; Liaw, C. C.; Wu, C. C.; Chiang, M. Y.; Wu, Y. C. *Planta Med.* **2005**, *71*, 153.
5. (a) Tsubuki, M.; Kanai, K.; Honda, T. *J. Chem. Soc., Chem. Commun.* **1992**, 1640; (b) Tsubuki, M.; Kanai, K.; Nagase, H.; Honda, T. *Tetrahedron* **1999**, *55*, 2493.
6. Fang, X. P.; Anderson, J. E.; Chang, C. J.; Mc Laughlin, J. L. *J. Nat. Prod.* **1991**, *54*, 1034.
7. Surivet, J. P.; Gore, J.; Vatele, J. M. *Tetrahedron Lett.* **1996**, *37*, 371.
8. Mukai, C.; Hirai, S.; Hanaoka, M. *J. Org. Chem.* **1997**, *2*, 6619.
9. Surivet, J. P.; Vatele, J. M. *Tetrahedron Lett.* **1998**, *39*, 7299.
10. Chen, J.; Lin, G. Q.; Wang, Z. M.; Liu, H. Q. *Synthesis* **2002**, 1265.
11. Ramachandran, P. V.; Chandra, J. S.; Reddy, M. V. R. *J. Org. Chem.* **2002**, *67*, 7547.
12. Tate, E. W.; Dixon, D. J.; Ley, S. V. *Org. Biomol. Chem.* **2006**, *4*, 1698.
13. Prasad, K. R.; Gholap, S. L. *Tetrahedron Lett.* **2007**, *48*, 4679.
14. Yoshida, T.; Yamauchi, S.; Tago, R.; Maruyama, M.; Akiyama, K.; Sugahara, T.; Kishida, T.; Koba, Y. *Biosci. Biotechnol. Biochem.* **2008**, *72*, 2342.
15. Yadav, J. S.; Premalatha, K.; Harshavardhan, S. J.; Reddy, B. V. S. *Tetrahedron Lett.* **2008**, *49*,

6765.

16. Sabitha, G.; Reddy, T. R.; Yadav, J. S. *Tetrahedron Lett.* **2011**, *52*, 6550.
17. Paioti, P. H. S.; Coelho, F. *Tetrahedron Lett.* **2011**, *52*, 6180.
18. Sabitha, G.; Bhikshapathi, M.; Ranjith, N.; Ashwini, N.; Yadav, J. S. *Synthesis* **2011**, 821.
19. Shing, T. K. M.; Tsui, H. C.; Zhou, Z. H. *J. Org. Chem.* **1995**, *60*, 3121.
20. (a) Reddy, R. S.; Chouthaiwale, P. V.; Suryavanshi, G.; Chavan .V. B.; Sudalai. A. *Chem. Commun.* **2010**, *46*, 5012; (b) Tokunaga, M.; Larrow, J. F.; Kakiuchi, F.; Jacobsen, E. N. *Science* **1997**, *277*, 936.
21. (a) Wilkinson, R. G.; Shepherd, R. G.; Thomas, J. P.; Baughn, C. *J. Am. Chem. Soc.* **1961**, *83*, 2212; (b) Shepherd, R. G.; Wilkinson, R. G. *J. Med. Chem.* **1962**, *5*, 823; (c) Wilkinson, R. G.; Cantrall, M. B.; Shepherd, R. G. *J. Med. Chem.* **1962**, *5*, 835.
22. Some, among many, notable examples: (a) Fumagillin: Tarbell, D. S.; Carman, R. M.; Chapman, D. D.; Cremer, S. E.; Cross, A. D.; Huffman, K. R.; Kuntsmann, M.; McCorkindale, N. J.; McNally, J. G.; Rosowsky, A.; Varino, F. H. L.; West, R. L. *J. Am. Chem. Soc.* **1961**, *83*, 3096; (b) Ovalicin: Sigg, H. P.; Weber, H. P. *Helv. Chim. Acta* **1968**, *51*, 1395; (c) Coriolin: Takeuchi, T.; Iinuma, H.; Iwanaga, J.; Takahashi, S.; Takita, T.; Umezawa, H. *J. Antibiot.* **1969**, *22*, 215; (d) Disparlure: Bierl, B. A.; Beroza, M.; Collier, C. W. *Science* **1970**, *170*, 87.
23. For reviews and lead references, see: (a) Winstein, S.; Henderson, R. B. *Heterocyclic Compounds*, Vol. 1; Elderfield, R. C., Ed.; Wiley: New York, 1950; Chapter 1; (b) Parker, R. E.; Isaacs, N. S. *Chem. Rev.* **1959**, *59*, 737; (c) Bartok, M.; Lang, K. L. *Small Ring Heterocycles In The Chemistry of Heterocyclic Compounds*, Vol. 42, Part 3; Hassner, A., Ed.; Wiley: New York, **1985**; Chapter 1.
24. Furrow, M. E.; Schaus, S. E.; Jacobsen, E. N. *J. Org. Chem.* **1998**, *63*, 6776.
25. For an in-depth discussion of practical considerations in kinetic resolution reactions, see: Keith, J. M.; Larrow, J. F.; Jacobsen, E. N. *Adv. Synth. Catal.* **2001**, *343*, 5.
26. Schaus, S. E.; Brandes, B. D.; Larrow, J. F.; Tokunaga, M.; Hansen, K. B.; Gould, A. E.; Furrow, M. E.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2002**, *124*, 1307.
27. Ford, D. D.; Nielsen, L. P. C.; Zuend, S. J.; Musgrave, C. B.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2013**, *135*, 15595.
28. (a) Nielsen, L. P. C.; Stevenson, C. P.; Blackmond, D. G.; Jacobsen, E. N. *J. Am. Chem. Soc.* **2004**, *126*, 1360; (b) Nielsen, L. P. C.; Zuend, S. J.; Ford, D. D.; Jacobsen, E. N. *J. Org. Chem.* **2012**, *77*, 2486; (c) Jain, S.; Zheng, X.; Jones, C. W.; Weck, M.; Davis, R. *J. Inorg. Chem.* **2007**, *46*, 8887; (d) Jain, S.; Venkatasubbaiah, K.; Jones, C. W.; Davis, R. *J. Mol. Catal. A: Chem.* **2010**, *316*, 8.
29. (a) Larrow, J. F.; Jacobsen, E. N.; Gao, Y.; Hong, Y.; Nie, X.; Zepp, C. M. *J. Org. Chem.* **1994**, *59*, 1939; (b) Larrow, J. F.; Jacobsen, E. N. *Org. Synth.* **1997**, *75*, 1.
30. For the most effective catalyst developed for the asymmetric dihydroxylation of terminal olefins, see: (a) Becker, H.; Sharpless, K. B. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 448; For a general review of the AD reaction, see: (b) Kolb, H. C.; Vannieuwenhze, M. S.; Sharpless, K. B. *Chem. Rev.* **1994**, *94*, 2483.

31. (a) Kumar, P.; Naidu, S. V.; Gupta, P. *Tetrahedron* **2007**, *63*, 2745; (b) Kumar, P.; Gupta, P. *Synlett*, **2009**, 1367; (c) Yu, Q.; Wu, Y.; Xia, L. J.; Tang, M. H.; Wu, Y. L. *Chem. Commun.* **1999**, 129.
32. Kotkar, S. P.; Sudalai, A. *Tetrahedron: Asymmetry* **2006**, *17*, 1738.
33. Keck, G. E.; Boden, E. P. *Tetrahedron Lett.* **1984**, *25*, 265.
34. Grieco, P. A.; Nishizawa, M.; Burke, S. D.; Marinovic, N. *J. Am. Chem. Soc.* **1976**, *98*, 1612.
35. Tanako, S.; Yanase, M.; Ogasawara, K. *Synthesis* **1989**, 39.
36. Hutchings, B. L.; Gordon, S.; Ablondi F.; Wolf, C. F.; Williams, J. H. *J. Org. Chem.* **1952**, *17*, 19.
37. (a) Marshall, E. *Science* **2000**, *290*, 428; (b) Kaiser, J. *Science* **1998**, *281*, 1930; (c) Ridley, R. *Nat. Med.* **1998**, *4*, 894.
38. Arison, B. H.; Hwang, S. B. *Acta Pharmacol. Sin.* **1991**, *5*, 345.
39. Zhang, H. J.; Tamez, P. A.; Hoang, V. D.; Tan, G. T.; Van Hung, N.; Xuan, L. T.; Huong, L. M.; Cuong, N. M.; Thao, D. T.; Soejarto, D. D.; Fong, H. H. S.; Pezzuto, J. M. *J. Nat. Prod.* **2001**, *64*, 772.
40. Lee, A.L.; Ley, S. V. *Org. Biomol. Chem.* **2003**, *1*, 3957.
41. (a) Ridley, R. G. *Science* **1999**, *285*, 1502; (b) Greenwood, B.; Mutabingwa, T. *Nature* **2002**, *415*, 670.
42. Barata, L. E. S.; Santos, L. S.; Ferri, P. H.; Phillipson, J. D.; Paine, A.; Croft, S. L. *Phytochemistry* **2000**, *55*, 589.
43. Zacchino, S.; Rodriguez, G.; Pezzenati, G.; Orellana, G.; Enriz, R.; Sierra, M. G. *J. Nat. Prod.* **1997**, *60*, 659.
44. Curti, C.; Zanardi, F.; Battistini, L.; Sartori, S.; Rassu, G.; Pinna, L.; Casiraghi, G. *J. Org. Chem.* **2006**, *71*, 8552.
45. Choppin, S.; Colobert, F.; Hanquet, G.; Leroux, F.; Mahmoudi, N.; Mazier, D. *EP 2177496 A1/2010*.
46. Nagaraju, M.; Chandra, R.; Gawali, B. B. *Synlett* **2012**, *23*, 1485.
47. Vanrheenen, V.; Kelly, R. C.; Cha, D. Y. *Tetrahedron Lett.* **1976**, *17*, 1973.
48. Raghavan, S.; Reddy S. R. *J. Org. Chem.* **2003**, *68*, 5754.
49. George, S.; Sudalai, A. *Tetrahedron: Asymmetry* **2007**, *18*, 975.
50. George, S.; Suryavanshi, G.; Sudalai, A. *Tetrahedron: Asymmetry* **2010**, *21*, 558.
51. Makabe, H.; Kong, L. K.; Hirota, M. *Org. Lett.* **2003**, *5*, 27.
52. Villieras, J.; Rambaud, M.; Graff, M. *Tetrahedron Lett.* **1985**, *26*, 53.

CHAPTER 2

**CN-Assisted Oxidative Cyclization of Cyano
Cinnamates and Styrene Derivatives: A Facile
Entry to 3-Substituted Chiral Phthalides and
its Application to the Synthesis of (-)-
Matteucen C and Butylphthalide**

Section I:

CN-Assisted Oxidative Cyclization of Cyano Cinnamates and Styrene Derivatives: a Facile Entry to 3-Substituted Chiral Phthalides

2.1.1 Introduction

Chiral phthalides [isobenzofuran-1(3H)-ones] comprising of 5-membered lactones are found in a large number of plant products displaying broad and potent biological activities.¹ Some representative examples are shown in **Fig. 1**. 3-Butylphthalide (**1**), a component in the Chinese folk medicine extracted from celery seed oil,^{2a} is in phase II clinical trials in China and potentially can be used for the treatment of stroke.^{2b} Moreover, it is employed for seasoning and flavoring purposes, shows anticonvulsant action,^{2c} increases the duration of anesthesia,^{2d} and exhibits cerebral antiischemic action.^{2e}

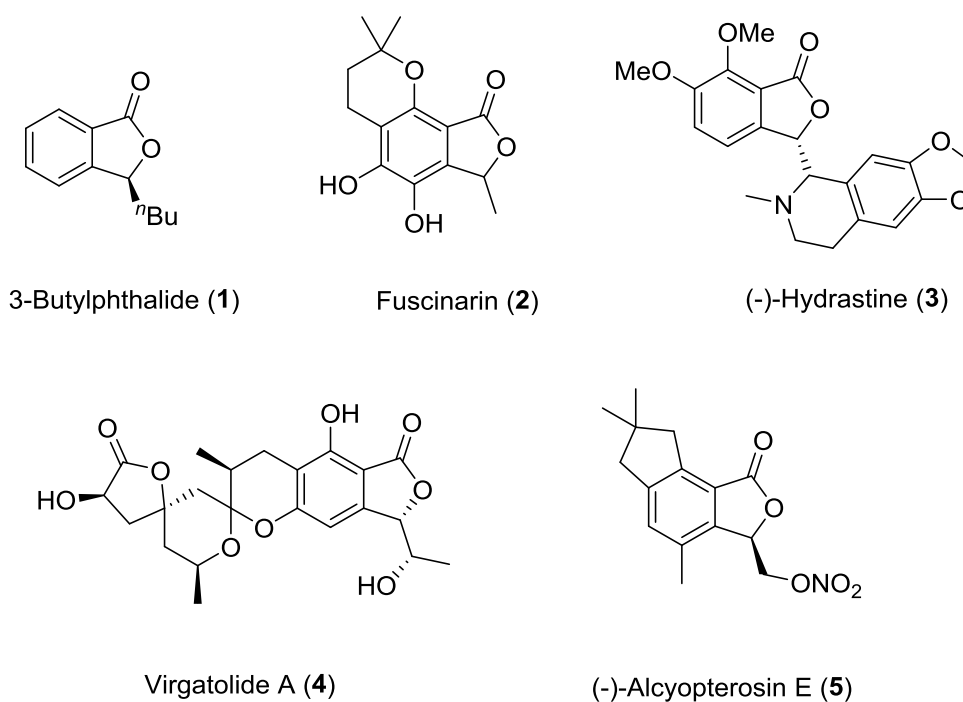


Fig. 1: Some of the examples of chiral phthalides

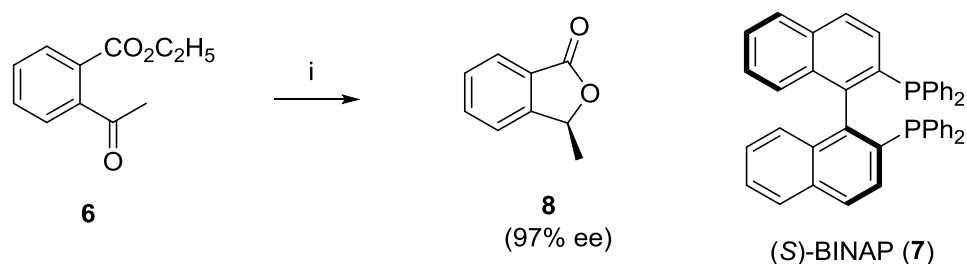
Fuscinarin (**2**) is a potent human CCR5 antagonist, used effectively for blocking HIV entry into host cells.³ (-)-Hydrastine (**3**) is active at the opioid receptor.⁴ In addition, it possesses antipaclitaxel-resistant human ovarian cancer activity through c-Jun kinasemediated apoptosis and is in phase I clinical trials.⁵ Both Virgatolide A (**4**) and (-)-Alcyopterosin E (**5**) show cytotoxic activity against HeLa cells.⁶ Due to the biological importance of 3-substituted phthalides **1-5** (**Fig. 1**), their molecular architectures have become a platform for new synthetic methodology development.⁷

2.1.2 Review of Literature

Literature search revealed that there are various methods available for the synthesis of 3-substituted phthalide derivatives, which are described below.

Noyori's approach (1990)⁸

Noyori *et al.* have described the synthesis of chiral phthalide ((*S*)-3-methyl isobenzofuran-1(3H)-one) **8** via asymmetric hydrogenation of ethyl *o*-acetylbenzoate **6** in ethanol with 0.4 mol% of the (*S*)-BINAP(**7**)-Ru catalyst at 100 atm H₂ pressure in 97% ee and 97% yield (**Scheme 1**).

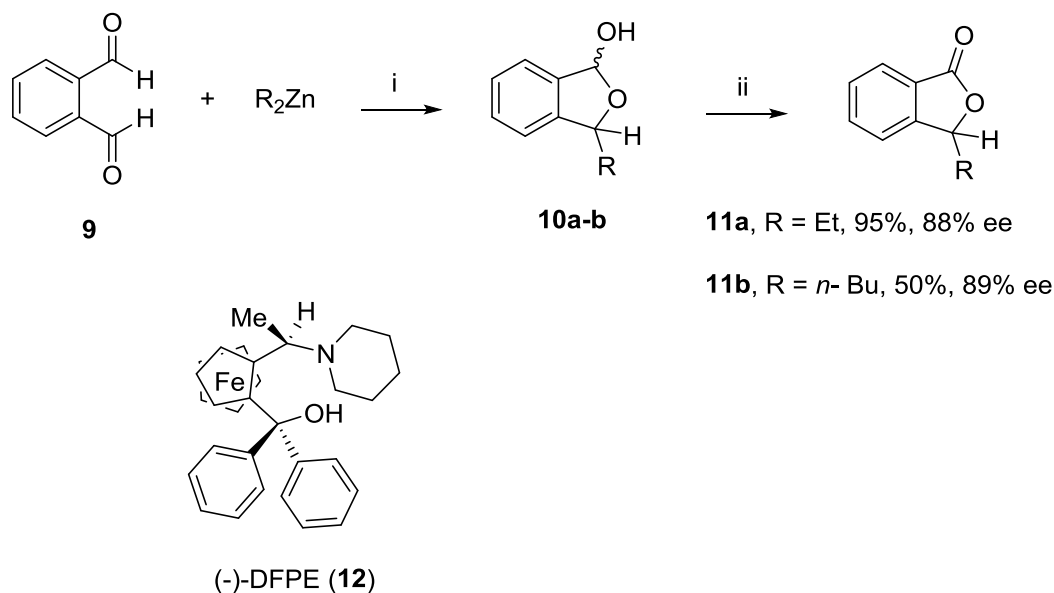


Scheme 1: (i) Ru(OCOCH₃)₂[(*S*)-BINAP] (0.4 mol%), H₂ (100 atm), EtOH, 0.5N HCl, 35 °C, 165 h, 97%.

Butsugan's approach (1992)⁹

Butsugan *et al.* have reported the synthesis of optically active 3-ethyl- and 3-*n*-butylphthalides using enantioselective addition of dialkylzinc reagents. Thus, *o*-phthalaldehyde **9** was subjected to asymmetric addition of dialkyl reagents, catalyzed

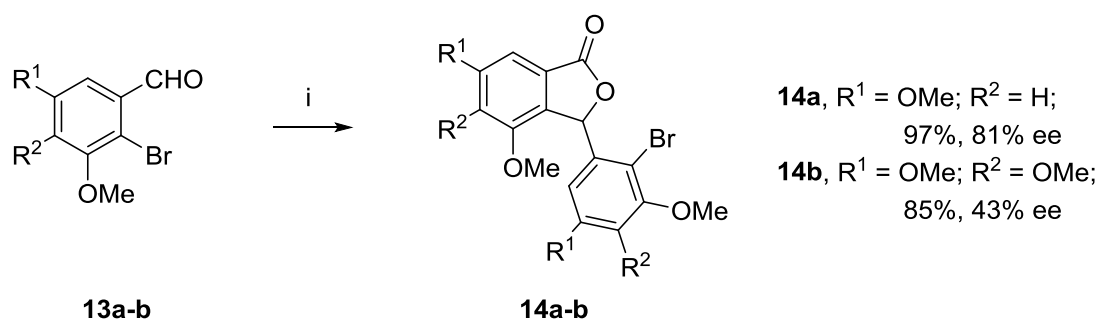
by chiral 1,2-disubstituted ferrocenyl amino alcohol **12**, followed by oxidation of the resulting lactols **10a-b** to provide phthalides **11a-b** in 88-89% ee (**Scheme 2**).



Scheme 2: (i) (-)-DFPE (**12**) (5 mol%), 25 °C, 1-3 h, (ii) Ag_2O , 0 °C.

Lin's approach (2002)¹⁰

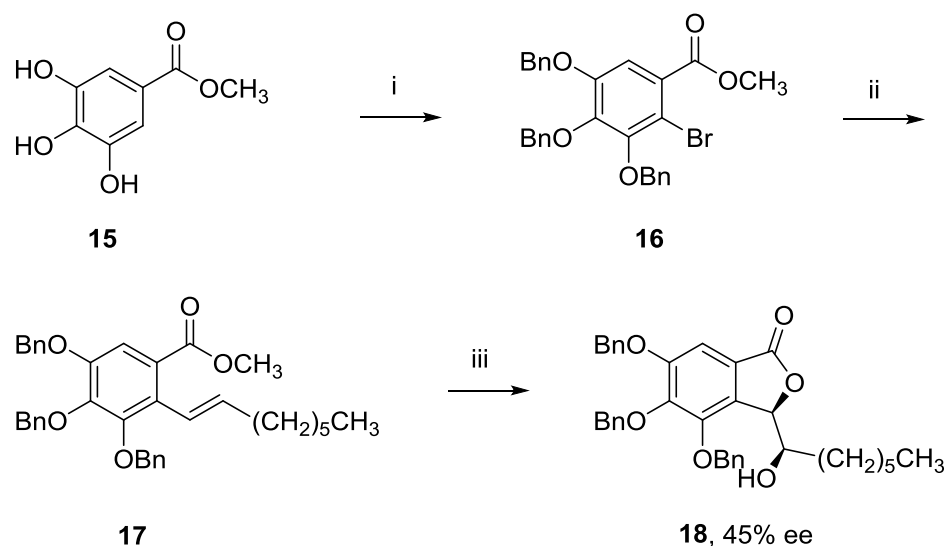
Ni-catalyzed tandem homo addition of *o*-bromoaldehydes **13a-b** via *in situ* cyclization was developed in presence of (*S*)-BINAP (**7**) and Zn that provided optically pure phthalides **14a-b** in good yields with moderate enantiomeric excess (**Scheme 3**).



Scheme 3: (i) $NiCl_2(PPh_3)_2$ (0.2 equiv). (*S*)-BINAP (**7**), Zn, toluene, 90 °C.

Mori's approach (2003)¹¹

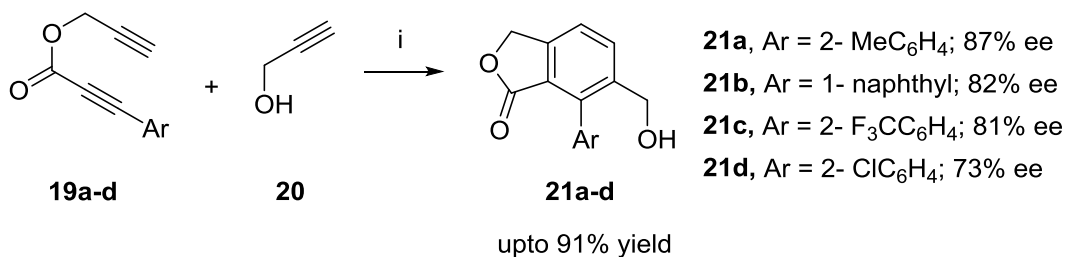
Mori *et al.* have used Sharpless asymmetric dihydroxylation as the key reaction. Thus, commercially available methyl 3,4,5-trihydroxybenzoate **15** was benzylated completely and subjected to bromination with NBS to afford bromo compound **16** in 96% yield. The bromo compound **16** was subjected to Miyaura-Suzuki coupling with (*E*)-1-octeneboronic acid to give olefin **17**. Asymmetric dihydroxylation of olefin **17** with AD-mix- β proceeded to furnish phthalide **18** in 54% yield with 45% ee (**Scheme 4**).



Scheme 4: (i) (a) BnBr, K₂CO₃, 96%; (b) NBS, DMF, 96%; (ii) (*E*)-CH₃(CH₂)₅CH=CHB(OH)₂, Pd(PPh₃)₄, K₂CO₃, C₆H₆:EtOH (5:1), 71%; (iii) AD-mix- β , CH₃SO₂NH₂, *tert*-BuOH:H₂O (1:1), 54%.

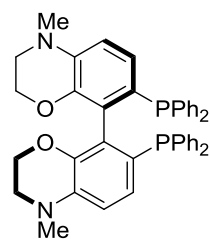
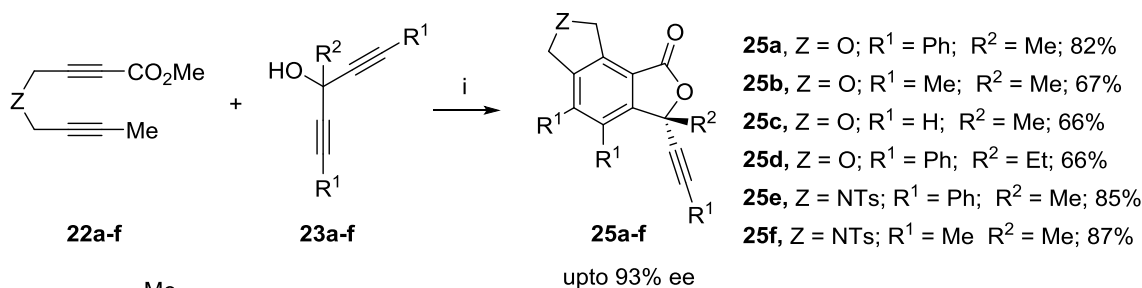
Tanaka's approach (2004)¹²

Tanaka *et al.* have described the enantioselective synthesis of axially chiral phthalides by the cationic [Rh^I(H₈-BINAP)] complex-catalyzed alkyne cyclotrimerization. The reaction of aryl-substituted 1,6-diyne **19a-d** with terminal monoyne **20** in the presence of the cationic complex [Rh^I(H₈-BINAP)] provided axially chiral phthalides **21a-d** in high yields with moderate enantioselectivity (**Scheme 5**).



Scheme 5: (i) 5% [Rh{(S)-H₈-BINAP}]BF₄ (1 mol%), CH₂Cl₂, 25 °C, 3 h.

Same authors have developed a cationic rhodium(I)/Solphos (**24**) complex-catalyzed asymmetric one-pot transesterification followed by [2+2+2] cycloaddition of 1,6-diyne esters **22a-f** with tertiary propargylic alcohols **23a-f** leading to enantioenriched tricyclic 3,3-disubstituted phthalides **25a-f** in good yields (66-87%) with moderate enantioselectivity (**Scheme 6**).



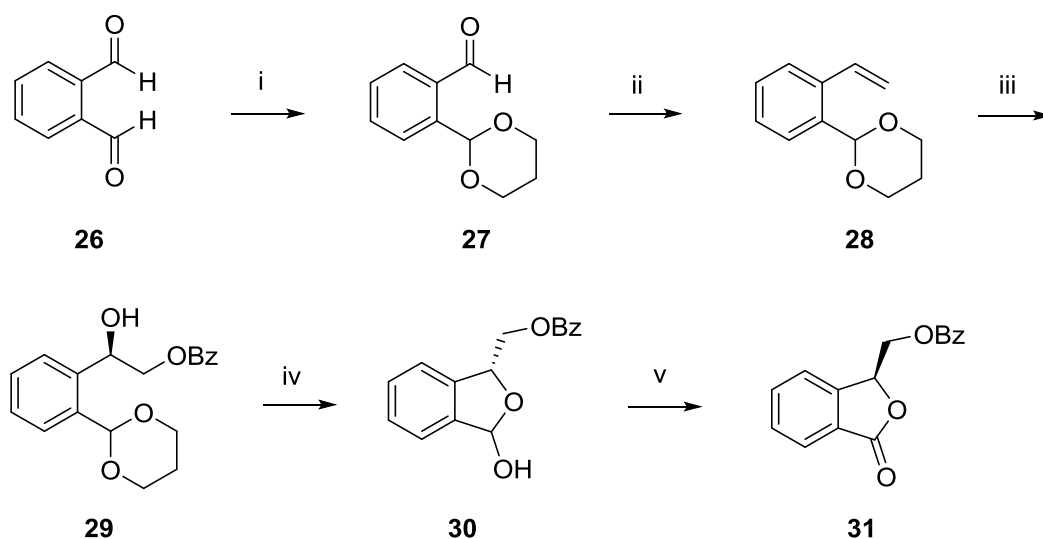
(R)-Solphos (**24**)

Scheme 6: (i) 5% [Rh(cod)₂]BF₄/(R)-Solphos (**24**) (1 mol%), CH₂Cl₂, 25 °C, 1-3 h.

Vaccher's approach (2005)¹³

Vaccher *et al.* have reported the synthesis of 3-benzoyloxy methylisobenzofuranone **30** using Sharpless asymmetric dihydroxylation as the key step. Thus, phthalaldehyde **26** was firstly protected using propane-1,3-diol to give the benzaldehyde derivative

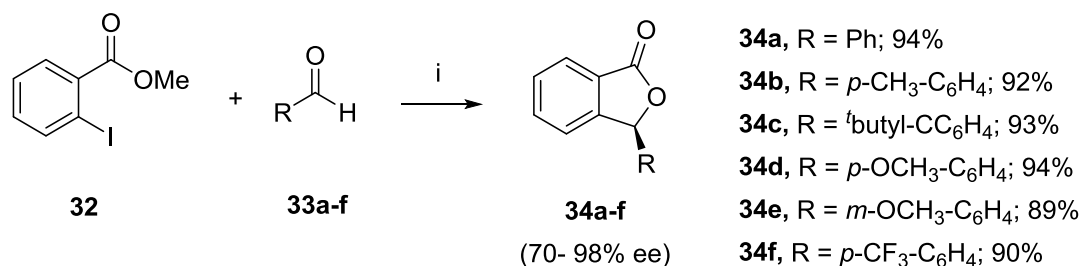
27, which was subjected to Wittig reaction to afford the styrene derivative **28**. Asymmetric dihydroxylation using AD-mix- β gave diol, which was selectively protected with benzoyl chloride to afford compound **29** with a free secondary hydroxyl group. Removal of the acetal from **29** in acidic medium gave benzo[*c*]furan **30**, which was converted to phthalide **31** in 50% yield using $\text{RuCl}_3\text{-NaIO}_4$ combination as oxidant (**Scheme 7**).



Scheme 7: (i) *p*-TSA, propan-1,3-diol, toluene, 5 h, 82%; (ii) *tert*-BuOK, $\text{CH}_3(\text{C}_6\text{H}_5)_3\text{PBr}$, toluene, 3 h, 78%; (iii) (a) AD-mix- β , *tert*-BuOH: H_2O (1:1), 12 h, 72%; (b) BzCl, $(\text{C}_2\text{H}_5)_3\text{N}$, toluene, 5 h, 78%; (iv) *p*-TSA, acetone, H_2O , 1 h, 82%; (v) NaIO_4 , RuCl_3 , CH_3CN , EtOAc, H_2O , 50%.

Cheng's approach (2007)¹⁴

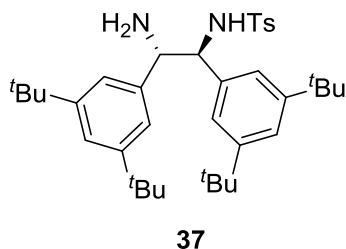
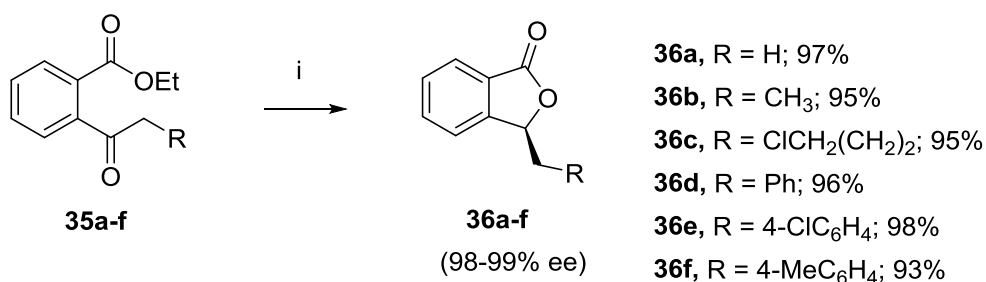
Cheng *et al.* have reported Co-bidentate phosphine complex-catalyzed synthesis of phthalides **34a-f**. Thus, methyl 2-iodobenzoates **32** underwent cyclization reactions with various aromatic aldehydes **33a-f** in the presence of $[\text{CoI}_2(\text{dppe})]$ (5 mol%) and Zn powder in dry THF at 75 °C for 24 h to give the corresponding phthalide derivatives **34a-f** in 89-94% yields with 70-98% ee (**Scheme 8**).



Scheme 8: (i) [CoI₂(dppf)] (5 mol%), Zn, THF, 75 °C, 24 h.

Xu's approach (2009)¹⁵

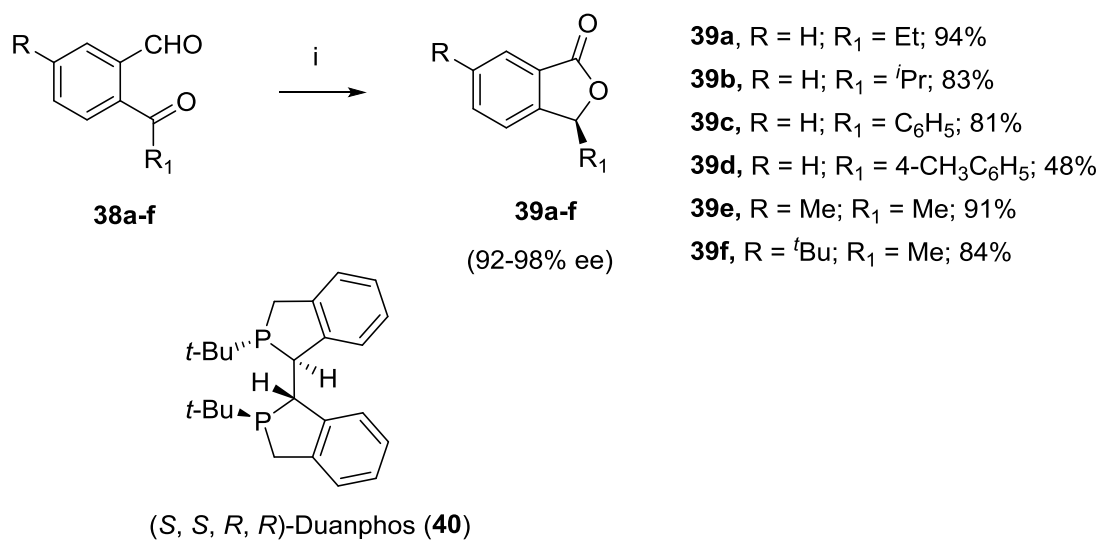
Xu *et al.* have reported a new diamine ligand **37** for asymmetric transfer hydrogenation (ATH) to synthesize 3-substituted phthalides. The reductive cyclization of 2-acylaryl carboxylates **35a-f** via the new [RuCl₂(*p*-cymene)]₂/**37**-catalyzed ATH and subsequent *in situ* lactonization under aqueous conditions proceeded to give a variety of 3-substituted phthalides **36a-f** in high yields (93-97%) with high ee (98-99%) (**Scheme 9**).



Scheme 9: (i) [RuCl₂(*p*-cymene)]₂/ligand **37** (1 mol%), HCO₂Na, H₂O/CH₂Cl₂, 40 °C, 24 h.

Dong's approach (2009)¹⁶

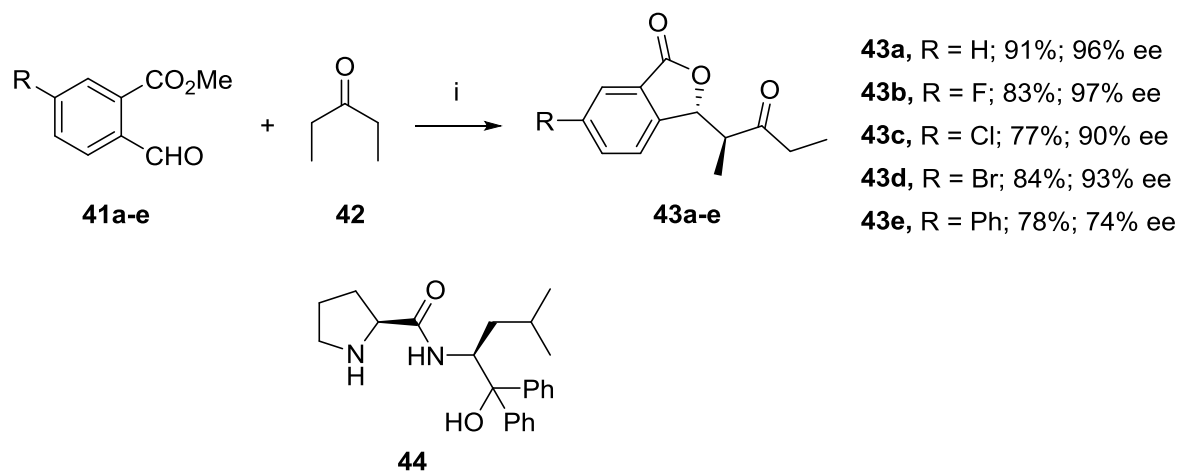
Dong *et al.* have employed [Rh(cod)Cl]₂-catalyzed hydroacylation of ketones **38a-f** in presence of duanphos **40** (10 mol%), and AgNO₃ (10 mol%) to give chiral phthalides **39a-f** in 81-94% yields with 92-98% ee (**Scheme 10**).



Scheme 10: (i) [Rh(cod)Cl]₂ (10 mol%), Duanphos (**40**) (10 mol%), AgNO₃ (10 mol%), toluene, 90 °C, 3-3.5 h.

Wang's approach (2010)¹⁷

Wang *et al.* have described the synthesis of chiral phthalides **43a-e** by employing organocatalytic asymmetric aldol-lactonization as the key reaction.

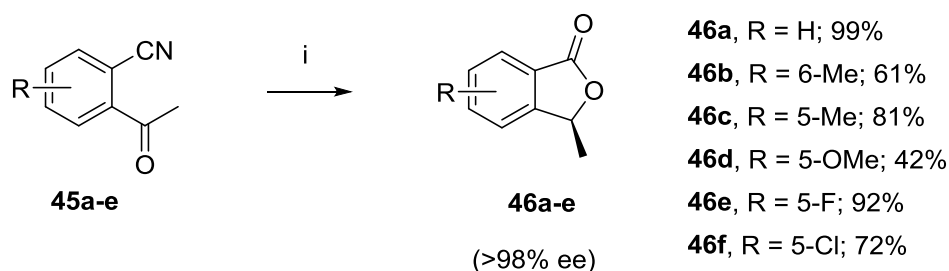


Scheme 11: (i) (a) *L*-prolinamide alcohol **44** (2.5 mol%), PhCO₂H (2.5 mol%), -40 °C, 12-24 h; (b) K₂CO₃, acetone:methanol (10:1), 15 min.

Thus, 2-formylbenzoic esters **41a-e** and ketone **42** were subjected to aldol reaction using *L*-prolinamide alcohol **44** as catalyst and PhCO₂H as an additive to give 3-substituted phthalides **43a-e** in 77-91% yield with 74-97% ee (**Scheme 11**).

Gotor's approach (2012)¹⁸

Gotor *et al.* have described Baker's yeast-catalyzed bioreduction of 2-acetylbenzo nitriles **45a-e** followed by aqueous HCl that provided access to enantiopure (*S*)-3-methylphthalides **46a-e** in moderate to excellent yields (42-99%) with >98% ee (**Scheme 12**).



Scheme 12: (i) (a) Baker's yeast, glucose, H₂O, 25 °C, 16-72 h; (b) HCl 1M, 25 °C, 48 h.

2.1.3 Present Work

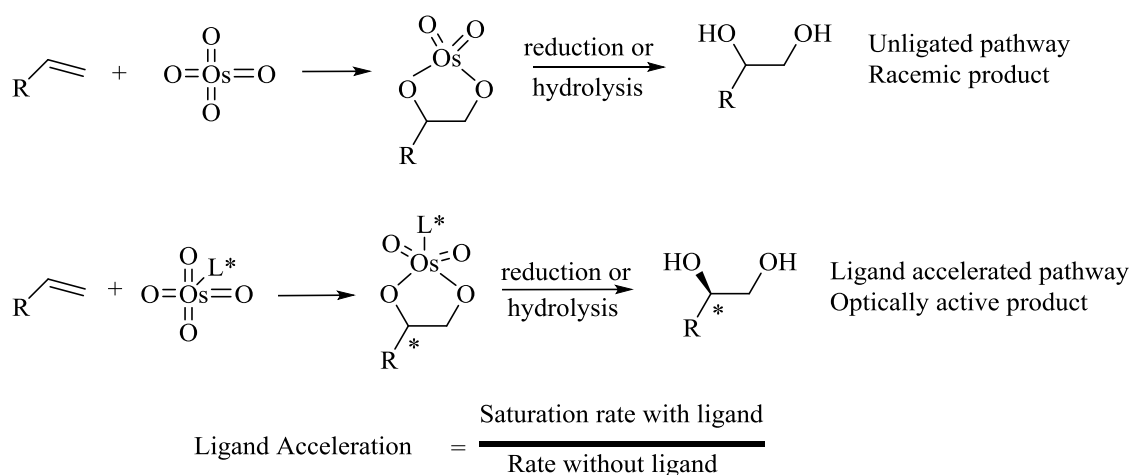
2.1.3.1 Objective

As can be seen from the above discussion, the reported methods for the synthesis of 3-substituted phthalides employ either chiral auxiliaries or expensive organometallic reagents in stoichiometric amounts and often lack in broad substrate scope and higher reaction stereoselectivity; only a few are atom economical. In this context, a more practical and efficient synthesis of functionalized 3-substituted phthalide derivatives is highly desirable. In this section, we present a single-step oxidative cyclization of cyanocinnamates and styrenic substrates that affords 3-substituted phthalides in high yields *via* synergetic acceleration of CN and osmate ester groups present in proximity

positions. Since the method involves asymmetric dihydroxylation (ADH) as the key chiral inducing reaction, a brief account of ADH is described below.

2.1.3.2 Asymmetric Dihydroxylation (ADH)

In recent years, much attention has been focused on the catalytic asymmetric synthesis. It often has significant economic advantages over stoichiometric asymmetric synthesis for industrial-scale production of enantiomerically pure compounds. All these asymmetric reactions crucially depend on ligand acceleration effect (LAE).¹⁹ Among all these reactions, Sharpless catalytic Asymmetric Dihydroxylation (ADH) is one of the most important practical and widely used reaction in organic synthesis. It has become the most general method for the preparation of optically active *vicinal-syn*-diols from activated as well as unactivated olefins.²⁰ In 1936, Criegee *et al.*²¹ found that addition of pyridine or any other tertiary amine to osmylation of olefins, accelerated the rate of reaction considerably. A major breakthrough had occurred in the field of asymmetric oxidation when Sharpless *et al.*^{20b} demonstrated that asymmetric induction could be achieved when chiral amines were added to OsO₄-mediated asymmetric oxidation of olefins.



Scheme 13: Mechanism of OsO₄-catalyzed dihydroxylation of olefin

Among the various ligands screened best results were obtained with ligands which were representatives of the cinchona alkaloid family, namely dihydroquinidine (DHQD) and dihydroquinine (DHQ) (**Scheme 13**).²²

To improve the %ee of the chiral diol, the second catalytic cycle of ADH should be avoided and this was achieved by employing the $\text{K}_3\text{Fe}(\text{CN})_6$ as reoxidant and using biphasic conditions (**Fig. 2**).

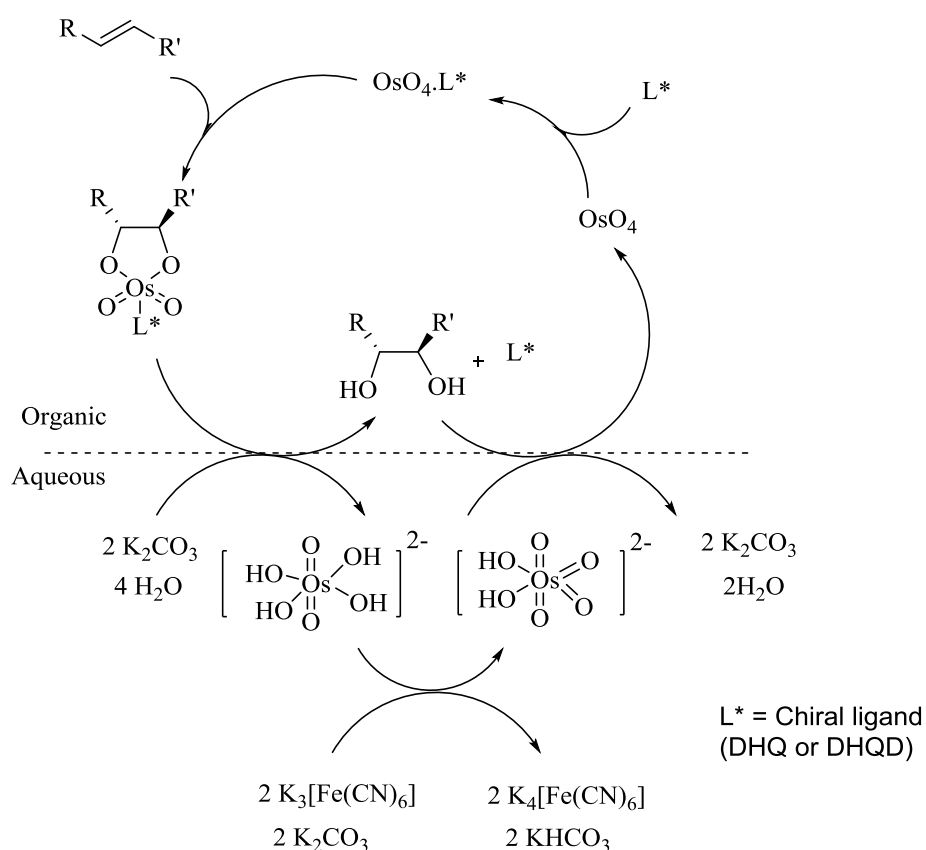


Fig. 2: Catalytic cycle for ADH using $\text{K}_3\text{Fe}(\text{CN})_6$ as co-oxidant

These conditions helped in protecting the organic osmate-(VI) monoglycolate ester from inopportune oxidation prior to hydrolysis and thereby releasing the diol and ligand to the organic phase and osmium-(VI) to the aqueous phase. Subsequently, osmium-(VI) reoxidized and recycled into the catalytic cycle. Further improvement in the ADH was realized by the addition of methyl sulfonamide (MeSO_2NH_2) to the

reaction mixture. It also helps to accelerate the hydrolysis, thus facilitating the dihydroxylation smoothly. Addition of methyl sulfonamide also allowed carrying out the reactions of 1,2-di-tri- and tetra-substituted olefins at 0 °C, which improved the selectivity as well as enantiomeric excess.

In order to develop the asymmetric version of the Os-catalyzed ADH reaction, Sharpless and coworkers screened various chiral ligands and found out that the derivatives of cinchona alkaloids gave excellent results. Among all the 250 derivatives of cinchona alkaloid ligands screened, the *bis*-DHQ or DHQD ethers of phthalazine-1,4-diol have proven to be the best for obtaining high enantioselective diols²³ (**Fig. 3**).

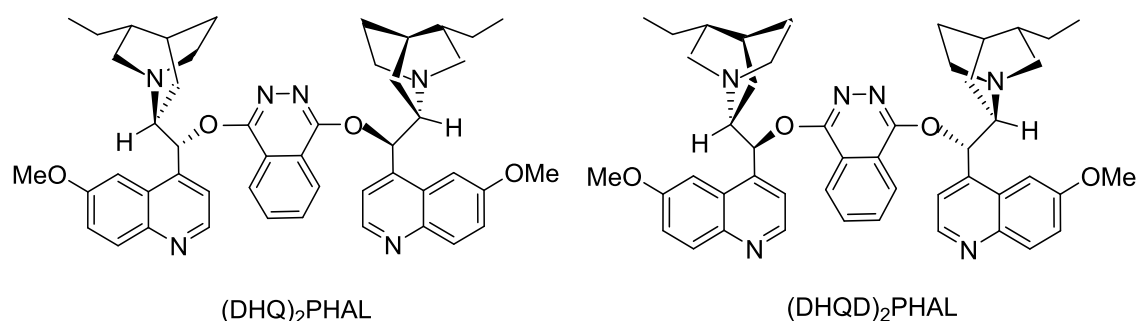


Fig. 3: Ligands for asymmetric dihydroxylation reaction

Studies have demonstrated the importance of enzyme-like binding pocket of the dimeric cinchona alkaloid for high enantioselectivity of the chiral diols.²⁴ Sharpless *et al.*²⁰ have shown that the facial selectivity for both ligands (DHQ)₂PHAL and (DHQD)₂PHAL is different, based on their ability to induce the ee into the diols. This observation has led to the development of mnemonic model (**Fig. 4**) in which olefin with the constraints will be attacked either from the top (i.e. β) face in the presence of

dihydroquinidine (DHQD) derivatives or from the bottom (i.e. α) face in the presence of dihydroquinine (DHQ) derived ligand.

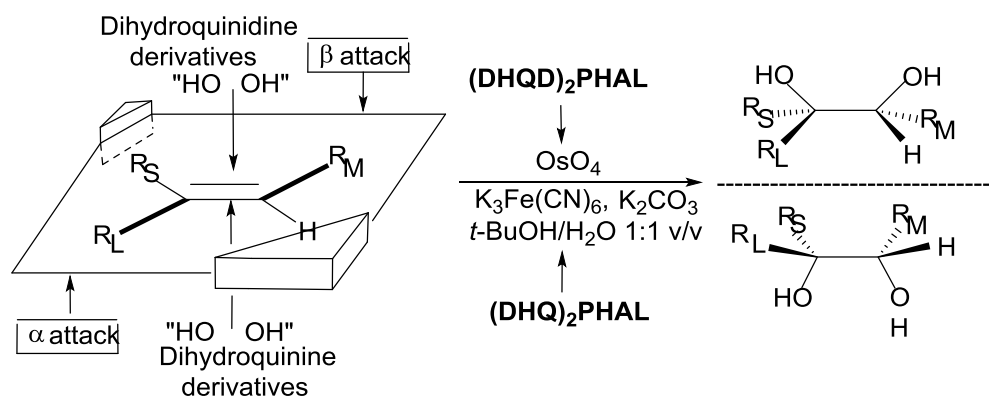
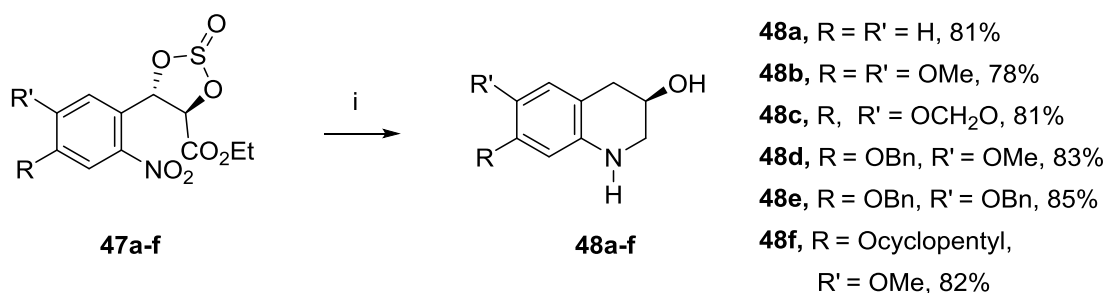


Fig. 4: Enantioselectivity mnemonic scheme

2.1.4 Results and Discussion

Recently, Sudalai *et al.* have developed a novel protocol of ADH process followed by Co-catalyzed “one-pot” reductive cyclization (CoCl₂-NaBH₄) of *nitro* cyclic sulfites **47a-f** that led to the construction of 3-substituted tetrahydroquinolin-3-ols **48a-f** (Scheme 14).²⁵



Scheme 14: (i) CoCl₂·6H₂O (1 mol%), NaBH₄ (4 equiv), EtOH, 0 to 25 °C.

In analogy with this, we reasoned that subjecting *ciano* cyclic sulfites to the same reaction conditions should afford synthetically useful benzazepines.²⁶ In order to synthesize cyano cyclic sulfite, we visualized a strategy in which cyano diol **52** could

serve as a starting material to *cyano* cyclic sulfite. Accordingly, *o*-cyanobenzaldehyde **49** was subjected to Wittig olefination to afford (*E*)- α,β -unsaturated ester **50a** in 88% yield (**Scheme 15**). The formation of cinnamate ester **50a** was confirmed from its ^1H NMR spectrum, which showed two doublets at δ 6.60 (d, $J = 16$ Hz, 1H) and 7.96 (d, $J = 16$ Hz, 1H) confirming the presence of α - and β -CH olefin protons of -CH=CHCO₂Et. This was further ascertained by the presence of characteristic carbon signals at δ 122.9 and 139.0 corresponding to α - and β -carbon signal of olefinic ester -CH=CHCO₂Et in its ^{13}C NMR spectrum (**Fig. 5**).

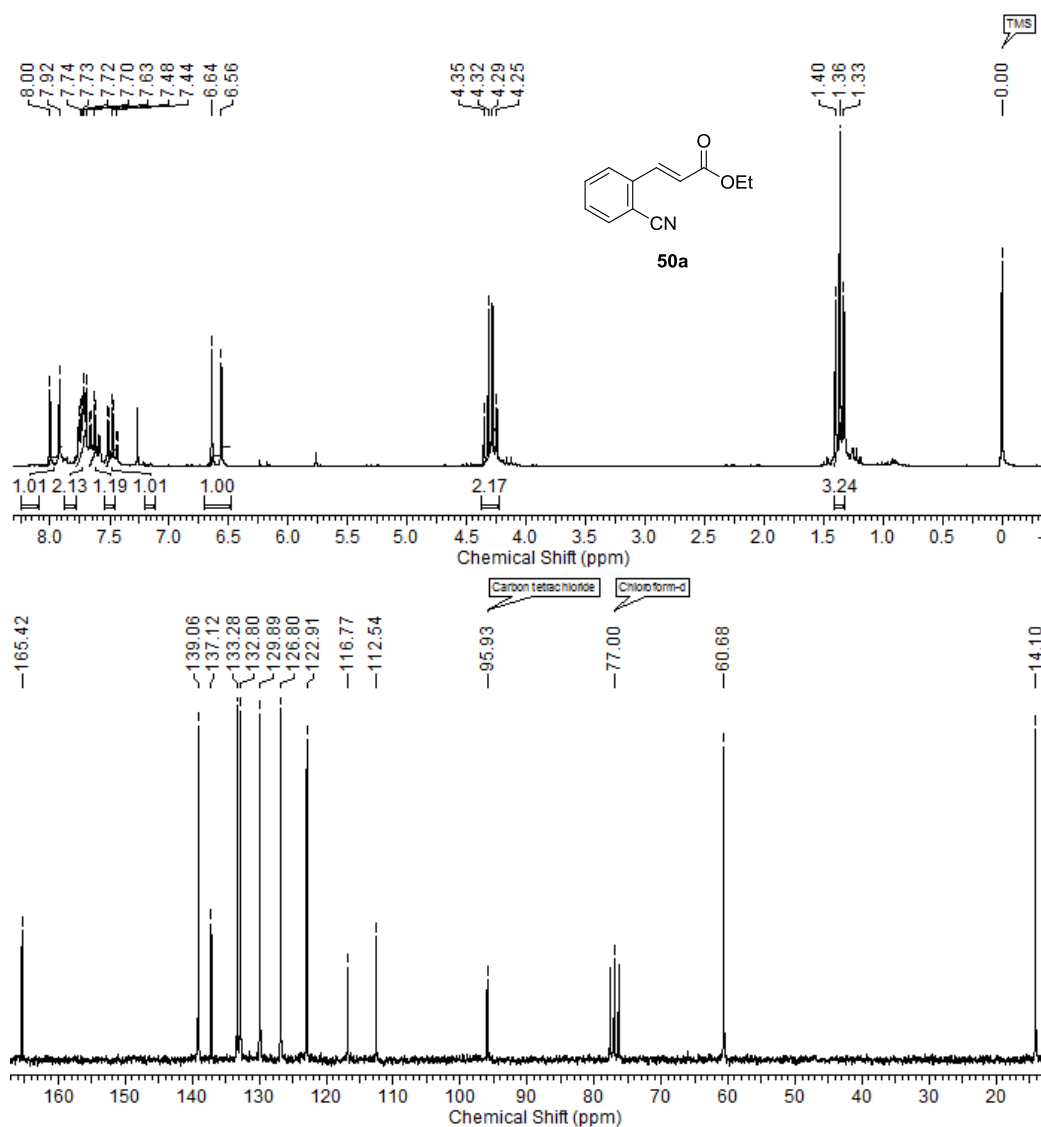
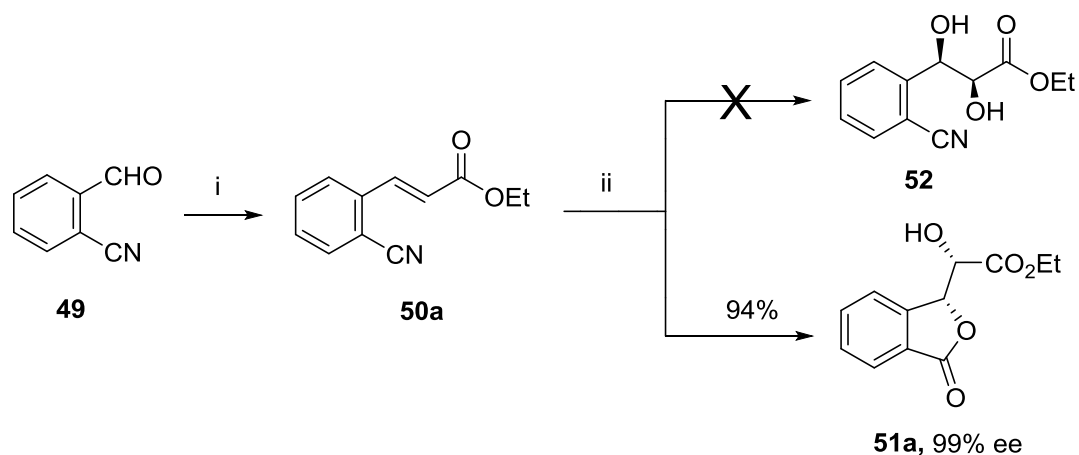


Fig. 5: ^1H and ^{13}C NMR spectra of *o*-cyanocinnamate **50a**

In order to validate our hypothesis, ethyl 2-cyanocinnamate **50a** was subjected to Sharpless asymmetric dihydroxylation using (DHQD)₂PHAL as the chiral ligand, with THF as co-solvent for better solubility. Surprisingly, the reaction took altogether a different course to give the cyclized chiral phthalide **51a** exclusively with 99% ee in a single step, instead of the expected cyano diol **52** (Scheme 15). This unexpected transformation is characterized by high rate, excellent yield and enantioselectivity, which is attributed to coordination assistance provided by the neighboring CN group to osmate ester, leading to faster hydrolysis of osmate ester in the catalytic cycle. Incidentally, the rate of ADH process for electron-deficient *o*-substituted cinnamates is generally reported to be sluggish (48 h to 7 days) giving products invariably with moderate enantioselectivity (88% ee).²⁷



Scheme 15: (i) $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$, benzene, reflux, 12 h, 88%; (ii) $\text{K}_2[\text{OsO}_2(\text{OH})_4]$ (0.1 mol%), (DHQD)₂PHAL (0.5 mol%), $\text{K}_3\text{Fe}(\text{CN})_6$ (3 equiv), K_2CO_3 (3 equiv), *tert*-BuOH:THF:H₂O (1:1:2), 25 °C, 7 h.

The formation of chiral phthalide **51a** was confirmed by the presence of a doublet of doublet at δ 4.66 (dd, $J = 2.1, 5.8$ Hz) integrating for one methine proton (-CH-OH) and a doublet at δ 5.79 (d, $J = 2.1$ Hz, 1H) integrating for benzylic proton (-CH-O-CO-) in its ¹H NMR spectrum. It was further substantiated by the carbon signals displaying at δ 70.3 and 80.3 in its ¹³C NMR spectrum, corresponding to carbons

attached to hydroxyl and lactone groups respectively (**Fig. 6**). The IR spectrum of phthalide **51a** displayed two strong absorption bands at 1720 and 1768 cm^{-1} due to the presence of ester and γ -lactone carbonyl groups respectively.

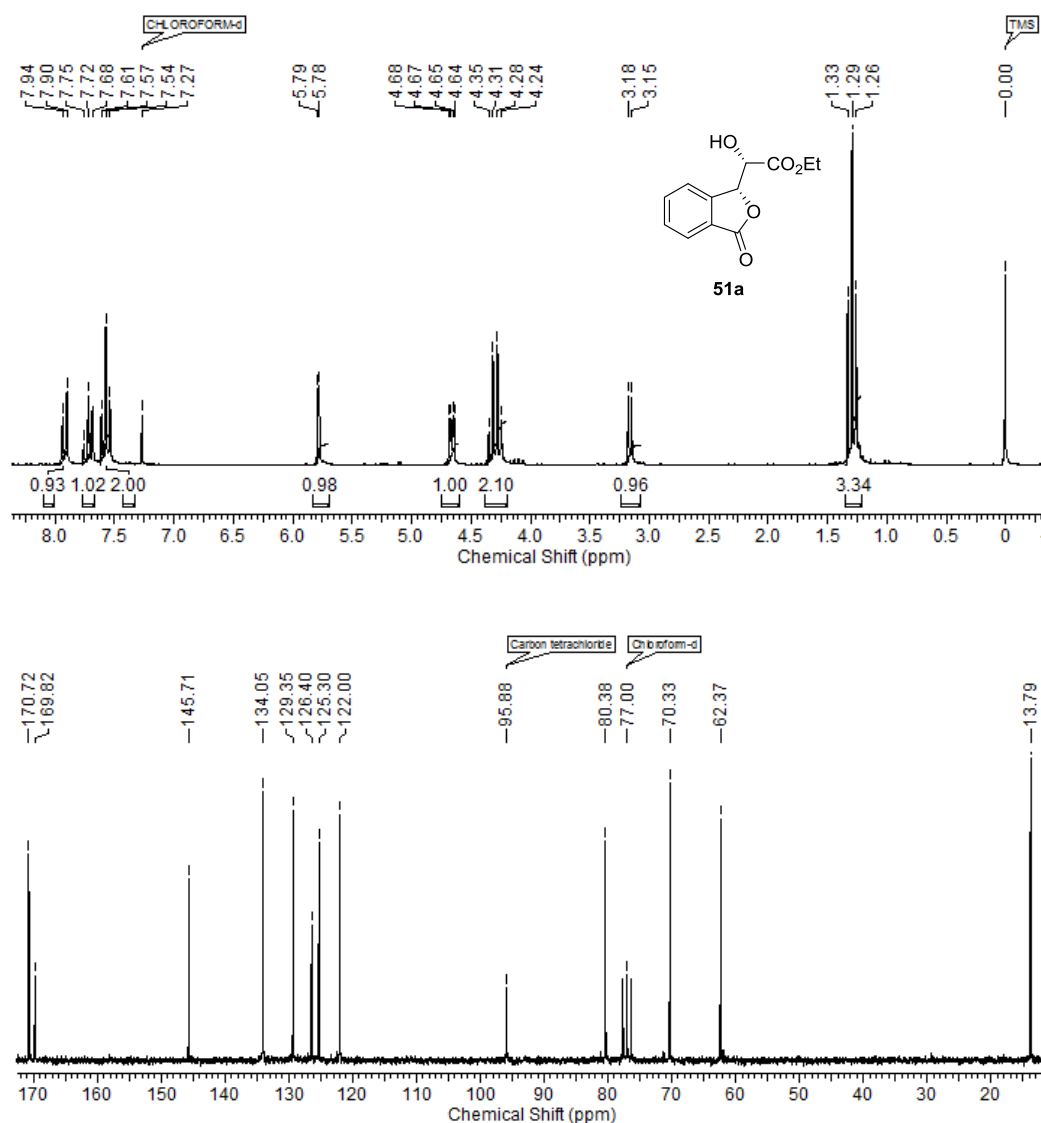


Fig. 6: ^1H and ^{13}C NMR spectra of phthalide **51a**

Further, the formation of chiral phthalide **51a** was confirmed by COSY and mass spectra (**Fig. 7**).

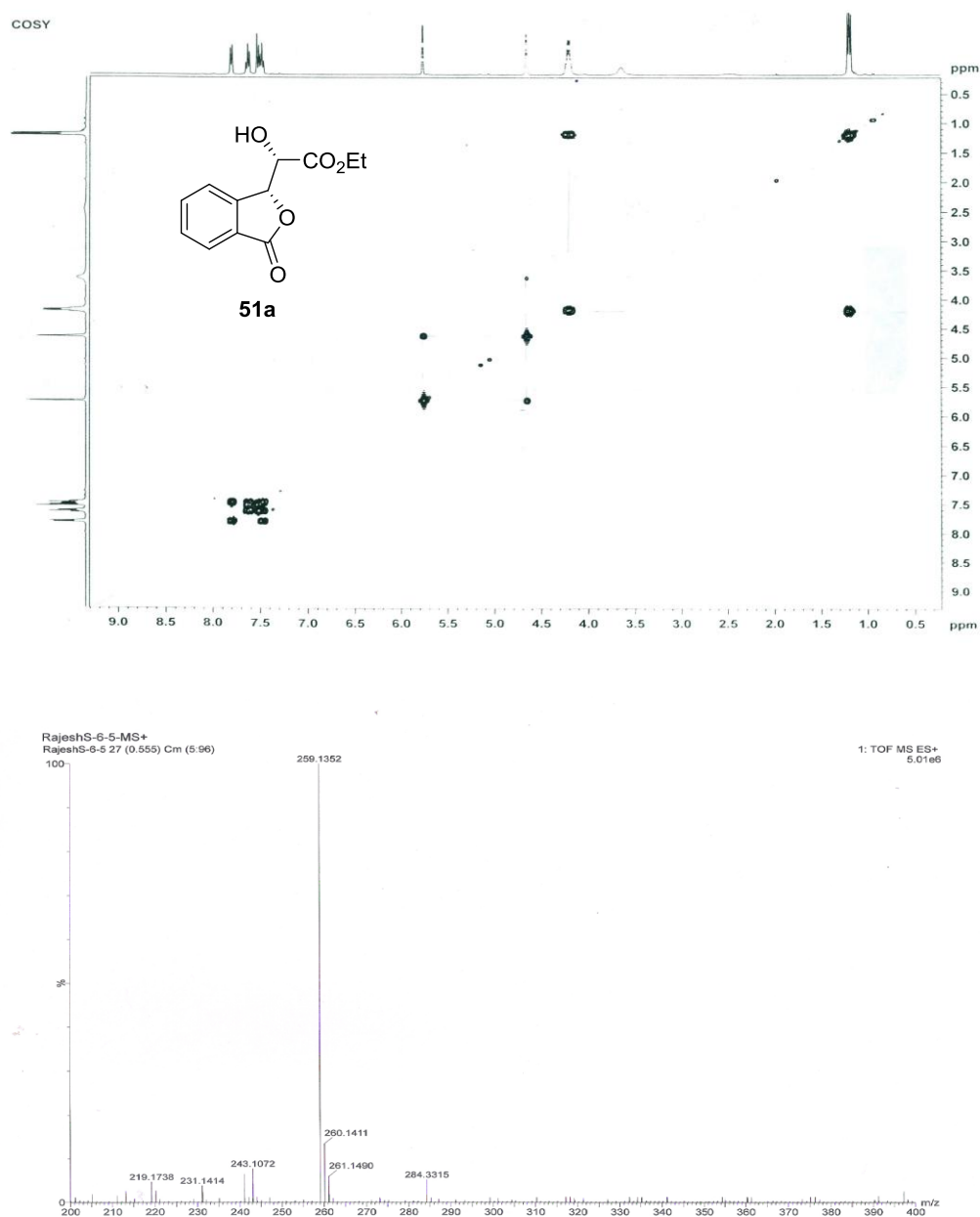


Fig. 7: COSY and mass spectra of phthalide **51a**

The enantiomeric excess (99% ee) of chiral phthalide **51a** was determined from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 12.16 (99.65%) and 13.80 (0.35%) (**Fig. 8**).

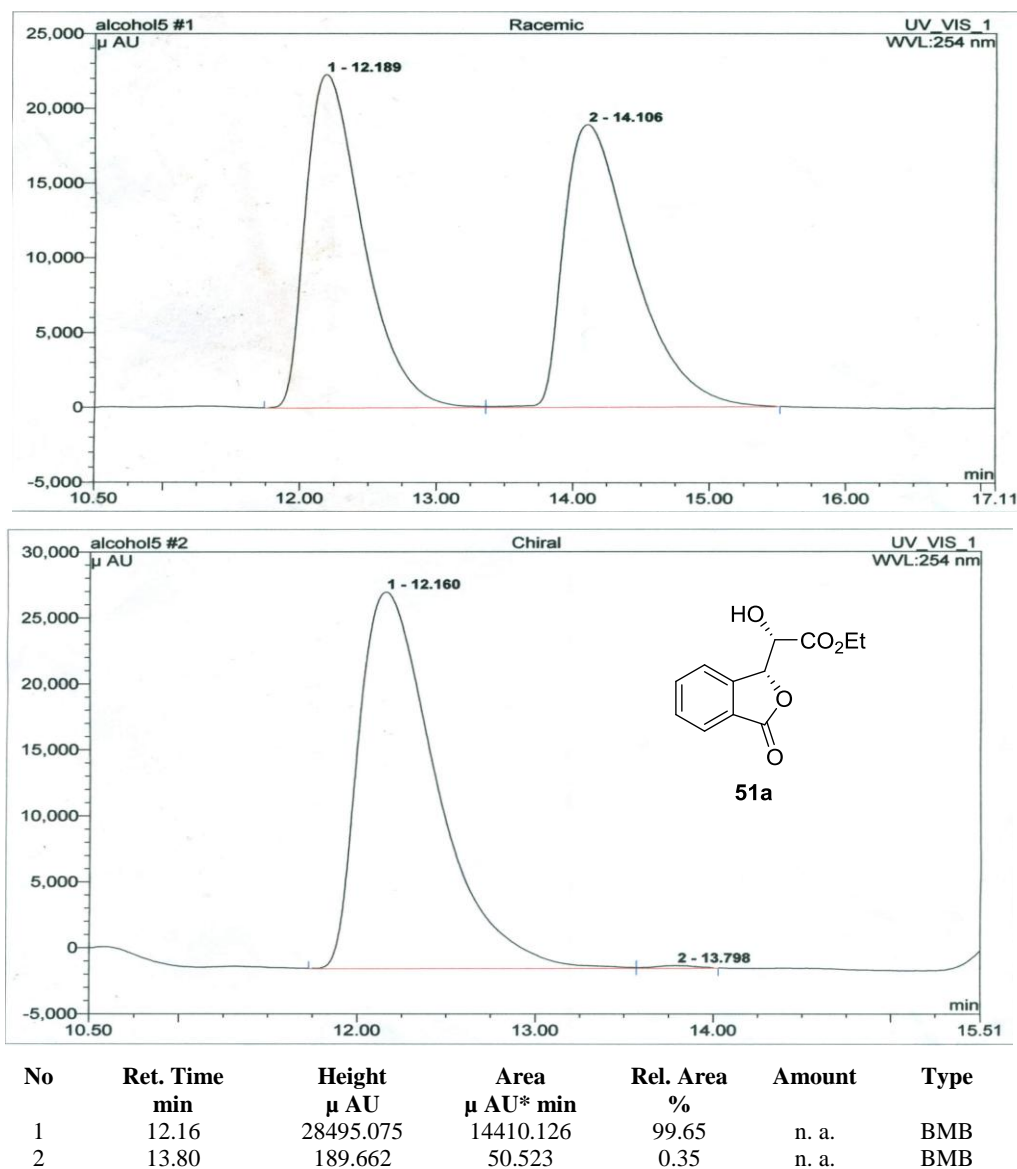
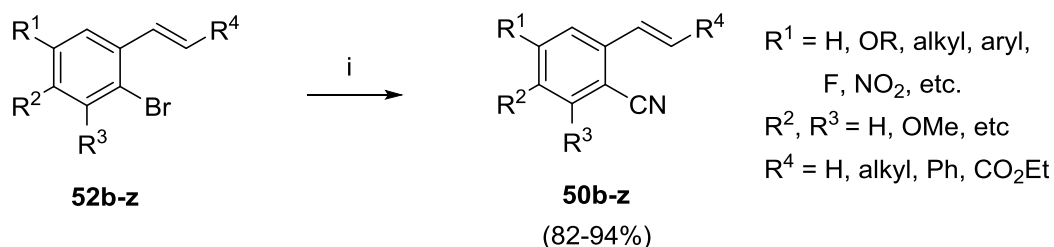


Fig. 8: HPLC chromatogram of phtalide **51a**

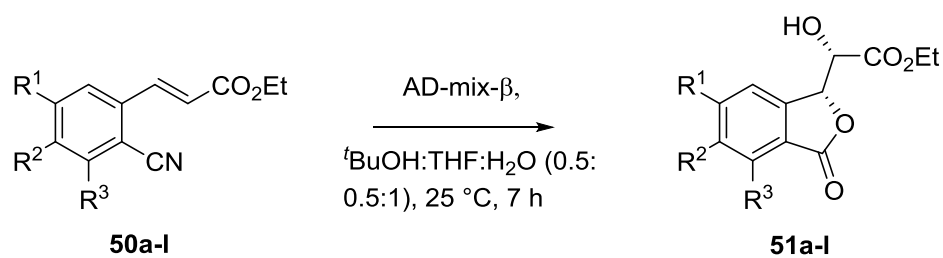
Encouraged by the result, we became interested in the scope of the reaction by subjecting other *o*-cyanoalkenes **50b-z**. *o*-Cyanoalkenes **50b-z** were prepared in 76-84% yield, in a single step, starting from the corresponding *o*-bromo alkene derivatives **52b-z** via Rosenmund-von Braun reaction with CuCN (3 equiv) and DMF as solvent at 150 °C (**Scheme 16**). *o*-Bromo alkene derivatives **52b-z** were in turn prepared in high yields via Wittig or Julia olefination of the respective benzaldehydes by following the literature procedures.²⁸



Scheme 16: (i) CuCN (3 equiv), DMF reflux, 18 h.

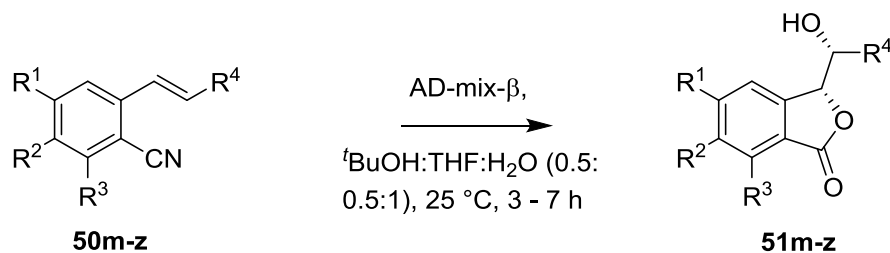
When subjected to Os-catalyzed asymmetric dihydroxylation (ADH) using (DHQD)₂PHAL as the chiral ligand, *o*-cyano α,β -unsaturated esters **50b-l** gave the corresponding chiral phthalide derivatives **51b-l** in 92-95% yields with excellent enantioselectivities (98-99% ee). Results of such studies are presented in **Table 1**. As can be seen, in every case, the reaction proceeded rapidly within 7 h giving the desired phthalides **51b-l** in excellent yields and ees (up to 99%) at ambient conditions. For instance, substrates having halogen (entry i), highly electron-rich (entry f) or electron-deficient (entry j) groups on the aromatic nucleus including 2-naphthyl system (entry l) underwent this oxidative cyclization smoothly affording the corresponding phthalides **51b-l** with excellent yields in one step.

Subsequently, we extended our study to include other styrene derivatives **50m-z** bearing different functionalities on the aromatic nucleus as well as on the β -position of the styrene derivative side chain (R^4) (**Table 2**). It was again found that this ADH process displayed a wide substrate scope tolerating alkyl, aryl, alkoxy, fluoro or tosyl groups. Excellent yields of phthalide derivatives **51m-z** (93-95%) and enantioselectivities (97-99% ee) were indeed realized in all the cases studied. The stereochemistry of the cyclized products was assigned according to the previously established absolute configuration of phthalides as well as in accordance with ADH rules.²⁹

Table 1: CN-assisted Os-catalyzed oxidative cyclization of cyano ethyl cinnamates

entry	R ¹	R ²	R ³	yield (%) ^a	ee (%) ^{b,c}
a	H	H	H	94	99
b	OMe	H	H	95	99
c	OMe	OMe	H	94	99
d	H	OMe	OMe	94	99
e	OMe	H	OMe	94	99
f	OMe	OMe	OMe	92	99
g	OTs	OMe	H	93	99
h	OBn	OMe	H	94	99
i	F	H	H	94	99
j	NO ₂	H	H	93	99
k	-O-CH ₂ -O-		H	95	98
l	<i>(E)</i> -ethyl 3-(1-cyanonaphthalen-2-yl)acrylate			94	98

^a Isolated yield after column chromatographic purification. ^b ee determined by chiral HPLC analysis. ^c ee determined by Mosher's ester analysis for entries h, i & l.

Table 2: CN-assisted Os-catalyzed oxidative cyclization of cyano styrene derivatives

entry	R ¹	R ²	R ³	R ⁴	yield (%) ^a	ee (%) ^{b,c,d}
m	H	H	H	H	95	99
n	OMe	H	H	H	95	99
o	OMe	OMe	H	H	93	99
p	H	OMe	OMe	H	94	99
q	OMe	H	OMe	H	94	99
r	OMe	OMe	OMe	H	92	99
s	OTs	OMe	H	H	93	99
t	OBn	OMe	H	H	94	99
u	F	H	H	H	94	99
v	-O-CH ₂ -O-		H	H	93	99
w	H	H	H	C ₃ H ₇	93	97
x	OMe	OMe	H	CH ₂ OTBS	94	97
y	OMe	OMe	H	Ph	94	97
z	OMe	OMe	H	<i>n</i> -C ₆ H ₁₃	92	98

^a Isolated yield after column chromatographic purification. ^b ee determined by chiral HPLC analysis.

^c ee determined by Mosher's ester analysis for entries t, u, w-z. ^d reaction completed in 3 h for m-v.

The formation of phthalide derivatives **51m-z** were confirmed by ¹H and ¹³C NMR spectroscopy. For example: The formation of phthalide **51m** was confirmed by the

appearance of typical signals at δ 3.90 (d, $J = 11.8$ Hz, 1H), 4.14 (d, $J = 11.8$ Hz, 1H) and 5.54-5.59 (m, 1H) due to diastereotopic methylene protons and benzylic proton respectively, in its ^1H NMR spectrum. Further, its ^{13}C NMR spectrum showed characteristic signals at δ 61.7, 81.5 and 170.6 corresponding to carbons attached to oxygen atoms and carbonyl carbon of lactone respectively (**Fig. 9**). Its IR spectrum displayed a strong absorption at 1756 cm^{-1} indicating the presence of lactone carbonyl group.

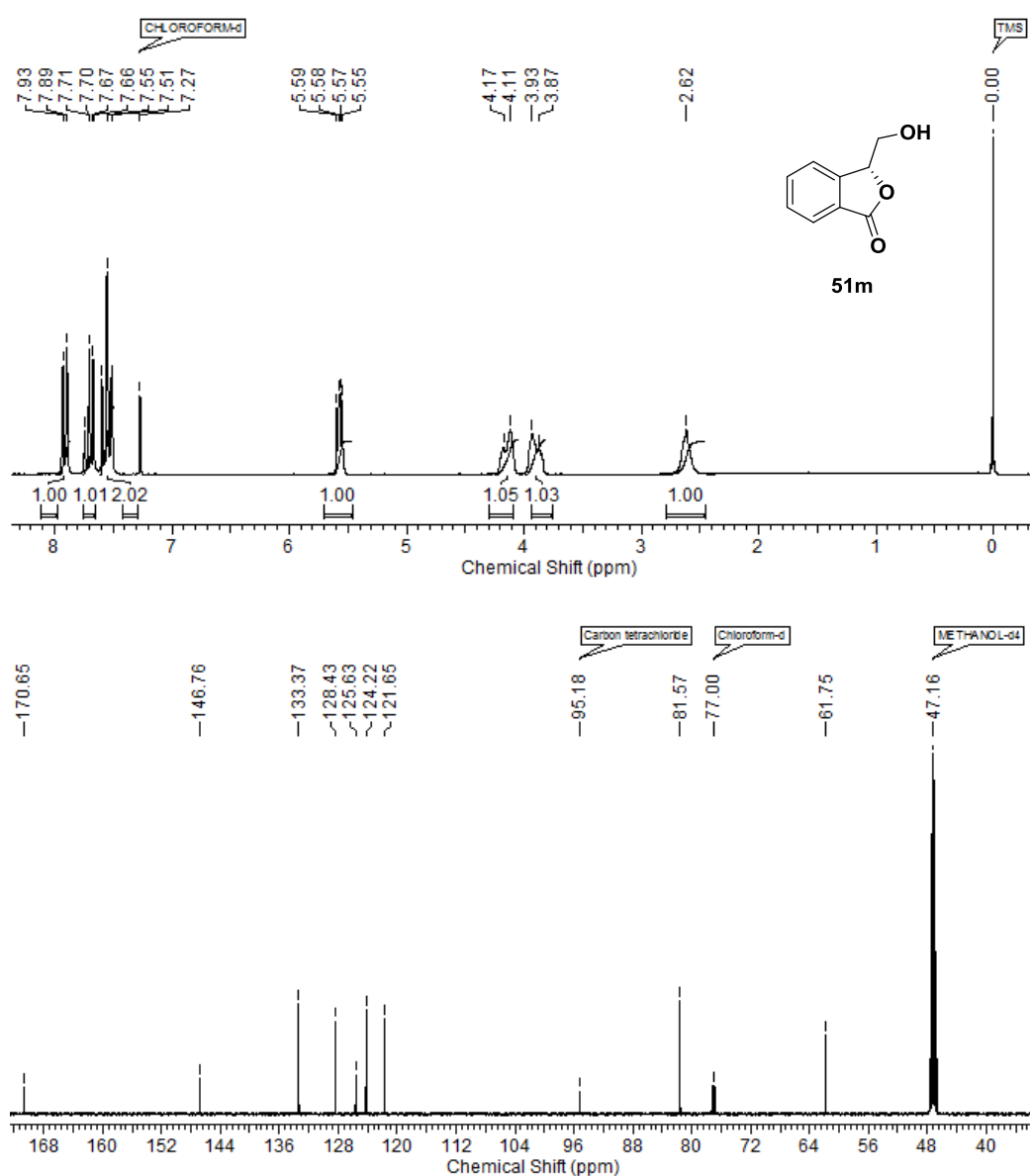


Fig. 9: ^1H and ^{13}C NMR spectra of phthalide **51m**

The enantiomeric excess of chiral phthalides **51m-z** was determined by chiral HPLC analysis and also by Mosher's ester analysis. For example: The enantiomeric excess of chiral phthalide **51n** was determined as 99% from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 1 mL/min) retention time 27.19 min (99.36%) and 39.72 min (0.64%) (**Fig. 10**).

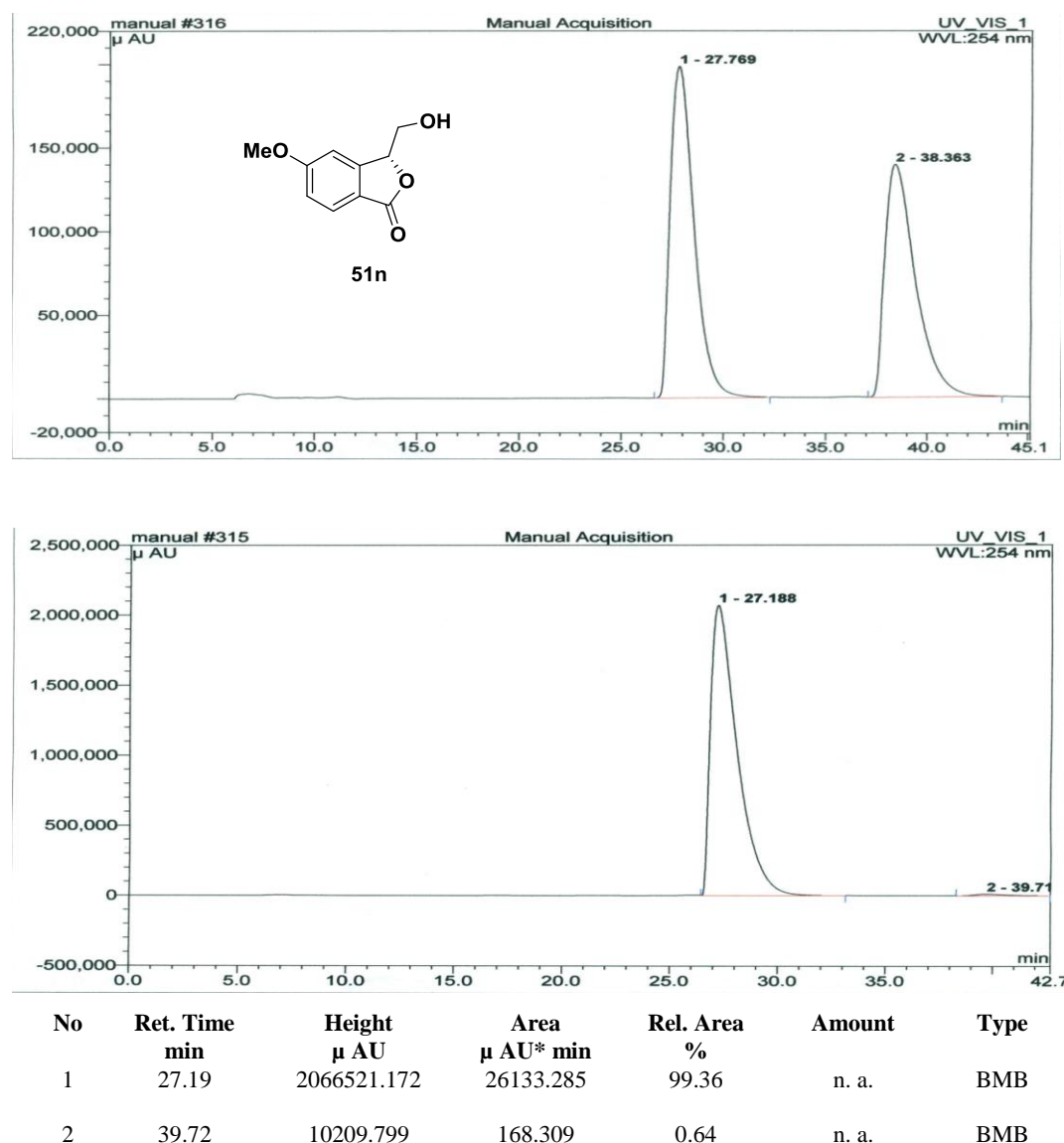


Fig. 10: HPLC chromatogram of phthalide **51n**

The higher reactivity of cyano substituted cinnamates and styrenes **50a-z** were substantiated by carrying out several competitive experiments involving 1:1 molar

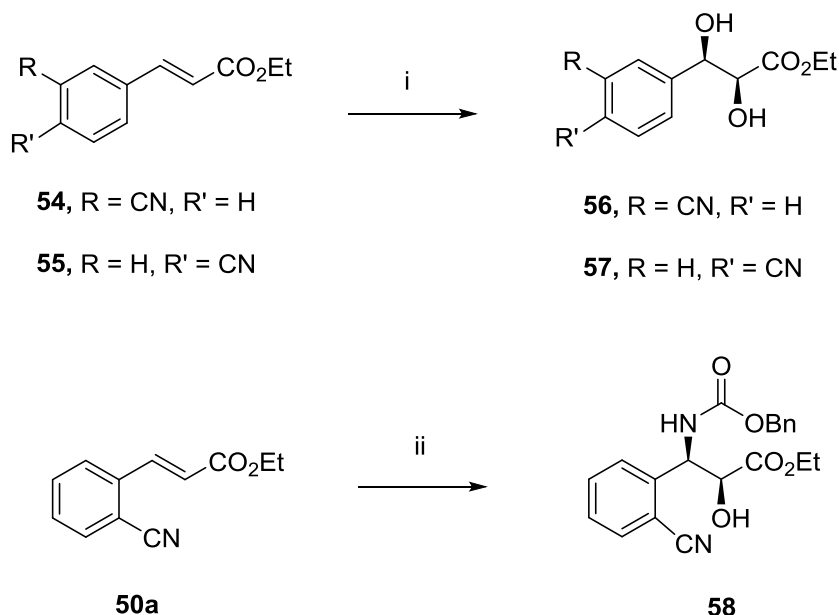
equivalents of aromatic substrates with or without cyano substitution; the results of which are presented in **Table 3**. The results clearly showed that cyano substituted substrates reacted almost 10-12 times faster than the one without cyano substitution, giving excellent yields of phthalides (92-94%).

Table 3: Competitive experiments^a

entry	substrates	product	yield (%) ^{b,c}
1	50a + ethyl cinnamate	51a	92
2	50e + 3,5-dimethoxyethyl cinnamate	51e	93
3	50m + styrene	51m	94
4	50o + 3,5-dimethoxystyrene	51o	92
5	50r + 3,4,5-trimethoxystyrene	51r	92

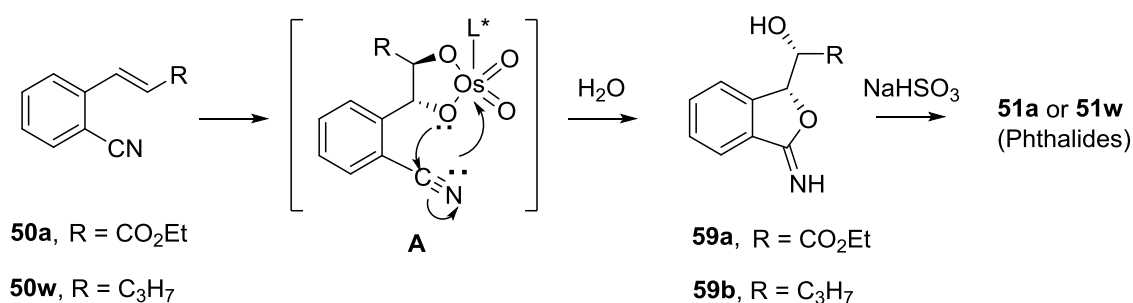
^a1:1 molar equivalents of aromatics substrates with and without cyano substitution (1 mmol each) AD-mix- β (0.5 mol%), *tert*-BuOH:THF:H₂O (0.5: 0.5:1), 25 °C, 3 h for entries 3-5 and 7 h for entries 1 and 2. ^bIsolated yields after column chromatographic purification. ^c5-8 % of 1, 2-diol from the corresponding substrates without cyano substitution was indeed isolated.

In order to account for the mechanistic course of the reaction, the following experiments (**Scheme 17**) were conducted: (i) AD-mix- β of substrates **54** & **55** for 36 h gave the corresponding cyanodiols **56** & **57** respectively, indicating that both CN and C=C groups must be positioned in proximity for CN coordination assistance to take place; (ii) asymmetric *aminohydroxylation*³⁰ of **50a** gave the expected amino alcohol **58** (64%) with no phthalide formation, suggesting that coordination of CN onto imino osmate ester is thermodynamically less favorable, due to its reduced Lewis acid character;³¹ (iii) in addition, imino intermediates **59a-b** were indeed isolated in 20% yield during the AD-mix- β of substrates **50a** and **50 w**.



Scheme 17: (i) $\text{K}_2[\text{OsO}_2(\text{OH})_4]$ (0.1 mol%), $(\text{DHQD})_2\text{PHAL}$ (0.5 mol%), $\text{K}_3\text{Fe}(\text{CN})_6$ (3 equiv), K_2CO_3 (3 equiv), *tert*-BuOH:THF:H₂O (1:1:2), 36 h, 94-95%; (ii) BnOCONH_2 , aq. NaOH, $\text{K}_2[\text{OsO}_2(\text{OH})_4]$, $(\text{DHQD})_2\text{PHAL}$, *tert*-BuOCl, ⁿPrOH:H₂O, 3 h, 64%, dr = 6:1.

This study clearly excludes the hydrolysis of CN to CO₂H followed by cyclization route, (iv) addition of benzonitrile as an external source of CN-assistance resulted in no rate enhancement for the ADH process.



Scheme 18: Mechanism of CN-assisted Os-catalyzed oxidative cyclization

On the basis of these results, a mechanistic model is presented in species **A** in which a synergism involving co-ordination of CN to Os(VI) and concurrent attack of osmate ester onto electropositive carbon of CN is shown that probably helps to accelerate the hydrolysis of osmate ester. These results indicate the 5-*exo-dig* type cyclization³² to

afford iminoesters **59a-b**, which finally lead to the formation of phthalides **51a** or **51w** (Scheme 18). The formation of iminoester **59b** was clearly demonstrated by IR data of the nonsubstituted imidate C=NH band (at 1687 cm^{-1}) and the phthalide (**51w**) C=O band (at 1752 cm^{-1}) (Fig. 11).

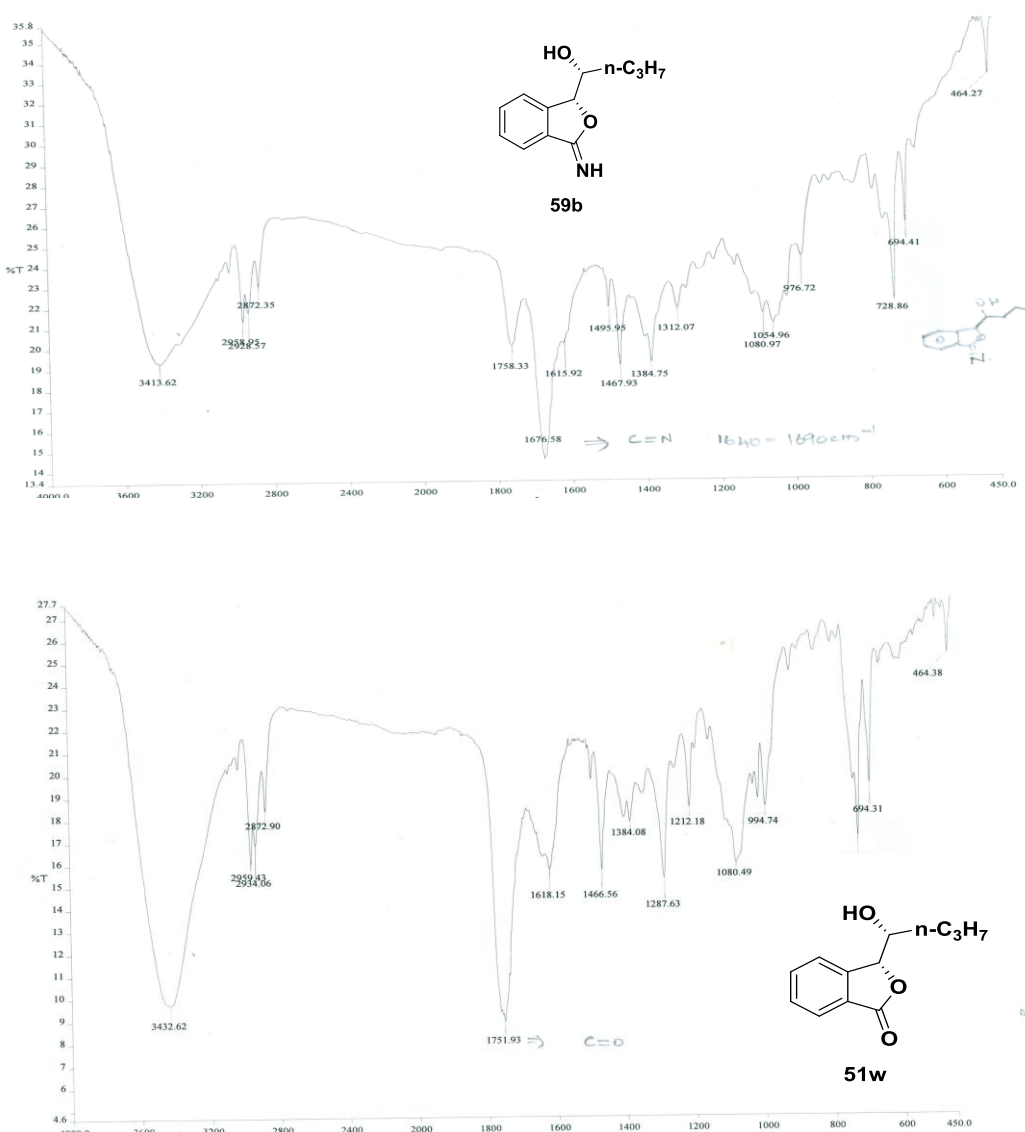


Fig. 11: IR spectra of iminoester **59b** and phthalide **51w**

2.1.5 Conclusion

A novel CN-assisted oxidative cyclization for the synthesis of a wide variety of 3-substituted phthalides and their structural analogues *via* ADH process of cyano cinnamates and styrene derivatives has been demonstrated. This reaction is highly practical in the sense that the products were obtained in excellent yields and optical purities (97-99% ee) and shows broad substrate scope and good functional group tolerance. The synergism shown by CN and osmate groups in proximity helps to enhance the rate of this reaction. We believe that this oxidative intramolecular cyclization ADH strategy should find wide applications in the total synthesis of other bioactive phthalide frameworks.

2.1.6 Experimental Section

Typical experimental procedure for the preparation of (*E*)-Ethyl 3-(2-cyano phenyl)acrylate (**50a**)

To a stirred solution of 2-cyanobenzaldehyde **49** (2 g, 7.9 mmol) in benzene (40 mL), $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$ (3.1 g, 8.6 mmol) was added. It was then refluxed for 12 h under N_2 atmosphere. After the completion of reaction, benzene was distilled out to give the crude product. Chromatographic purification of crude product [silica gel (230-400 mesh) and pet. ether:EtOAc (90:10) as eluent] afforded the cyano cinnamate **50a** (1.4 g).

Yield: 88%, colorless solid; **mp:** 60-62 °C; **IR** (CHCl_3 , cm^{-1}): ν_{max} 765, 784, 1031, 1184, 1318, 1447, 1480, 1594, 1640, 1712, 2225, 2938, 2983; **^1H NMR** (200 MHz, CDCl_3): δ 1.36 (t, $J = 7.3$ Hz, 3H), 4.31 (q, $J = 7.3$ Hz, 2H), 6.60 (d, $J = 16$ Hz, 1H), 7.47 (td, $J = 1.4, 7.5$ Hz, 1H), 7.62 (td, $J = 1.4, 7.5$ Hz, 1H), 7.70-7.76 (m, 2H), 7.96 (d, $J = 16$ Hz, 1H); **^{13}C NMR** (50 MHz, CDCl_3): δ 14.1, 60.7, 112.5, 116.8, 122.9, 126.8, 129.9, 132.8, 133.3, 137.1, 139.1, 165.4; **Analysis:** $\text{C}_{12}\text{H}_{11}\text{NO}_2$ requires C, 71.63; H, 5.51; N, 6.96; found: C, 71.59; H, 5.56; N, 6.93%.

Typical experimental procedure for the preparation of (S)-Ethyl 2-((R)-1,3-dihydro-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51a)

A 50 mL RB flask was charged with $\text{K}_3\text{Fe}(\text{CN})_6$ (1 g, 3 mmol), K_2CO_3 (414 mg, 3 mmol), *tert*-BuOH (2.5 mL), THF (2.5 mL) and H_2O (5 mL) and stirred for 10 min. Subsequently, $(\text{DHQD})_2\text{PHAL}$ (8 mg, 1 mol%) and $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$ (2 mg, 0.5 mol%) were added and the stirring continued for additional 30 min. To this reaction mixture, (*E*)-ethyl 3-(2-cyanophenyl)acrylate (**50a**) (200 mg, 1 mmol) was added and allowed to stir for 7 h at 25 °C. After completion of reaction (as monitored by TLC), sodium bisulphite (1 g) was added slowly at 0 °C. The organic layer was separated, aqueous layer extracted with ethyl acetate (3 x 10 ml) and the combined organic layers washed with brine (15 mL), dried over anhyd. Na_2SO_4 and concentrated under reduced pressure to yield the crude product. Flash column chromatographic purification [silica gel (230-400 mesh) and pet. ether:EtOAc (7:3) as an eluent] gave **51a** (221 mg).

Yield: 94%; colorless solid; **mp:** 146-148 °C; $[\alpha]_{25}^{\text{D}}$ -95.65 (*c* 1.24, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 12.16 (99.65%) and 13.80 (0.35%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 762, 856, 968, 1027, 1068, 1078, 1210, 1298, 1349, 1467, 1611, 1652, 1720, 1768, 2924, 3014, 3440; **^1H NMR** (200 MHz, CDCl_3): δ 1.29 (t, *J* = 7.1 Hz, 3H), 3.16 (d, *J* = 5.7 Hz, 1H), 4.30 (q, *J* = 7.1 Hz, 2H), 4.66 (dd, *J* = 2.1, 5.8 Hz, 1H), 5.79 (d, *J* = 2.1 Hz, 1H), 7.57 (t, *J* = 7.0 Hz, 2H), 7.68-7.75 (m, 1H), 7.90-7.93 (m, 1H); **^{13}C NMR** (50 MHz, CDCl_3): δ 13.7, 62.3, 70.3, 80.3, 122.0, 125.3, 126.4, 129.3, 134.0, 145.7, 169.8, 170.7; **HRMS** (ESI) *m/z* calcd for $\text{C}_{12}\text{H}_{12}\text{O}_5$ [$\text{M} + \text{Na}$] $^+$: 259.1357, found: 259.1352; **Analysis:** $\text{C}_{12}\text{H}_{12}\text{O}_5$ requires C, 61.01; H, 5.12; found: C, 60.96; H, 5.07%.

General experimental procedure for the preparation of *o*-cyanoalkenes (50b-z)

o-Bromo alkenes **52b-z** (1 mmol) were taken in dry DMF (10 mL) and CuCN (3 mmol) was added and the mixture refluxed under N₂ for 18 h (monitored by TLC). It was then cooled to 25 °C, and then diluted with water (30 mL) and EtOAc (25 mL). The organic layer was separated and the aqueous layer was extracted with EtOAc (3 x 20 mL). The combined organic extracts were washed with brine and dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to give crude products which were purified by column chromatography [silica gel (230-400 mesh) and pet. ether:EtOAc (7:3) as an eluent] to give *o*-cyanoalkenes **50b-z** in 76-84% yield.

(*E*)-Ethyl 3-(2-cyano-5-methoxyphenyl)acrylate (50b)

Yield: 86%, colorless solid; **mp:** 130-132 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 728, 868, 1026, 1256, 1490, 1594, 1607, 1640, 1712, 2228, 2853, 2923 3023; **¹H NMR** (200 MHz, CDCl₃): δ 1.36 (t, *J* = 7 Hz, 3H), 3.90 (s, 3H), 4.29 (q, *J* = 7 Hz, 2H), 6.56 (d, *J* = 16 Hz, 1H), 6.97 (dd, *J* = 2.5, 8.7 Hz, 1H), 7.15 (d, *J* = 2.5 Hz, 1H), 7.63 (d, *J* = 8.7 Hz, 1H), 7.90 (d, *J* = 16 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.2, 55.6, 60.8, 104.6, 112.1, 116.0, 117.3, 123.1, 135.0, 139.4, 162.7, 165.5; **Analysis:** C₁₃H₁₃NO₃ requires C, 67.52; H, 5.67; N, 6.06; found: C, 67.49; H, 5.61; N, 6.01%.

(*E*)-Ethyl 3-(2-cyano-4,5-dimethoxyphenyl)acrylate (50c)

Yield: 87%, colorless solid; **mp:** 159-161 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 761, 848, 1094, 1149, 1204, 1326, 1462, 1571, 1594, 1709, 2222, 2984, 3018; **¹H NMR** (200 MHz, CDCl₃): δ 1.36 (t, *J* = 7.3 Hz, 3H), 3.94 (s, 3H), 3.97 (s, 3H), 4.29 (q, *J* = 7.3 Hz, 2H), 6.47 (d, *J* = 16 Hz, 1H), 7.07 (s, 1H), 7.11 (s, 1H), 7.89 (d, *J* = 16 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.2, 55.9, 56.2, 60.7, 105.2, 108.2, 114.2, 117.1, 120.7, 131.5, 139.2, 150.5, 152.6, 165.8; **Analysis:** C₁₄H₁₅NO₄ requires C, 64.36; H, 5.79; N, 5.36; found: C, 64.32; H, 5.71; N, 5.34%.

(E)-Ethyl 3-(2-cyano-3,4-dimethoxyphenyl)acrylate (50d)

Yield: 88%, colorless solid; **mp:** 145-147 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 758, 894, 1078, 1138, 1208, 1318, 1326, 1462, 1571, 1594, 1608, 1710, 2222, 2984, 3018; **¹H NMR** (200 MHz, CDCl₃): δ 1.35 (t, $J = 7.0$ Hz, 3H), 3.93 (s, 3H), 4.03 (s, 3H), 4.27 (q, $J = 7.0$ Hz, 2H), 6.48 (d, $J = 16$ Hz, 1H), 7.10 (d, $J = 8.6$ Hz, 1H), 7.40 (d, $J = 8.6$ Hz, 1H), 7.83 (d, $J = 16$ Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.3, 56.1, 60.6, 61.6, 107.9, 114.1, 116.4, 120.7, 122.9, 129.7, 139.2, 152.1, 153.5, 165.9; **Analysis:** C₁₄H₁₅NO₄ requires C, 64.36; H, 5.79; N, 5.36; found: C, 64.34; H, 5.71; N, 5.32%.

(E)-Ethyl 3-(2-cyano-3,5-dimethoxyphenyl)acrylate (50e)

Yield: 87%, colorless solid; **mp:** 119-122 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 734, 876, 1069, 1128, 1208, 1326, 1326, 1478, 1568, 1594, 1608, 1712, 2228, 2958, 3082; **¹H NMR** (200 MHz, CDCl₃): δ 1.36 (t, $J = 7.1$ Hz, 3H), 3.89 (s, 3H), 3.93 (s, 3H), 4.29 (q, $J = 7.1$ Hz, 2H), 6.47 (d, $J = 2.1$ Hz, 1H), 6.55 (d, $J = 16$ Hz, 1H), 6.73 (d, $J = 2.1$ Hz, 1H), 7.86 (d, $J = 16$ Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.3, 55.7, 56.1, 60.8, 94.9, 96.1, 99.4, 103.4, 114.8, 123.3, 139.6, 140.1, 163.4, 163.9, 165.6; **Analysis:** C₁₄H₁₅NO₄ requires C, 64.36; H, 5.79; N, 5.36; found: C, 64.32; H, 5.71; N, 5.34%.

(E)-Ethyl 3-(2-cyano-3,4,5-trimethoxyphenyl)acrylate (50f)

Yield: 88%, colorless solid; **mp:** 150-152 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 669, 703, 749, 940, 1260, 1311, 1573, 1607, 1640, 1708, 2210, 2979, 3016; **¹H NMR** (200 MHz, CDCl₃): δ 1.36 (t, $J = 7.1$ Hz, 3H), 3.90 (s, 3H), 3.96 (s, 3H), 4.06 (s, 3H), 4.28 (q, $J = 7.1$ Hz, 2H), 6.50 (d, $J = 16$ Hz, 1H), 6.91 (s, 1H), 7.84 (d, $J = 16$ Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.3, 55.8, 60.3, 109.1, 115.4, 117.0, 118.5, 126.2, 142.5, 148.5, 151.1, 161.2; **Analysis:** C₁₅H₁₇NO₅ requires C, 61.85; H, 5.88; N, 4.81; found: C, 61.82; H, 5.79; N, 4.75%.

5-((E)-2-(Ethoxycarbonyl)vinyl)-4-cyano-2-methoxyphenyl 4-methylbenzene sulfonate (50g)

Yield: 87%, colorless solid; **mp:** 150-151 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 742, 865, 1030, 1128, 1232, 1318, 1329, 1478, 1571, 1594, 1608, 1708, 2225, 2982, 3025; **¹H NMR** (200 MHz, CDCl₃): δ 1.35 (t, *J* = 6.9 Hz, 3H), 2.48 (s, 3H), 3.73 (s, 3H), 4.30 (q, *J* = 6.9 Hz, 2H), 6.54 (d, *J* = 16 Hz, 1H), 7.09 (s, 1H), 7.35 (d, *J* = 8.5 Hz, 2H), 7.39 (s, 1H), 7.77 (d, *J* = 8.5 Hz, 2H), 7.91 (d, *J* = 16 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.2, 21.7, 56.0, 61.0, 104.5, 110.3, 116.0, 123.8, 128.3, 129.7, 132.6, 137.9, 138.4, 139.1, 145.8, 155.6, 165.2; **Analysis:** C₂₀H₁₉NO₆S requires C, 59.84; H, 4.77; N, 3.49; found: C, 59.78; H, 4.69; N, 3.42%.

(E)-Ethyl 3-(5-(benzyloxy)-2-cyano-4-methoxyphenyl)acrylate (50h)

Yield: 86%, colorless solid; **mp:** 146-148 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 738, 825, 1031, 1098, 1234, 1334, 1380, 1467, 1568, 1575, 1608, 1710, 2228, 2982, 3034; **¹H NMR** (200 MHz, CDCl₃): δ 1.35 (t, *J* = 7.2 Hz, 3H), 3.93 (s, 3H), 4.28 (q, *J* = 7.3 Hz, 2H), 5.20 (s, 2H), 6.34 (d, *J* = 15.4 Hz, 1H), 7.08 (s, 2H), 7.34-7.43 (m, 5H), 7.84 (d, *J* = 15.4 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.3, 56.2, 60.8, 71.0, 105.5, 110.5, 114.7, 117.2, 120.9, 127.3, 128.8, 131.5, 135.4, 139.3, 151.1, 151.8, 165.9; **Analysis:** C₂₀H₁₉NO₄ requires C, 71.20; H, 5.68; N, 4.15; found: C, 71.14; H, 5.61; N, 4.09%.

(E)-Ethyl 3-(2-cyano-5-fluorophenyl)acrylate (50i)

Yield: 85%, colorless solid; **mp:** 72-74 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 828, 866, 981, 1030, 1186, 1226, 1276, 1325, 1370, 1480, 1574, 1603, 1640, 1693, 2984, 3012; **¹H NMR** (200 MHz, CDCl₃): δ 1.36 (t, *J* = 7.2 Hz, 3H), 4.31 (q, *J* = 7.2 Hz, 2H), 6.58 (d, *J* = 15.9 Hz, 1H), 7.19 (td, *J* = 2.8, 8.4 Hz, 1H), 7.41 (dd, *J* = 2.6, 9.1 Hz, 1H), 7.74 (dd, *J* = 5.4, 8.4 Hz, 1H), 7.91 (d, *J* = 15.9 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.1, 60.9, 108.8, 133.9 (d, *J* = 23.6 Hz), 116.0, 117.6 (d, *J* = 23.6 Hz), 124.2, 135.7

(d, $J = 9.8$ Hz), 140.2 (d, $J = 8.8$ Hz), 164.5 (d, $J = 257.7$ Hz), 164.9; **Analysis:** $C_{12}H_{10}FNO_2$ requires C, 65.75; H, 4.60; N, 6.39; found: C, 65.68; H, 4.56; N, 6.36%.

(E)-Ethyl 3-(2-cyano-5-nitrophenyl)acrylate (50j)

Yield: 87%, colorless solid; **mp:** 105-107 °C; **IR** ($CHCl_3$, cm^{-1}): ν_{max} 739, 830, 968, 1032, 1106, 1346, 1540, 1708, 2233, 2980, 3087; **1H NMR** (500 MHz, $CDCl_3$): δ 1.38 (t, $J = 7.0$ Hz, 3H), 4.33 (q, $J = 7.0$ Hz, 2H), 6.78 (d, $J = 15.8$ Hz, 1H), 7.95 (d, $J = 2.0$ Hz, 1H), 7.98 (d, $J = 15.8$ Hz, 1H), 8.33 (dd, $J = 2.0, 8.4$ Hz, 1H) 8.59 (d, $J = 2.0$ Hz, 1H); **^{13}C NMR** (125 MHz, $CDCl_3$): δ 14.2, 61.3, 115.2, 117.8, 121.7, 124.1, 125.9, 134.7, 136.9, 139.5, 150.2, 164.8; **Analysis:** $C_{12}H_{10}N_2O_4$ requires C, 58.54; H, 4.09; N, 11.38; found: C, 58.48; H, 4.02; N, 11.31%.

(E)-Ethyl 3-(5-cyanobenzo[d][1,3]dioxol-6-yl)acrylate (50k)

Yield: 86%, colorless solid; **mp:** 148-149 °C; **IR** ($CHCl_3$, cm^{-1}): ν_{max} 728, 878, 1042, 1134, 1256, 1366, 1382, 1478, 1568, 1594, 1608, 1712, 2218, 2958, 3082; **1H NMR** (200 MHz, $CDCl_3$): δ 1.35 (t, $J = 7$ Hz, 3H), 4.28 (q, $J = 7$ Hz, 2H), 6.12 (s, 2H), 6.41 (d, $J = 15.8$ Hz, 1H), 7.05 (s, 1H), 7.13 (s, 1H), 7.90 (d, $J = 15.8$ Hz, 1H); **^{13}C NMR** (50 MHz, $CDCl_3$): δ 14.3, 60.8, 102.8, 105.9, 106.8, 111.8, 116.9, 121.4, 133.9, 138.9, 149.2, 151.9, 165.7; **Analysis:** $C_{13}H_{11}NO_4$ requires C, 63.67; H, 4.52; N, 5.71; found: C, 63.59; H, 4.48; N, 5.65%.

(E)-Ethyl 3-(1-cyanonaphthalen-2-yl)acrylate (50l)

Yield: 88%, colorless solid; **mp:** 118-119 °C; **IR** ($CHCl_3$, cm^{-1}): ν_{max} 784, 865, 989, 1030, 1106, 1210, 1275, 1291, 1319, 1368, 1573, 1607, 1712, 2218, 2978, 3084; **1H NMR** (200 MHz, $CDCl_3$): δ 1.35 (t, $J = 7.0$ Hz, 3H), 4.32 (q, $J = 7.0$ Hz, 2H), 6.68 (d, $J = 16$ Hz, 1H), 7.59-7.78 (m, 3H), 7.90 (d, $J = 7.7$ Hz, 1H), 8.04 (d, $J = 8.7$ Hz, 1H), 8.19 (d, $J = 16$ Hz, 1H), 8.28 (d, $J = 8.7$ Hz, 1H); **^{13}C NMR** (50 MHz, $CDCl_3$): δ 14.2, 60.8, 110.8, 115.5, 122.1, 123.5, 125.8, 128.3, 129.1, 132.5, 132.9, 137.0, 139.5,

165.5; **Analysis:** C₁₆H₁₃NO₂ requires C, 76.48; H, 5.21; N, 5.57; found: C, 76.42; H, 5.19; N, 5.52%.

2-Vinylbenzotrile (50m)

Yield: 86%, colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 752, 839, 962, 1014, 1072, 1118, 1202, 1308, 1347, 1368, 1444, 1573, 1607, 1625, 1675, 2215, 2889, 2923, 3012; **¹H NMR** (200 MHz, CDCl₃): δ 5.54 (d, J = 10.6 Hz, 1H), 5.95 (d, J = 17.8 Hz, 1H), 7.08 (dd, J = 10.6, 17.8 Hz, 1H), 7.34 (td, J = 1.2, 7.5 Hz, 1H), 7.51-7.70 (m, 3H); **¹³C NMR** (50 MHz, CDCl₃): δ 111.0, 117.4, 118.7, 125.2, 127.8, 132.5, 132.7, 140.4; **Analysis:** C₉H₇N requires C, 83.69; H, 5.46; N, 10.84; found: C, 83.62; H, 5.41; N, 10.78%.

4-Methoxy-2-vinylbenzotrile (50n)

Yield: 84%, colorless gum, **IR** (CHCl₃, cm⁻¹): ν_{\max} 752, 839, 1030, 1083, 1119, 1256, 1308, 1347, 1368, 1456, 1573, 1607, 1625, 1668, 2208, 2923, 3081; **¹H NMR** (200 MHz, CDCl₃): δ 3.88 (s, 3H), 5.53 (d, J = 11.1 Hz, 1H), 5.92 (d, J = 17.7 Hz, 1H), 6.85 (dd, J = 2.3, 8.5 Hz, 1H), 7.03 (dd, J = 11.1, 17.7 Hz, 1H), 7.11 (s, 1H), 7.55 (d, J = 8.5 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.4, 103.2, 110.4, 114.1, 117.9, 118.7, 132.9, 134.4, 142.5, 162.7; **Analysis:** C₁₀H₉NO requires C, 75.45; H, 5.70; N, 8.80; found: C, 75.41; H, 5.67; N, 8.73%.

4,5-Dimethoxy-2-vinylbenzotrile (50o)

Yield: 88%, colorless solid; **mp:** 106-107 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 752, 839, 936, 1031, 1086, 1119, 1256, 1308, 1378, 1389, 1456, 1575, 1612, 1628, 1656, 2210, 2923, 3052; **¹H NMR** (200 MHz, CDCl₃): δ 3.91 (s, 3H), 3.97 (s, 3H), 5.45 (d, J = 11.0 Hz, 1H), 5.80 (d, J = 17.2 Hz, 1H), 6.94-7.08 (m, 3H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.7, 55.9, 102.7, 106.9, 113.5, 116.5, 117.7, 132.5, 134.9, 148.7, 152.5;

Analysis: C₁₁H₁₁NO₂ requires C, 69.83; H, 5.86; N, 7.40; found: C, 69.75; H, 5.75; N, 7.39%.

2,3-Dimethoxy-6-vinylbenzotrile (50p)

Yield: 86%, colorless solid; **mp:** 108-110 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 748, 840, 936, 1028, 1086, 1119, 1256, 1308, 1378, 1389, 1456, 1575, 1612, 1628, 1656, 2202, 2981, 3029; **¹H NMR** (200 MHz, CDCl₃): δ 3.88 (s, 3H), 3.90 (s, 3H), 5.53 (d, J = 10.9 Hz, 1H), 5.90 (d, J = 17.3 Hz, 1H), 6.37 (d, J = 2.2 Hz, 1H), 6.69 (d, J = 2.2 Hz, 1H), 7.01 (dd, J = 10.9, 17.3 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 56.1, 61.5, 106.7, 114.7, 116.7, 120.8, 132.4, 133.4, 151.5, 151.7; **Analysis:** C₁₁H₁₁NO₂ requires C, 69.83; H, 5.86; N, 7.40; found: C, 69.73; H, 5.78; N, 7.39%.

2,4-Dimethoxy-6-vinylbenzotrile (50q)

Yield: 83%, colorless solid; **mp:** 76-79 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 724, 867, 968, 1030, 1086, 1119, 1259, 1308, 1386, 1389, 1456, 1578, 1612, 1636, 1656, 2212, 2985, 3029; **¹H NMR** (200 MHz, CDCl₃): δ 3.90 (s, 3H), 4.00 (s, 3H), 5.40 (d, J = 10.8 Hz, 1H), 5.79 (d, J = 17.6 Hz, 1H), 6.94 (dd, J = 10.8, 17.6 Hz, 1H), 7.07 (d, J = 8.5 Hz, 1H), 7.32 (d, J = 8.5 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.5, 55.9, 93.6, 97.5, 101.7, 115.5, 118.9, 133.1, 143.4, 163.0, 163.8; **Analysis:** C₁₁H₁₁NO₂ requires C, 69.83; H, 5.86; N, 7.40; found: C, 69.79; H, 5.78; N, 7.39%.

2,3,4-Trimethoxy-6-vinylbenzotrile (50 r)

Yield: 87%, colorless solid; **mp:** 102-103 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 771, 867, 1051, 1105, 1204, 1238, 1257, 1580, 1609, 1753, 2228, 2979, 3013; **¹H NMR** (200 MHz, CDCl₃): δ 3.86 (s, 3H), 3.95 (s, 3H), 4.04 (s, 3H), 5.48 (d, J = 11.2 Hz, 1H), 5.83 (d, J = 17.3 Hz, 1H), 6.85 (s, 1H), 6.97 (dd, J = 11.2, 17.3 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.9, 60.8, 61.5, 98.7, 103.4, 114.8, 117.8, 132.6, 137.2, 141.1, 155.4,

157.2; **Analysis:** C₁₂H₁₃NO₃ requires C, 65.74; H, 5.98; N, 6.39; found: C, 65.72; H, 5.91; N, 6.37%.

4-Cyano-2-methoxy-5-vinylphenyl 4-methylbenzenesulfonate (50s)

Yield: 82%, colorless solid; **mp:** 149-150 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 746, 845, 938, 1034, 1086, 1119, 1256, 1308, 1378, 1389, 1456, 1575, 1612, 1628, 1656, 2220, 2978, 3075; **¹H NMR** (200 MHz, CDCl₃): δ 2.48 (s, 3H), 3.74 (s, 3H), 5.57 (d, J = 10.9 Hz, 1H), 5.86 (d, J = 17.6 Hz, 1H), 6.93-7.08 (m, 2H), 7.28 (s, 1H), 7.35 (d, J = 8.0 Hz, 2H), 7.77 (d, J = 8.2 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 21.7, 55.8, 102.9, 108.9, 116.6, 119.7, 127.7, 128.5, 129.6, 132.3, 132.7, 137.7, 141.4, 145.6, 155.5; **Analysis:** C₁₇H₁₅NO₄S requires C, 61.99; H, 4.59; N, 4.25; found: C, 61.89; H, 4.53; N, 4.23%.

4-(Benzyloxy)-5-methoxy-2-vinylbenzonitrile (50t)

Yield: 84%, colorless solid; **mp:** 111-113 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 747, 858, 934, 1028, 1065, 1119, 1232, 1308, 1394, 1389, 1456, 1574, 1612, 1631, 1656, 2220, 2988, 3086; **¹H NMR** (200 MHz, CDCl₃): δ 3.90 (s, 3H), 5.21 (s, 2H), 5.39 (d, J = 11.1 Hz, 1H), 5.66 (d, J = 17.4 Hz, 1H), 6.89-7.04 (m, 2H), 7.10 (s, 1H), 7.32-7.47 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 56.0, 70.8, 103.1, 109.2, 114.0, 116.6, 117.8, 127.2, 128.2, 128.6, 132.6, 134.9, 135.7, 149.3, 151.8; **Analysis:** C₁₇H₁₅NO₂ requires C, 76.96; H, 5.70; N, 5.28; found: C, 76.91; H, 5.67; N, 5.27%.

4-Fluoro-2-vinylbenzonitrile (50u)

Yield: 86%, colorless solid; **mp:** 105-107 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 752, 839, 962, 1014, 1072, 1118, 1202, 1308, 1347, 1368, 1444, 1573, 1607, 1625, 1675, 2853, 2923, 3012; **¹H NMR** (500 MHz, CDCl₃): δ 5.62 (d, J = 11.0 Hz, 1H), 5.96 (d, J = 17.3 Hz, 1H), 7.02-7.08 (m, 2H), 7.34 (dd, J = 2.2, 9.4 Hz, 1H), 7.64 (dd, J = 5.5, 8.5 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 107.6, 112.6 (d, J = 23.7 Hz), 115.8 (d, J =

23.7 Hz), 116.8, 120.2, 132.2, 143.8 (d, $J = 9.5$ Hz), 165.1 (d, $J = 243.8$ Hz);

Analysis: C₉H₆FN requires C, 73.46; H, 4.11; N, 9.52; found: C, 73.44; H, 4.08; N, 9.49%.

6-Vinylbenzo[*d*][1,3]dioxole-5-carbonitrile (50v)

Yield: 88%, colorless solid; **mp:** 88-91 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 868, 930, 1038, 1162, 1263, 1359, 1486, 1505, 1604, 1615, 2219, 2916, 3018; **¹H NMR** (200 MHz, CDCl₃): δ 5.44 (d, $J = 11.1$ Hz, 1H), 5.77 (d, $J = 17.3$ Hz, 1H), 6.07 (s, 2H), 6.95-7.04 (m, 2H), 7.09 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 102.3, 104.0, 104.8, 110.9, 117.2, 117.7, 132.5, 137.5, 147.4, 151.8; **Analysis:** C₁₀H₇NO₂ requires C, 69.36; H, 4.07; N, 8.09; found: C, 69.34; H, 4.02; N, 7.99%.

2-((*E*)-Pent-1-enyl)benzotrile (50w)

Yield: 87%, colorless solid; **mp:** 126-128 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 857, 974, 1037, 1095, 1184, 1202, 1216, 1251, 1275, 1291, 1319, 1347, 1368, 1393, 1444, 1477, 1573, 1607, 1640, 1716, 2219, 2984, 3023; **¹H NMR** (200 MHz, CDCl₃): δ 0.98 (t, $J = 7.3$ Hz, 3H), 1.45-1.63 (m, 2H), 2.22-2.33 (m, 2H), 6.43 (dt, $J = 15.3, 6.8$ Hz, 1H), 6.74 (d, $J = 15.3$ Hz, 1H), 7.25 (dd, $J = 15.3, 1.4$ Hz, 1H), 7.45-7.62 (m, 3H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.7, 22.2, 35.2, 110.5, 117.9, 125.2, 126.0, 126.8, 132.5, 132.7, 136.4, 141.1; **Analysis:** C₁₂H₁₃N requires C, 84.17; H, 7.65; N, 8.18; found: C, 84.14; H, 7.61; N, 8.15%.

4,5-Dimethoxy-2-((*E*)-3-*tert*-butyldimethylsilyloxyprop-1-enyl)benzotrile (50x)

Yield: 85%, colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 748, 876, 932, 1032, 1098, 1276, 1339, 1486, 1505, 1604, 1615, 2220, 2989, 3054; **¹H NMR** (200 MHz, CDCl₃): δ 0.12 (s, 6H), 0.94 (s, 9H), 3.88 (s, 3H), 3.94 (s, 3H), 4.39 (dd, $J = 1.7, 4.7$ Hz, 2H), 6.27-6.39 (m, 1H), 6.89 (d, $J = 15.5$ Hz, 1H), 6.98 (s, 1H), 7.00 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ -5.3, 18.3, 25.9, 55.8, 55.9, 63.3, 102.6, 107.4, 113.7, 117.9, 124.9,

132.4, 134.8, 148.4, 152.5; **Analysis:** C₁₈H₂₇NO₃Si requires C, 64.83; H, 8.16; N, 4.20; found: C, 64.79; H, 8.09; N, 4.13%.

4,5-Dimethoxy-2-styrylbenzotrile (50y)

Yield: 83%, colorless solid; **mp:** 158-159 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 696, 761, 1149, 1204, 1326, 1462, 1571, 1594, 2215, 2984, 3023; **¹H NMR** (200 MHz, CDCl₃): δ 3.91 (s, 3H), 4.01 (s, 3H), 7.01-7.17 (m, 3H), 7.26-7.42 (m, 4H), 7.54 (d, $J = 6.9$ Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.9, 102.9, 106.8, 113.6, 118.0, 123.8, 126.7, 128.7, 128.6, 131.2, 134.9, 136.1, 148.5, 152.6; **Analysis:** C₁₇H₁₅NO₂ requires C, 76.96; H, 5.70; N, 5.28; found: C, 76.89; H, 5.57; N, 5.19%.

2,3,4-Trimethoxy-6-((E)-oct-1-enyl)benzotrile (50z)

Yield: 86%, colorless solid; **mp:** 172-174 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 694, 755, 878, 969, 989, 1097, 1216, 1271, 1452, 1464, 1513, 1600, 2220, 2970, 3025, 3059; **¹H NMR** (200 MHz, CDCl₃): δ 0.87-0.93 (m, 3H), 1.26-1.46 (m, 8H), 2.21-2.31 (m, 2H), 3.85 (s, 3H), 3.94 (s, 3H), 4.03 (s, 3H), 6.23-6.36 (m, 1H), 6.63 (d, $J = 15.6$ Hz, 1H), 6.78 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.1, 22.6, 28.9, 29.0, 31.6, 33.1, 56.0, 61.1, 61.6, 98.3, 103.3, 115.4, 125.8, 135.9, 138.1, 140.5, 155.6, 157.2; **Analysis:** C₁₈H₂₅NO₃ requires C, 71.26; H, 8.31; N, 4.62; found: C, 71.22; H, 8.28; N, 4.58%.

(E)-Ethyl 3-(3-cyanophenyl)acrylate (54)

Yield: 93%; colorless solid; **mp:** 62-65 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 710, 765, 977, 1032, 1185, 1278, 1318, 1447, 1480, 1640, 1712, 2225, 2938, 2983; **¹H NMR** (200 MHz, CDCl₃): δ 1.35 (d, $J = 7$ Hz, 3H), 4.28 (d, $J = 7$ Hz, 2H), 6.48 (d, $J = 16.1$ Hz, 1H), 7.48-7.80 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.1, 60.5, 113.2, 117.8, 120.8, 129.6, 131.1, 131.6, 132.8, 135.5, 141.5, 165.7; **Analysis:** C₁₂H₁₁NO₂ requires C, 71.63; H, 5.51; N, 6.96; found: C, 71.59; H, 5.45; N, 6.85%.

(E)-Ethyl 3-(4-cyanophenyl)acrylate (55)

Yield: 93%; colorless solid; **mp:** 68-70 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 730, 795, 955, 1065, 1194, 1268, 1375, 1445, 1495, 1652, 1721, 2226, 2983; **¹H NMR** (200 MHz, CDCl₃): δ 1.35 (d, $J = 7.1$ Hz, 3H), 4.28 (d, $J = 7.1$ Hz, 2H), 6.51 (d, $J = 15.8$ Hz, 1H), 7.59-7.71 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.1, 60.6, 113.2, 117.9, 121.6, 128.2, 132.4, 138.5, 141.8, 165.7; **Analysis:** C₁₂H₁₁NO₂ requires C, 71.63; H, 5.51; N, 6.96; found: C, 71.58; H, 5.48; N, 6.88%.

General experimental procedure for the preparation of chiral phthalides (51b-z)

A 50 mL RB flask was charged with K₃Fe(CN)₆ (3 mmol), K₂CO₃ (3 mmol), *tert*-BuOH (2.5 mL), THF (2.5 mL) and H₂O (5 mL) and stirred for 10 min. Subsequently, (DHQD)₂PHAL (1 mol%) and K₂OsO₄·2H₂O (0.5 mol%) were added and the stirring continued for additional 30 min. To this reaction mixture, *o*-cyanoalkenes **50b-z** (1 mmol) was added and allowed to stir for 3-7 h at 25 °C. After completion of reaction (as monitored by TLC), sodium bisulphite (1 g) was added slowly at 0 °C. The organic layer was separated, aqueous layer extracted with ethyl acetate (3 x 10 ml) and the combined organic layers washed with brine (15 mL), dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to yield the crude product. Flash column chromatographic purification [silica gel (230-400 mesh) and pet. ether:EtOAc (7:3) as an eluent] gave phthalides **51b-z** in 92-95% yield.

(S)-Ethyl 2-((R)-1,3-dihydro-5-methoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51b)

Yield: 95%; colorless solid; **mp:** 121-122 °C; $[\alpha]_{25}^D -94.49$ (c 1.15, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 1 mL/min) retention time 25.80 min (99.55%) and 30.33 min (0.45%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 724, 876, 1031, 1084, 1191, 1212, 1278, 1295, 1357, 1398, 1445, 1486, 1578, 1607, 1721, 1765, 2984, 3023, 3415; **¹H NMR** (200 MHz, CDCl₃): δ 1.29 (t, $J = 7.2$ Hz,

3H), 3.14 (br s, 3H), 3.91 (s, 3H), 4.29 (q, $J = 7.2$ Hz, 2H), 4.63 (d, $J = 1.7$ Hz, 1H), 5.69 (d, $J = 2.2$ Hz, 1H), 6.96 (d, $J = 2.1$ Hz, 1H), 7.05 (dd, $J = 2.1, 8.6$ Hz, 1H), 7.80 (d, $J = 8.6$ Hz, 1H); ^{13}C NMR (50 MHz, CDCl_3): δ 14.0, 55.7, 62.6, 70.5, 79.6, 106.0, 116.9, 118.9, 127.0, 148.5, 164.7, 169.5, 170.8; **Analysis:** $\text{C}_{13}\text{H}_{14}\text{O}_6$ requires C, 58.64; H, 5.30; found: C, 58.62; H, 5.19%.

(S)-Ethyl 2-((R)-1,3-dihydro-5,6-dimethoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51c)

Yield: 94%; colorless solid; **mp:** 144-146 °C; $[\alpha]_{25}^{\text{D}}$ -95.12 (c 1.12, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel OJ-H, n -hexane/ i PrOH, 90:10, 0.5 mL/min) retention time 23.18 min (99.36%) and 27.60 min (0.64%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 758, 945, 1125, 1297, 1507, 1722, 1764, 2925, 3010, 3341; ^1H NMR (200 MHz, CDCl_3): δ 1.30 (t, $J = 7.2$ Hz, 3H), 3.20 (d, $J = 6.2$ Hz, 1H), 3.94 (s, 3H), 3.98 (s, 3H), 4.29 (q, $J = 7.2$ Hz, 2H), 4.62 (dd, $J = 2.4, 6.1$ Hz, 1H), 5.66 (d, $J = 2.2$ Hz, 1H), 6.93 (s, 1H), 7.27 (s, 1H); ^{13}C NMR (50 MHz, $\text{DMSO}-d_6$): δ 14.3, 56.1, 56.3, 61.1, 70.1, 81.1, 105.2, 105.7, 118.2, 141.6, 150.4, 154.7, 170.2, 171.3; **Analysis:** $\text{C}_{14}\text{H}_{16}\text{O}_7$ requires C, 64.36; H, 5.79; N, 5.36; found: C, 64.32; H, 5.71; N, 5.34%.

(S)-Ethyl 2-((R)-1,3-dihydro-6,7-dimethoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (41d)

Yield: 94%, colorless solid; **mp:** 110-112 °C; $[\alpha]_{25}^{\text{D}}$ -95.28 (c 1.0, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel OJ-H, n -hexane/ i PrOH, 90:10, 0.5 mL/min) retention time 23.90 min (99.44%) and 27.87 min (0.56%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 762, 946, 1132, 1298, 1518, 1728, 1764, 2985, 3034, 3425; ^1H NMR (200 MHz, CDCl_3): δ 1.30 (t, $J = 7.1$ Hz, 3H), 3.19 (br s, 1H), 3.91 (s, 3H), 4.10 (s, 3H), 4.29 (q, $J = 7.1$ Hz, 2H), 4.57 (s, 1H), 5.65 (d, $J = 2.0$ Hz, 1H), 7.13 (d, $J = 8.1$ Hz, 1H), 7.23 (d, $J = 8.1$ Hz, 1H); ^{13}C NMR (50 MHz, CDCl_3): δ 14.0, 56.6, 62.2, 62.6, 70.7, 79.0,

116.4, 118.7, 119.2, 138.5, 148.3, 152.9, 167.2, 170.9; **Analysis:** C₁₄H₁₆O₇ requires C, 64.36; H, 5.79; found: C, 64.34; H, 5.71%.

(S)-Ethyl 2-((R)-1,3-dihydro-5,7-dimethoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51e)

Yield: 94%, colorless solid; **mp:** 154-156 °C; $[\alpha]_{25}^D$ -96.29 (*c* 1.15, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 18.37 min (99.60%) and 21.74 min (0.40%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 746, 985, 1130, 1287, 1514, 1723, 1762, 2954, 3085, 3414; **¹H NMR** (200 MHz, CDCl₃): δ 1.32 (t, *J* = 7.2 Hz, 3H), 3.37 (br s, 1H), 3.91 (s, 3H), 3.94 (s, 3H), 4.30 (q, *J* = 7.2 Hz, 2H), 4.61 (s, 1H), 5.67 (s, 1H), 6.47 (s, 1H), 6.59 (s, 1H); **¹³C NMR** (50 MHz, CD₃OD): δ 14.5, 56.5, 56.8, 62.9, 71.9, 82.1, 99.8, 100.2, 108.0, 153.0, 160.9, 168.9, 170.6, 172.4; **Analysis:** C₁₄H₁₆O₇ requires C, 64.36; H, 5.79; found: C, 64.34; H, 5.76%.

(S)-Ethyl 2-((R)-1,3-dihydro-5,6,7-trimethoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51f)

Yield: 92%, colorless solid; **mp:** 111-112 °C; $[\alpha]_{25}^D$ -94.65 (*c* 1.23, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 1012, 1094, 1140, 1254, 1350, 1475, 1602, 1765, 2954, 3085, 3408 cm⁻¹; **¹H NMR** (200 MHz, CDCl₃): δ 1.31 (t, *J* = 7.2 Hz, 3H), 3.09 (s, 1H), 3.86 (s, 3H), 3.96 (s, 3H), 4.13 (s, 3H), 4.31 (q, *J* = 7.2 Hz, 2H), 4.58 (d, *J* = 2.1 Hz, 1H), 5.58 (d, *J* = 2.1 Hz, 1H), 6.70 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.9, 56.3, 61.1, 62.0, 62.4, 79.1, 99.5, 111.0, 141.9, 143.5, 152.1, 159.6, 167.3, 176.7; **Analysis:** C₁₅H₁₈O₈ requires C, 55.21; H, 5.56; found: C, 55.18; H, 5.53%.

(S)-Ethyl 2-((R)-5-(*p*-toluenesulfonyloxy)-1,3-dihydro-6-methoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51g)

Yield: 93%, colorless solid; **mp:** 107-108 °C; $[\alpha]_{25}^D$ -94.89 (*c* 1.15, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 768, 819, 1025, 1050, 1120, 1180, 1190, 1330, 1374, 1494, 1614, 1767, 2924, 3012, 3371; **¹H NMR** (200 MHz, CDCl₃): δ 1.27 (t, *J* = 7.2 Hz, 3H), 2.48

(s, 3H), 3.07 (s, 1H), 3.78 (s, 3H), 4.26 (q, $J = 7.2$ Hz, 2H), 4.63 (d, $J = 2.1$ Hz, 1H), 5.67 (d, $J = 2.1$ Hz, 1H), 6.99 (s, 1H), 7.35 (d, $J = 8.1$ Hz, 2H), 7.49 (s, 1H), 7.76 (d, $J = 8.1$ Hz, 2H); ^{13}C NMR (50 MHz, DMSO- d_6): δ 14.4, 21.7, 56.6, 61.4, 70.3, 71.6, 81.3, 107.3, 118.6, 119.7, 128.6, 130.0, 132.5, 139.5, 145.9, 147.9, 156.9, 168.8, 170.9; **Analysis:** $\text{C}_{20}\text{H}_{20}\text{O}_9\text{S}$ requires C, 55.04; H, 4.62; found: C, 55.01; H, 4.59%.

(S)-Ethyl 2-((R)-5-(benzyloxy)-1,3-dihydro-6-methoxy-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51h)

Yield: 94%, colorless solid; **mp:** 138-140 °C; $[\alpha]_{25}^{\text{D}}$ -96.04 (c 1.21, CHCl_3); **IR** (CHCl_3 , cm^{-1}): ν_{max} 738, 856, 1025, 1078, 1130, 1184, 1195, 1336, 1395, 1494, 1645, 1765, 2942, 3035, 3413; **^1H NMR** (200 MHz, CDCl_3): δ 1.28 (t, $J = 7.0$ Hz, 3H), 3.04 (d, $J = 5.9$ Hz, 1H), 3.94 (s, 3H), 4.27 (q, $J = 7.0$ Hz, 2H), 4.55 (dd, $J = 2.5, 5.9$ Hz, 1H), 5.22 (d, $J = 3.5$ Hz, 2H), 5.61 (d, $J = 2.0$ Hz, 1H), 6.94 (s, 1H), 7.26 (s, 1H), 7.29-7.45 (m, 5H); ^{13}C NMR (50 MHz, DMSO- d_6): δ 13.7, 55.7, 61.4, 70.3, 70.6, 79.9, 105.2, 105.8, 118.5, 127.0, 127.8, 128.2, 135.3, 139.9, 150.7, 153.5, 169.6, 170.4; **Analysis:** $\text{C}_{20}\text{H}_{20}\text{O}_7$ requires C, 64.51; H, 5.41; found: C, 64.39; H, 5.36%.

(S)-Ethyl 2-((R)-5-fluoro-1,3-dihydro-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (50i)

Yield: 94%, colorless solid; **mp:** 108-109 °C; $[\alpha]_{25}^{\text{D}}$ -95.41 (c 1.15, CHCl_3); **IR** (CHCl_3 , cm^{-1}): ν_{max} 756, 891, 1052, 1097, 1130, 1190, 1325, 1374, 1485, 1629, 1765, 2928, 3015, 3351; **^1H NMR** (200 MHz, CDCl_3): δ 1.30 (t, $J = 7.2$ Hz, 3H), 3.17 (d, $J = 6.7$ Hz, 1H), 4.31 (q, $J = 7.2$ Hz, 2H), 4.62 (dd, $J = 2.2, 5.8$ Hz, 1H), 5.74 (d, $J = 7.2$ Hz, 1H), 7.23 (d, $J = 8.4$ Hz, 2H), 7.91 (dd, $J = 5.8, 8.3$ Hz, 1H); ^{13}C NMR (50 MHz, CDCl_3): δ 13.9, 62.2, 70.2, 79.7, 109.5 (d, $J = 24.6$ Hz), 117.6 (d, $J = 24.6$ Hz), 122.7, 127.7, 148.6 (d, $J = 10.3$ Hz), 166.3 (d, $J = 256.3$ Hz), 168.5, 170.5; **Analysis:** $\text{C}_{12}\text{H}_{11}\text{FO}_5$ requires C, 56.70; H, 4.36; found: C, 56.67; H 4.33%.

(S)-Ethyl 2-((R)-1,3-dihydro-5-nitro-1-oxoisobenzofuran-3-yl)-2-hydroxy acetate (51j)

Yield: 93%, colorless solid; **mp:** 146-148 °C; $[\alpha]_{25}^D$ -95.28 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 738, 829, 967, 1037, 1106, 1346, 1540, 1740, 1779, 2853, 2918, 3009, 3444; **¹H NMR** (500 MHz, CDCl₃): δ 1.35 (t, *J* = 7.2 Hz, 3H), 3.21 (d, *J* = 6.1 Hz, 1H), 4.36 (q, *J* = 7.2 Hz, 2H), 4.71 (d, *J* = 3.6 Hz, 1H), 5.90 (s, 1H), 8.09 (d, *J* = 8.2 Hz, 1H), 8.42-8.46 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃): δ 14.1, 63.2, 70.1, 80.1, 117.8, 125.3, 127.0, 131.8, 146.8, 151.7, 167.3, 170.2; **Analysis:** C₁₂H₁₁NO₇ requires C, 51.25; H, 3.94; N, 4.98; found: C, 51.24; H, 3.85; N, 4.93%.

(S)-Ethyl 2-((R)-5-1,3-dihydro-5,6-dioxomethyl-1-oxoisobenzofuran-3-yl)-2-hydroxyacetate (51k)

Yield: 95%, colorless solid; **mp:** 150-153 °C; $[\alpha]_{25}^D$ -95.74 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 786, 891, 1015, 1054, 1122, 1183, 1196, 1356, 1395, 1489, 1618, 1755, 2942, 3021, 3410; **¹H NMR** (200 MHz, CDCl₃): δ 1.31 (t, *J* = 7.1 Hz, 3H), 3.10 (br s, 1H), 4.30 (qd, *J* = 1.4, 7.1 Hz, 2H), 4.56 (s, 1H), 5.62 (d, *J* = 2.1 Hz, 1H), 6.14 (dd, *J* = 1.4, 4.4 Hz, 2H), 6.89 (s, 1H), 7.20 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.0, 62.6, 70.4, 79.6, 101.8, 102.7, 104.2, 120.4, 142.2, 149.6, 153.7, 169.2, 170.8; **Analysis:** C₁₃H₁₂O₇ requires C, 55.72; H, 4.32; found: C, 55.65; H, 4.29%.

(S)-Ethyl 2-((R)-1,3-dihydro-1-oxonaphtho[2,1-c]furan-3-yl)-2-hydroxyacetate (51l)

Yield: 94%, colorless solid; **mp:** 107-109 °C; $[\alpha]_{25}^D$ -95.69 (*c* 1.15, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 784, 865, 989, 1010, 1106, 1210, 1275, 1291, 1319, 1368, 1573, 1607, 1750, 2978, 3084, 3457; **¹H NMR** (200 MHz, CDCl₃): δ 1.28 (t, *J* = 7.4 Hz, 3H), 3.14 (d, *J* = 6.0 Hz, 1H), 4.31 (q, *J* = 7.4 Hz, 2H), 4.74 (dd, *J* = 2.1, 6.0 Hz, 1H), 5.85 (d, *J* = 2.1 Hz, 1H), 7.58 (d, *J* = 8.5 Hz, 1H), 7.63-7.78 (m, 2H), 7.97 (d, *J* = 8.5 Hz, 1H), 8.16 (d, *J* = 8.5 Hz, 1H) 8.97 (d, *J* = 8.5 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃+ CD₃OD): δ 13.2, 61.4, 69.8, 80.3, 118.1, 118.6, 120.2, 122.4, 126.7, 128.0,

128.3, 133.0, 135.2, 147.6, 170.3; **Analysis:** C₁₆H₁₄O₅ requires C, 67.13; H, 4.93; found: C, 67.11; H, 4.89%.

(R)-3-(Hydroxymethyl)isobenzofuran-1(3H)-one (51m)

Yield: 95%, colorless solid; **mp:** 101-104 °C; $[\alpha]_{25}^D$ -78.12 (*c* 1.23, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 8.03 (99.36%) and 9.24 (0.64%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 744, 847, 968, 1025, 1067, 1089, 1211, 1288, 1349, 1467, 1607, 1640, 1756, 2924, 3012, 3440; **¹H NMR** (200 MHz, CDCl₃): 2.61 (s, 1H), 3.90 (d, *J* = 11.8 Hz, 1H), 4.14 (d, *J* = 11.8 Hz, 1H), 5.54-5.59 (m, 1H), 7.55 (t, *J* = 7.7 Hz, 2H), 7.70 (td, *J* = 1.1, 7.4 Hz, 1H), 7.89 (d, *J* = 7.4 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃+CD₃OD): δ 61.7, 81.5, 121.6, 124.2, 125.6, 128.4, 133.3, 146.7, 170.6; **Analysis:** C₉H₈O₃ requires C, 65.85; H, 4.91; found: C, 65.83; H, 4.85%.

(R)-3-(Hydroxymethyl)-5-methoxyisobenzofuran-1(3H)-one (51n)

Yield: 95%, colorless solid; **mp:** 137-140 °C; $[\alpha]_{25}^D$ -78.36 (*c* 1.12, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 1 mL/min) retention time 27.19 min (99.36%) and 39.72 min (0.64%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 728, 868, 1026, 1256, 1490, 1607, 1640, 1749, 2853, 2923, 3440; **¹H NMR** (200 MHz, CDCl₃): δ 2.31 (br s, 1H), 3.84-3.91 (m, 4H), 4.06-4.14 (m, 1H), 5.46 (t, *J* = 5.3 Hz, 1H), 6.94 (d, *J* = 2.0 Hz, 1H), 7.04 (dd, *J* = 2.0, 8.6 Hz, 1H), 7.80 (d, *J* = 8.6 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃+CD₃OD): δ 54.8, 62.1, 81.0, 105.5, 116.3, 117.7, 126.0, 149.8, 164.5, 170.8; **Analysis:** C₁₀H₁₀O₄ requires C, 61.85; H, 5.19; found: C, 61.79; H, 5.12%.

(R)-3-(Hydroxymethyl)-5,6-dimethoxyisobenzofuran-1(3H)-one (51o)

Yield: 93%, colorless solid; **mp:** 165-167 °C; $[\alpha]_{25}^D$ -77.89 (*c* 1, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min)

retention time 23.18 min (99.36%) and 27.60 min (0.64%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 698, 828, 956, 1027, 1056, 1225, 1266, 1309, 1335, 1474, 1508, 1612, 1752, 2922, 3023, 3358; **^1H NMR** (200 MHz, CDCl_3): δ 2.71 (t, $J = 6.4$ Hz, 1H), 3.81-3.90 (m, 1H), 3.93 (s, 3H), 3.99 (s, 3H), 4.04-4.15 (m, 1H), 5.42-5.47 (m, 1H), 6.93 (s, 1H), 7.25 (s, 1H); **^{13}C NMR** (50 MHz, $\text{DMSO}-d_6$): δ 56.1, 56.3, 62.4, 81.6, 105.0, 105.8, 117.9, 142.4, 150.3, 154.6, 170.5; **Analysis**: $\text{C}_{11}\text{H}_{12}\text{O}_5$ requires C, 58.93; H, 5.39; found: C, 58.85; H, 5.37%.

(R)-3-(Hydroxymethyl)-6,7-dimethoxyisobenzofuran-1(3H)-one (51p)

Yield: 94%, colorless solid; **mp**: 85-88 °C; $[\alpha]_{25}^{\text{D}}$ -78.21 (c 1, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 18.35 min (99.35%) and 20.85 min (0.56%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 698, 798, 956, 1030, 1067, 1220, 1328, 1339, 1458, 1605, 1745, 2976, 3012, 3457; **^1H NMR** (200 MHz, CDCl_3): δ 2.24 (br s, 1H), 3.79-3.85 (m, 1H), 3.90 (s, 3H), 3.95 (s, 3H), 4.03-4.09 (m, 1H), 5.35-5.39 (m, 1H), 6.42 (s, 1H), 6.48(s, 1H); **^{13}C NMR** (50 MHz, CDCl_3): δ 56.6, 62.0, 63.7, 80.7, 116.8, 118.4, 119.4, 139.6, 148.0, 152.5, 168.2; **Analysis**: $\text{C}_{11}\text{H}_{12}\text{O}_5$ requires C, 58.93; H, 5.39; found: C, 58.83; H, 5.36%.

(R)-3-(Hydroxymethyl)-5,7-dimethoxyisobenzofuran-1(3H)-one (51q)

Yield: 94%, colorless solid; **mp**: 152-153 °C; $[\alpha]_{25}^{\text{D}}$ -78.1, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 18.27 min (99.36%) and 20.40 min (0.64%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 695, 765, 950, 1030, 1058, 1232, 1331, 1365, 1463, 1615, 1751, 2982, 3010, 3443; **^1H NMR** (200 MHz, CDCl_3): δ 2.53 (br s, 1H), 3.77-3.88 (m, 3H), 3.91 (s, 3H), 3.99-4.04 (m, 3H), 4.10 (s, 1H), 5.40-5.45 (m, 1H), 7.09 (dd, $J = 8.4, 8.2$ Hz, 1H), 7.22 (d, $J = 8.2$ Hz, 1H); **^{13}C NMR** (50 MHz, CDCl_3): δ 54.6, 54.9, 62.2, 80.4, 97.6, 98.2, 105.8,

151.7, 158.9, 166.6, 168.9; **Analysis:** C₁₁H₁₂O₅ requires C, 58.93; H, 5.39; found: C, 58.89; H, 5.37%.

(R)-3-(Hydroxymethyl)-5,6,7-trimethoxyisobenzofuran-1(3H)-one (51r)

Yield: 93%, colorless solid; **mp:** 178-180 °C; $[\alpha]_{25}^D$ -78.05 (*c* 1.15, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 1014, 1097, 1254, 1345, 1483, 1600, 1754, 2947, 3017, 3444; **¹H NMR** (200 MHz, CDCl₃): δ 2.62 (br s, 1H), 3.84-3.90 (m, 4H), 3.96 (s, 3H), 4.03-4.09 (m, 1H), 4.13 (s, 3H), 5.35-5.39 (m, 1H), 6.69 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃+ CD₃OD): δ 56.3, 61.1, 62.0, 63.7, 80.6, 99.9, 110.6, 141.8, 144.8, 152.1, 159.7, 168.3; **Analysis:** C₁₂H₁₄O₆ requires C, 56.09; H, 5.55; found: C, 56.05; H, 5.53%.

(R)-1,3-Dihydro-1-(hydroxymethyl)-5-methoxy-3-oxoisobenzofuran-6-yl 4-methylbenzenesulfonate (51s)

Yield: 95%, colorless solid; **mp:** 152-154 °C; $[\alpha]_{25}^D$ -77.79 (*c* 1.18, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 734, 849, 973, 103, 1053, 1178, 1345, 1372, 1494, 1614, 1755, 2919, 3018, 3437; **¹H NMR** (200 MHz, CDCl₃): δ 2.24 (br s, 1H), 2.48 (s, 3H), 3.80 (s, 3H), 3.92 (dd, *J* = 4.6, 12.3 Hz, 1H), 4.03 (dd, *J* = 4.7, 12.3 Hz, 1H), 5.44 (t, *J* = 4.6 Hz, 1H), 6.97 (s, 1H), 7.35 (d, *J* = 8.2 Hz, 2H), 7.48 (s, 1H), 7.78 (d, *J* = 8.2 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃+CD₃OD): δ 20.0, 55.0, 61.4, 80.9, 105.3, 117.4, 119.3, 127.6, 128.8, 131.9, 138.9, 145.1, 147.7, 156.6, 169.5; **Analysis:** C₁₇H₁₆O₇S requires C, 56.04; H, 4.43; found: C, 55.97; H, 4.37%.

(R)-5-(Benzyloxy)-3-(hydroxymethyl)-6-methoxyisobenzofuran-1(3H)-one (51t)

Yield: 94%, colorless solid; **mp:** 126-128 °C; $[\alpha]_{25}^D$ -78.22 (*c* 1.10, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 689, 825, 975, 1025, 1076, 1223, 1268, 1312, 1334, 1494, 1528, 1621, 1752, 2924, 3032, 3385; **¹H NMR** (200 MHz, CDCl₃): δ 2.31 (br s, 1H), 3.75-3.85 (m, 1H), 3.92 (s, 3H), 3.98-4.06 (m, 1H), 5.22 (s, 2H), 5.36-5.41 (m, 1H), 6.92 (s, 1H), 7.28 (s, 1H), 7.32-7.45 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 56.2, 64.1,

71.1, 81.0, 105.4, 106.6, 118.6, 127.3, 128.3, 128.7, 135.6, 140.9, 151.2, 154.0, 170.5;

Analysis: C₁₇H₁₆O₅ requires C, 67.99; H, 5.37; found: C, 67.91; H, 5.35%.

(R)-5-Fluoro-3-(hydroxymethyl)isobenzofuran-1(3H)-one (51u)

Yield: 93%, colorless gum; $[\alpha]_{25}^D$ -77.21 (*c* 1.2, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 689, 825, 975, 1025, 1076, 1223, 1268, 1312, 1334, 1494, 1528, 1621, 1752, 2924, 3032, 3385; **¹H NMR** (500 MHz, CDCl₃): δ 2.87 (br s, 1H), 3.93 (dd, *J* = 4.0, 12.5 Hz, 1H), 4.11 (dd, *J* = 4.0, 12.5 Hz, 1H), 5.51-5.53 (t, *J* = 4.0 Hz, 1H), 7.21-7.27 (m, 2H), 7.88-7.91 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 63.4, 80.9, 109.7 (d, *J* = 24.6 Hz), 117.7 (d, *J* = 24.6 Hz), 122.6, 128.2 (d, *J* = 9.4 Hz), 149.7 (d, *J* = 9.4 Hz), 167.3 (d, *J* = 398.5 Hz), 167.9; **Analysis:** C₉H₇FO₃ requires C, 59.35; H, 3.87; found: C, 73.44, H, 4.08%.

(R)-3 (Hydroxymethyl)-5,6-dioxomethylisobenzofuran-1(3H)-one (51v)

Yield: 94%, colorless solid; **mp:** 144-145 °C; $[\alpha]_{25}^D$ -78.11 (*c* 1.2, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 698, 852, 957, 1024, 1067, 1232, 1286, 1319, 1343, 1484, 1582, 1612, 1766, 2942, 3054, 3389; **¹H NMR** (200 MHz, CDCl₃): δ 2.40 (br s, 1H), 3.84 (dd, *J* = 4.0, 12.4 Hz, 1H), 4.06 (dd, *J* = 4.0, 12.4 Hz, 1H), 5.41 (m, 1H), 6.13 (d, *J* = 2.3 Hz, 2H), 6.87 (s, 1H), 7.20 (s, 1H); **¹³C NMR** (50 MHz, DMSO-*d*₆): δ 62.1, 81.3, 102.8, 103.3, 119.7, 144.5, 149.0, 153.2, 169.6; **Analysis:** C₁₀H₈O₅ requires C, 57.70; H, 3.87; found: C, 57.68; H, 3.85%.

(R)-3-((R)-1-Hydroxybutyl)isobenzofuran-1(3H)-one (51w)

Yield: 93%; colorless solid; **mp:** 103-109 °C; $[\alpha]_{25}^D$ -76.89 (*c* 1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 694, 728, 1080, 1212, 1287, 1467, 1618, 1752, 2873, 2959, 3433; **¹H NMR** (200 MHz, CDCl₃): δ 0.93 (t, *J* = 6.8 Hz, 3H), 1.44-1.72 (m, 4H), 1.97 (br s, 1H), 3.99 (br s, 1H), 5.40 (d, *J* = 3.6 Hz, 1H), 7.51-7.57 (m, 2H), 7.65-7.73 (m, 1H), 7.87-7.92 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.9, 18.8, 34.9, 71.9, 83.2, 122.4,

125.6, 126.6, 129.2, 134.0, 147.2, 170.5; **Analysis:** C₁₂H₁₄O₃ requires C, 69.88; H, 6.84; found: C, 69.82; H, 6.81%.

(R)-3-((R)-1-Hydroxy-2-tertiarybutyldimethylsilylethyl)-5,6-dimethoxyisobenzofuran-1(3H)-one (51x)

Yield: 94%, colorless solid; **mp:** 166-168 °C; $[\alpha]_{25}^D$ -79.24 (*c* 1.1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 775, 837, 1060, 1137, 1471, 1503, 1740, 2855, 2926, 3406; **¹H NMR** (200 MHz, CDCl₃): δ 0.08 (d, *J* = 5.4 Hz, 6H), 0.90 (s, 9H), 2.30 (d, *J* = 5.5 Hz, 1H), 3.62-3.82 (m, 2H), 3.94 (s, 3H), 3.98 (s, 3H), 4.05-4.12 (m, 1H), 5.51 (d, *J* = 3.4 Hz, 1H), 6.98 (s, 1H), 7.28 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ -5.0, -4.5, 17.8, 25.5, 56.2, 63.1, 73.3, 80.1, 104.2, 106.0, 118.9, 141.5, 150.6, 154.6, 170.6; **Analysis:** C₁₈H₂₈O₆Si requires C, 58.67; H, 7.66; found: C, 58.65; H, 7.56%.

(R)-3-(Hydroxy(phenyl)methyl)-5,6-dimethoxyisobenzofuran-1(3H)-one (51y)

Yield: 94%, colorless solid; **mp:** 113-115 °C; $[\alpha]_{25}^D$ -79.23 (*c* 1.15, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 857, 974, 1026, 1064, 1158, 1216, 1334, 1604, 1743, 2858, 2928, 3430; **¹H NMR** (200 MHz, CDCl₃): δ 3.05 (br s, 1H), 3.64 (s, 3H), 3.90 (s, 3H), 4.69 (d, *J* = 7.4 Hz, 1H), 5.47 (d, *J* = 7.4 Hz, 1H), 5.85 (s, 1H), 7.20 (s, 1H), 7.34-7.41 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃+CD₃OD): δ 54.7, 74.3, 83.0, 104.3, 117.4, 126.6, 127.4, 138.1, 140.6, 149.8, 153.5, 170.6; **Analysis:** C₁₇H₁₆O₅ requires C, 67.99; H, 5.37; found: C, 67.92; H, 5.29%.

(R)-3-((R)-1-Hydroxyheptyl)-5,6,7-trimethoxyisobenzofuran-1(3H)-one (51z)

Yield: 92%, colorless solid; **mp:** 113-115 °C; $[\alpha]_{25}^D$ -78.36 (*c* 1.08, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 796, 1089, 1130, 1254, 1326, 1465, 1543, 1749, 2898, 2974, 3988 cm⁻¹; **¹H NMR** (200 MHz, CDCl₃): δ 0.87-0.93 (m, 3H), 1.26-1.37 (m, 8H), 1.64-1.78 (m, 2H), 3.87 (s, 3H), 3.94-3.96 (m, 4H), 4.13 (s, 3H), 5.23 (d, *J* = 3.0 Hz, 1H), 6.68 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.0, 22.5, 25.7, 29.1, 31.7, 32.8, 56.3, 61.2,

62.1, 72.1, 81.7, 99.8, 111.1, 141.8, 145.3, 152.2, 159.6, 168.1; **Analysis:** C₁₈H₂₆O₆ requires C, 63.89; H, 7.74; found: C, 63.81; H, 7.68%.

(2S, 3R)-Ethyl 3-(3-cyanophenyl)-2,3-dihydroxypropanoate (56)

Yield: 93%; colorless gum; $[\alpha]_{25}^D$ -36.06 (*c* 1.2, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 680, 725, 954, 1057, 1118, 1214, 1291, 1734, 2229, 2985, 3443; **¹H NMR** (200 MHz, CDCl₃): δ 1.30 (d, *J* = 7.1 Hz, 3H), 3.26 (d, *J* = 7.5 Hz, 1H), 3.43 (d, *J* = 5.8 Hz, 1H), 4.24-4.34 (m, 3H), 5.02 (dd, *J* = 2.3, 7.5 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 1H), 7.57-7.67 (m, 2H), 7.72 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.1, 62.3, 73.5, 74.5, 112.2, 118.6, 129.0, 130.2, 130.9, 131.3, 141.9, 172.3; **Analysis:** C₁₂H₁₃NO₄ requires C, 61.27; H, 5.57; N, 5.95; found: C, 61.26; H, 5.54; N, 5.89.

(2S, 3R)-Ethyl 3-(4-cyanophenyl)-2,3-dihydroxypropanoate (57)

Yield: 93%; colorless solid; **mp:** 102-103 °C; $[\alpha]_{25}^D$ -36.42 (*c* 1.1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 685, 765, 1017, 1050, 1105, 1204, 1257, 1752, 2228, 2978, 3332; **¹H NMR** (200 MHz, CDCl₃): δ 1.31 (d, *J* = 7.0 Hz, 3H), 3.03 (d, *J* = 7.6 Hz, 1H), 3.27 (d, *J* = 5.3 Hz, 1H), 4.24-4.35 (m, 3H), 5.05 (dd, *J* = 2.3, 7.5 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 1H), 7.57-7.67 (m, 2H), 7.72 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃+CD₃OD): δ 12.9, 60.6, 73.2, 74.2, 110.0, 117.9, 126.7, 131.0, 146.2, 171.4; **Analysis:** C₁₂H₁₃NO₄ requires C, 61.27; H, 5.57; N, 5.95; found: C, 61.23; H, 5.52; N, 5.84%.

Benzyl(1R,2S)-2-(ethoxycarbonyl)-1-(2-cyanophenyl)-2-hydroxyethylcarbamate (58)

Sodium hydroxide (60 mg, 1.5 mmol) was dissolved in water (4 mL), and 0.5 mL of this NaOH solution was transferred to a small vial containing K₂OsO₂(OH)₄ (0.02 mmol for 4 mol %) for later use. To the remainder of the NaOH solution were added the carbamate (1.55 mmol) and *n*-PrOH (2 mL). The mixture was stirred for 2-3 min and placed in a water bath before *tert*-butylhypochlorite (175 μ L, 1.52 mmol) was slowly added with vigorous stirring. Then, the resulting solution was sequentially

treated with a solution of (DHQD)₂PHAL (0.025 mmol for 5 mol %) in *n*-PrOH (1 mL), the *o*-cyano ethylcinnamate (0.50 mmol), the previously prepared solution of K₂OsO₂(OH)₄, and *n*-PrOH (1 mL). The reaction mixture was monitored by TLC to establish completion, quenched by the addition of saturated aqueous sodium sulfite (4 mL) while being cooled in an ice-water bath, and stirred for an additional 30 min. The separated aqueous phase was extracted with EtOAc (3 X 5 mL), and the combined organic extracts were washed with water (3 mL) followed by brine (5 mL), dried over anhyd.Na₂SO₄, and concentrated under reduced pressure to give the crude products which were purified by column chromatography [silica gel (230-400 mesh) and pet. ether:EtOAc (60:40) as an eluent] to give product **58** in 64% yield with dr 6:1.

Yield: 64%; colorless gum; $[\alpha]_{25}^D$ -36.06 (*c* 1.1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 857, 974, 1037, 1095, 1184, 1202, 1275, 1291, 1319, 1347, 1368, 1393, 1477, 1573, 1607, 1640, 1716, 2219, 2984, 3023, 3415; **¹H NMR** (200 MHz, CDCl₃): δ 1.28 (t, *J* = 7.1 Hz, 3H), 3.34 (d, *J* = 7.5 Hz, 1H), 4.29 (q, *J* = 7.1 Hz, 2H), 4.50 (s, 1H), 5.06 (dd, *J* = 2.3, 7.5 Hz, 1H), 5.62 (d, *J* = 8.9 Hz, 1H), 5.85 (d, *J* = 8.9 Hz, 1H), 7.32-7.36 (m, 5H), 7.39-7.56 (m, 3H), 7.66-7.77 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.2, 55.3, 60.3, 62.8, 72.5, 111.1, 117.0, 122.0, 128.4, 132.8, 133.2, 142.9, 145.8, 155.3, 172.0; **Analysis:** C₁₂H₁₃NO₄ requires C, 61.27; H, 5.57; N, 5.95; found: C, 61.26; H, 5.54; N, 5.89.

Section II:

First Enantioselective Synthesis of (-)-Matteucen C and Facile Synthesis of antileptic Stroke Drug, 3-Butylphthalide

2.2.1 Introduction

3-Alkylated phthalide frameworks are present in a large number of natural products and biologically active compounds.¹ In recent years such chiral phthalide systems have attracted considerable attention as they play a pivotal role in modern drug discovery. Among them (-)-matteucen C (**62**) and 3-butylphthalide (**1**) are of particular interest due to their biological activities. *Matteuccia orientalis* (HOOK.) TREV (Onocleaceae), mainly distributed in Southern China, is a Chinese medicinal herb used for the treatment of hemostatics and relieving ostealgia.³³ Search for new bioactive constituents from the rhizomes of this plant led to the isolation of new isocoumarin derivatives, such as matteucen A (**60**), racemic-matteucen B (**61**) and new phthalide derivative, (-)-matteucen C (**62**) (Fig. 12).³⁴

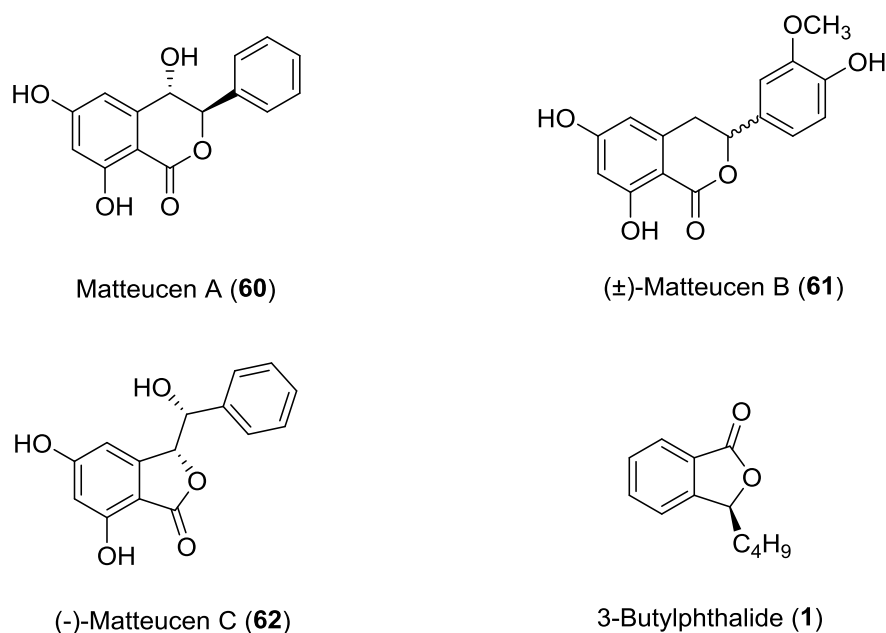


Fig. 12: Structures of matteucen A-C and 3-butylphthalide

Worldwide ischemic stroke is the second leading cause of human morbidity and mortality.³⁵ Currently, ischemic stroke approximately causes loss of 5 million people each year, and the mortality rate of stroke is increasing.³⁶ Studies have suggested that the pathogenesis of ischemic stroke is attributed to the interaction of multiple factors, including genetic high risk, thrombosis, and chronic inflammatory diseases such as hypertension, diabetes, and etc.³⁷ During the process of ischemic stroke, the platelet aggregation related thrombosis limits sufficient blood flow in the special region of the brain and leads to ischemic inflammation and brain damage.³⁸ Although few drugs are available for the intervention of ischemic stroke, the efficacy of these drugs are not satisfactory and need to be used in combinations with other active drugs. For centuries in China, seeds of *Apium graveolens* Linn, Chinese celery have been employed against ischemic stroke and 3-butylphthalide (**1**) has been reported as the active ingredient in the seed oil of *Apium graveolens* Linn celery.^{2a} 3-Butylphthalide (**1**) (**Fig. 12**), was approved as antiischemic drug by the State Food and Drug Administration (SFDA) of China in 2002. It intervenes ischemic stroke through multiple mechanism for example, by improving energy metabolism, reducing oxidative damage,³⁹ improving micro circulation in arterioles,⁴⁰ decreasing neuronal apoptosis,⁴¹ improving mitochondrial function and inhibiting inflammation.⁴² Moreover, it showed promising preclinical potential as a multi-target drug for the prevention and treatment of Alzheimer's disease and vascular dementia.⁴³ It is noteworthy that Alzheimer's disease is the most common form of senile dementia, characterized by progressive memory loss and vascular dementia, the second most common cause of dementia especially in the Asian population.⁴⁴ Currently, there is no specific drug available to prevent or cure vascular dementia.

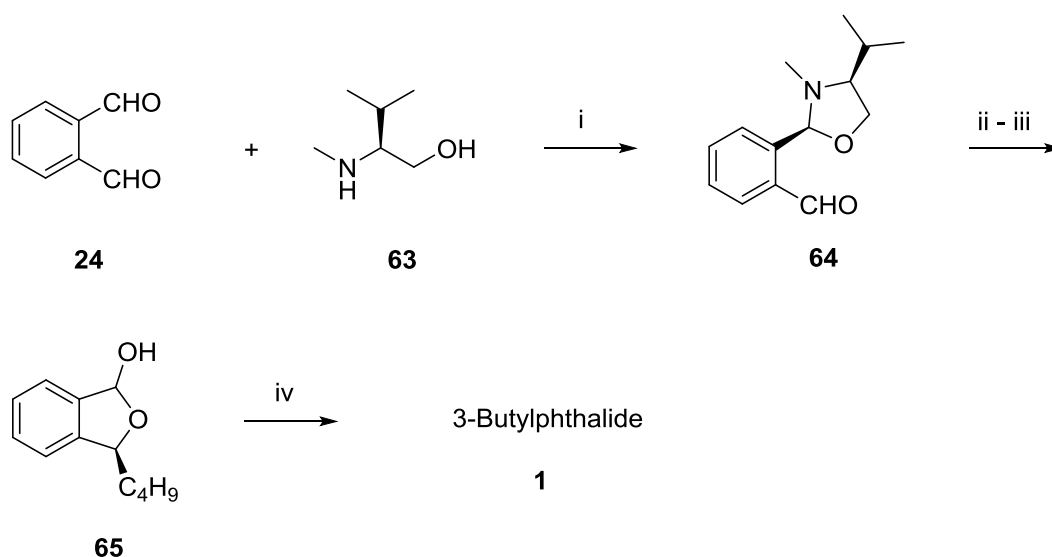
2.2.2 Review of Literature

Literature search revealed that there is no report available for the synthesis of (-)-matteucen C (**62**), where as there are six reports for the synthesis of 3-butylphthalide.

⁴⁵⁻⁵⁰ A short description of all the six methods are presented below.

Takahashi's approach (1991)⁴⁵

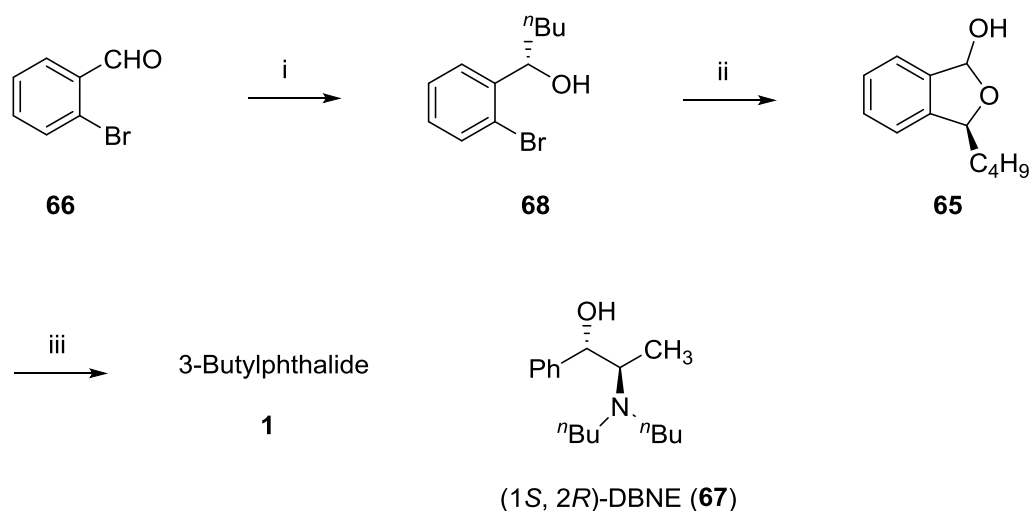
Takahashi *et al.* have utilized valine-based chiral auxiliary for the introduction of chirality in 3-butylphthalide (**1**) synthesis. Thus, chiral oxazolidine **64** was prepared by condensation of phthalaldehyde **24** with (*S*)-*N*-methyl valinol **63** in 56% yield (dr = 93:7). Diastereoselective addition of dibutylcupriolithium onto oxazolidine **64** followed by subsequent hydrolysis under acidic condition afforded lactol **65** in 85% ee. Further, oxidation of lactol **65** with PCC gave 3-butylphthalide (**1**) (**Scheme 19**).



Scheme 19: (i) anhyd. Na_2SO_4 , CH_2Cl_2 , 25 °C, 16 h, 56%; (ii) $(^i\text{Bu})_2\text{CuLi}$, Et_2Zn , $^i\text{BuMgCl}$, THF, -50 °C, 24 h; (iii) *p*-TSA, THF:H₂O (5:1), reflux, 1 h, 86% (over 2 steps); (iv) PCC, CH_2Cl_2 , 25 °C, 30 min, 41%.

Soai's approach (1991)⁴⁶

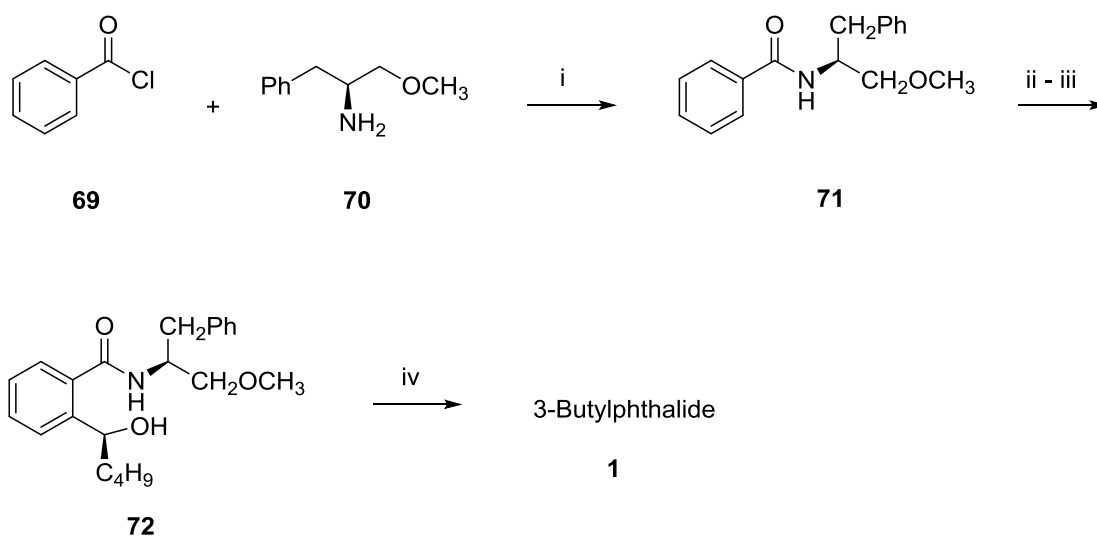
Soai *et al.* have described the synthesis of 3-butylphthalide (**1**) using enantioselective addition of dibutylzinc reagent as key step. Thus, *o*-bromobenzaldehyde **66** was subjected to asymmetric addition of dibutylzinc reagent catalyzed by chiral N,N-dibutylnorephedrine (**67**) to give alcohol **68** in 94% yield and 90% ee. Treatment of alcohol **68** with *n*-BuLi followed by quenching with DMF afforded lactol **65** in 82% yield, which was finally transformed to 3-butylphthalide (**1**) using Ag₂O oxidation (Scheme 20).



Scheme 20: (i) (1*S*, 2*R*)-DBNE (**67**), *n*-Bu₂Zn, hexane, 25 °C, 17 h, 94%; (ii) *n*-BuLi, DMF, 82%; (iii) Ag₂O, 25 °C, 76%.

Matsui's approach (1993)⁴⁷

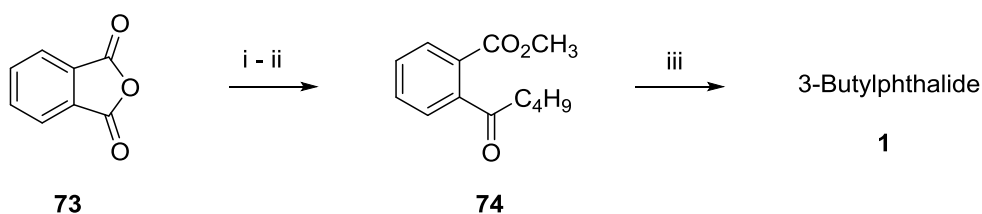
Matsui *et al.* have commenced their synthesis from commercially available benzoyl chloride **69**. The chiral benzamide **71** was obtained in 90% yield by treating benzoyl chloride **69** with chiral amine **70**. *ortho*-Lithiation of chiral benzamide **71** followed by diastereoselective addition onto *n*-pentanal afforded alcohol **72** in 57% yield (dr = 90:10). Finally, cyclization under acidic condition provided 3-butylphthalide (**1**) (Scheme 21).



Scheme 21: (i) Et₃N, THF, 25 °C, 16 h, 90%; (ii) *n*-BuLi, TMEDA, THF, -78 °C to 0 °C, (iii) C₄H₉CHO, THF, -78 °C, 4 h, 57% (over two steps); (iv) 5% HCl, dioxane, reflux, 4 h, 95%.

Kitayama's approach (2002)⁴⁸

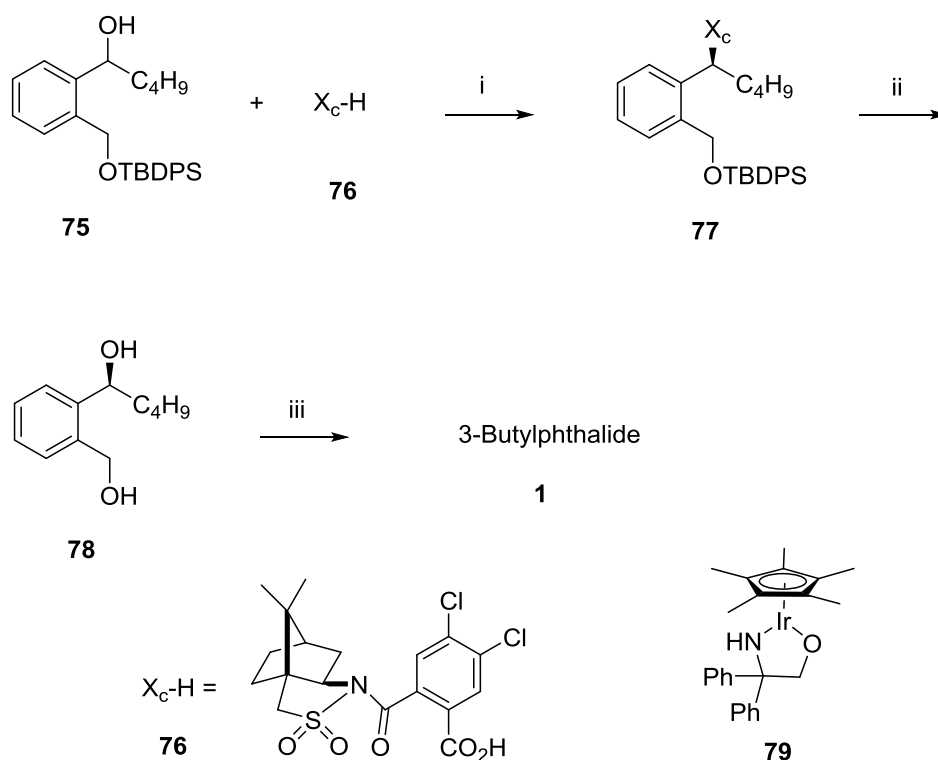
Kitayama have reported the synthesis of 3-butylphthalide (1) by employing enzymatic reduction as the key reaction. Methyl 2-pentanoyl benzoate (74) was prepared from phthalic anhydride (73) by reacting with dibutylcadmium, followed by esterification in acidic methanol. Methyl 2-pentanoylbenzoate (74) was subjected to enzymatic reduction with *Pseudomonas putida* ATCC to give 3-butylphthalide (1) in 80% yield and 99% ee (Scheme 22).



Scheme 22: (i) CdCl₂, *n*-BuBr, Mg, THF, reflux, 30 min, 64%; (ii) 97% H₂SO₄, CH₃OH, reflux, 30 min, 64%; (iii) *Pseudomonas putida* ATCC 12633, H₂O, 0 °C, 24 h, 80%.

Kosaka's approach (2002)⁴⁹

Kosaka *et al.* have utilized camphorsultam dichlorophthalic acid (**76**) for enantio-resolution of racemic alcohol as key step in 3-butylphthalide (**1**) synthesis. Thus, racemic alcohol **75** was esterified with chiral auxiliary **76** to give diastereomeric mixture of esters, which were separated by HPLC to provide optically pure ester **77** in 49% yield. Enantiopure diol **78** was obtained by treating ester **77** with methanolic KOH. Selective oxidation of primary alcohol in **78** followed by lactol formation in a single step was achieved with Ir catalyst **79** to afford 3-butylphthalide (**1**) in 99% ee (**Scheme 23**).

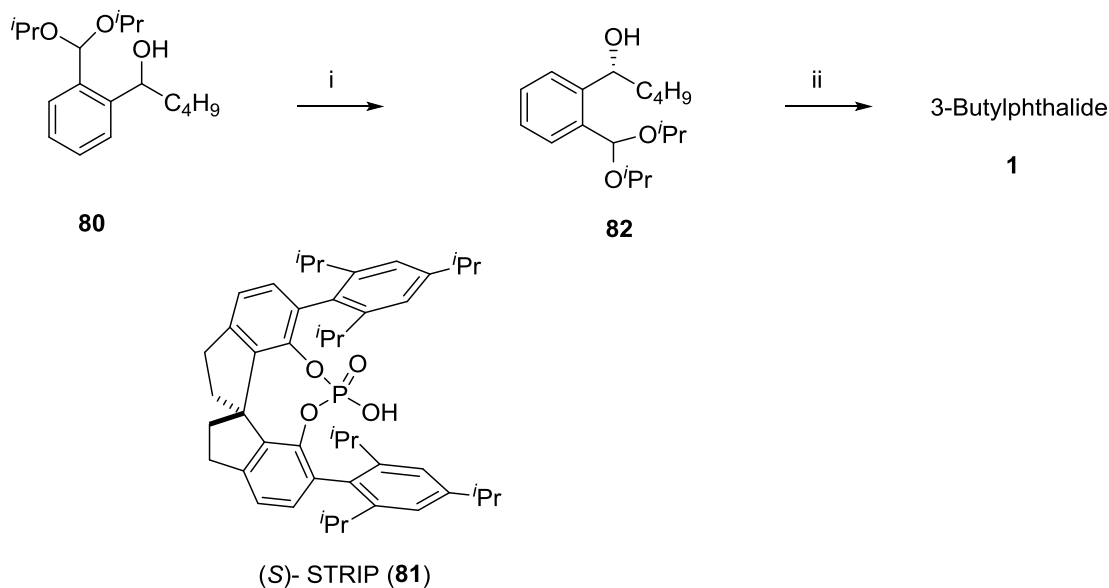


Scheme 23: (i) DCC, DMAP, CH₂Cl₂, 25 °C, 18 h, HPLC separation, 49%; (ii) KOH, CH₃OH, 16 h, 99%; (iii) Ir catalyst **79**, acetone, 25 °C to 50 °C, 48 h, 84%.

List's approach (2010)⁵⁰

List *et al.* have achieved the synthesis of 3-butylphthalide (**1**) by employing kinetic resolution as the key reaction. These authors have developed a new spirocyclic

phosphoric acid catalyst [(*S*)-STRIP (**81**)], for kinetic resolution of homoaldol **80** to afford chiral alcohol **82** in 34% yield and 95% ee. Jones oxidation of chiral alcohol **82** provided access to 3-butylphthalide (**1**) in 85% yield (**Scheme 24**).



Scheme 24: (i) (*S*)-STRIP (**81**), CH₂Cl₂, 4 Å MS, 25 °C, 6 h, 34%; (ii) CrO₃, conc. H₂SO₄, acetone:water (3:1), 0 °C, 1 h, 85%.

2.2.3 Present Work

2.2.3.1 Objective

Even though few methods have been reported for the synthesis of 3-butylphthalide (**1**),⁴⁵⁻⁵⁰ they suffer from certain limitations such as use of chiral auxiliary, expensive and exotic reagents, low overall yields, etc. In this context, a more practical method for the synthesis of 3-butylphthalide (**1**) is highly desirable. In Section I of this Chapter, we have described an elegant method for the synthesis of 3-substituted chiral phthalide derivatives **51a-z**. In continuation of the work on Os-catalyzed oxidative cyclization of *o*-cyano alkenes, a short synthesis of two natural products namely (-)-matteucen C (**62**) and 3-butylphthalide (**1**) is described in this section.⁵¹

Retrosynthetic analysis of (-)-matteucen C (**62**) reveals that *o*-cyanostilbene derivative (**85**) could serve as the key intermediate for the oxidative cyclization leading to the synthesis of (-)-matteucen C (**62**). *o*-Cyanostilbene derivative (**85**) can in turn be obtained from *o*-bromobenzaldehyde (**83**) (Fig. 13).

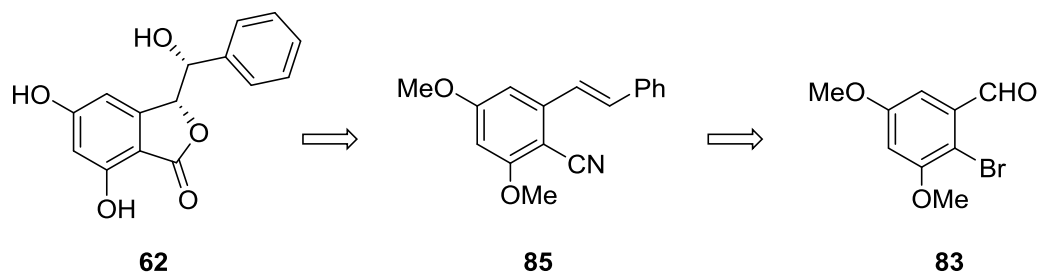


Fig. 13: Retrosynthetic analysis of (-)-matteucen C (**62**)

Similarly, we envisaged that 3-butylphthalide (**1**) could be obtained by means of Barton-McCombie deoxygenation of phthalide **51w**. The required phthalide **51w** could be prepared by means of oxidative cyclization of *o*-cyanostyrene **50w**. The *o*-cyanostyrene **50w** could in turn be obtained from *o*-bromobenzaldehyde (**66**) (Fig. 14).

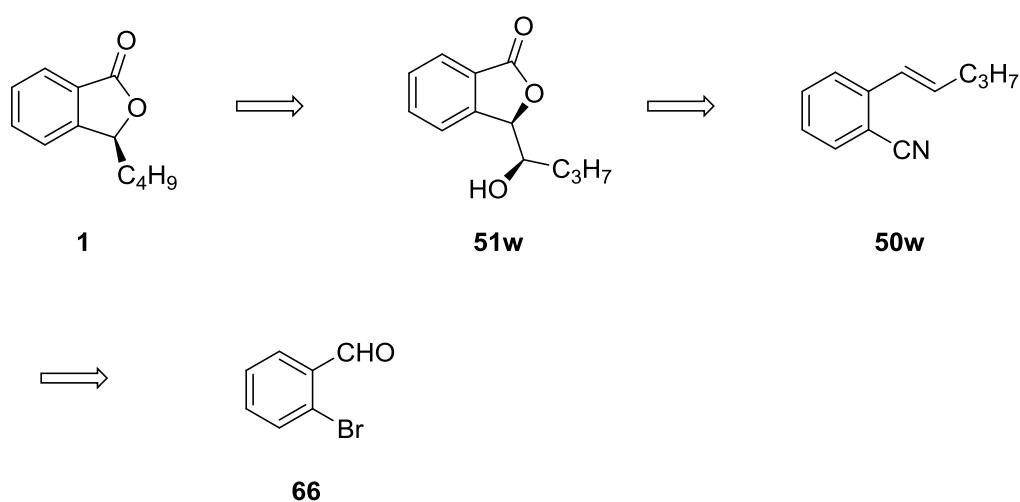
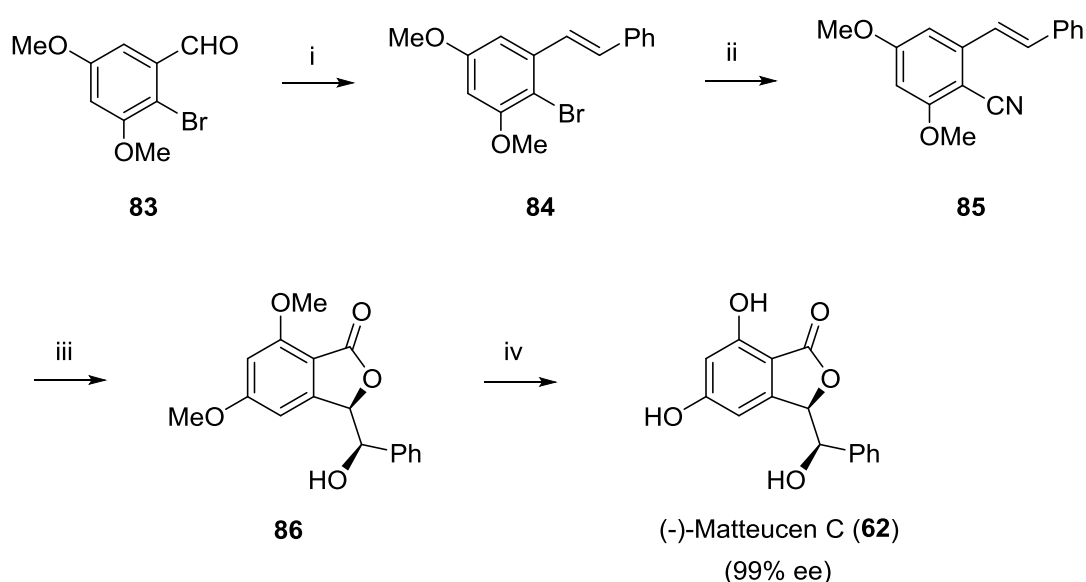


Fig. 14: Retrosynthetic analysis of 3-butylphthalide (**1**)

2.2.4 Results and Discussion

(a) First Enantioselective Synthesis of (-)-Matteucen C (62)

The complete synthetic sequence for (-)-matteucen C (**62**), wherein Os-catalyzed CN-assisted oxidative cyclization of *o*-cyanostilbene derivative (**85**) constitutes a key step for the introduction of chirality, is presented in **Scheme 25**.



Scheme 25: (i) $\text{PhCH}_2\text{Ph}_3\text{P}^+\text{I}^-$, *n*-BuLi, THF, 0 to 25 °C, 3 h, 82%; (ii) CuCN (3.5 equiv), DMF, reflux, 14 h, 83%; (iii) AD-mix- β , *tert*-BuOH:THF:H₂O (0.5:0.5:1), 25 °C, 7 h, 93%; (iv) BBr₃, CH₂Cl₂, 25 °C, 12 h, 69%.

Accordingly, the synthesis of (-)-matteucen C (**62**) was undertaken starting from *o*-bromobenzaldehyde derivative **83**, which on subjecting to Wittig olefination [$\text{PhCH}_2\text{Ph}_3\text{P}^+\text{I}^-$, *n*-BuLi, THF] gave (*E*)-*o*-bromostilbene derivative **84** in 82% yield. Two doublets at δ 6.99 (d, $J = 16.1$ Hz, 1H) and 7.52 (d, $J = 16.1$ Hz, 1H) integrating for one proton each in the ¹H NMR spectrum of **84** accounted for olefinic protons. It was further supported by the typical olefinic carbon signals at δ 127.9 and 128.6 in its ¹³C NMR spectrum (**Fig. 15**).

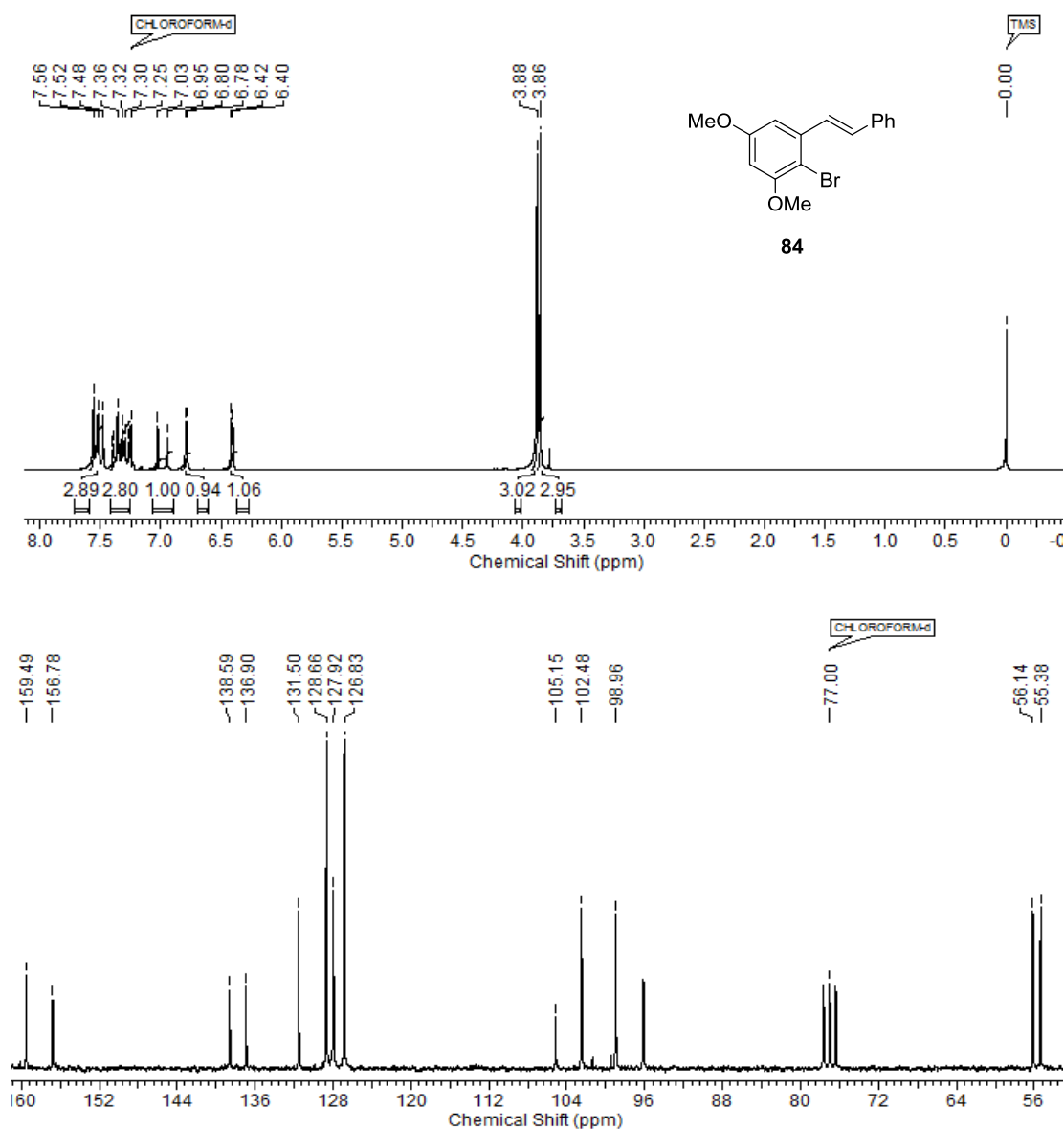


Fig. 15: ¹H and ¹³C NMR spectra of *o*-bromostilbene derivative **84**

o-Bromostilbene derivative **84** was then converted to *o*-cyanostilbene derivative **85** using Rosenmund-von Braun reaction [CuCN, DMF, reflux, 83%]. The formation of the *o*-cyanostilbene derivative **85** was confirmed by the appearance of CN carbon at δ 115.7 in its ¹³C NMR spectrum. Its IR spectrum displayed a sharp CN stretching vibrational frequency at 2216 cm⁻¹ (**Fig. 16**).

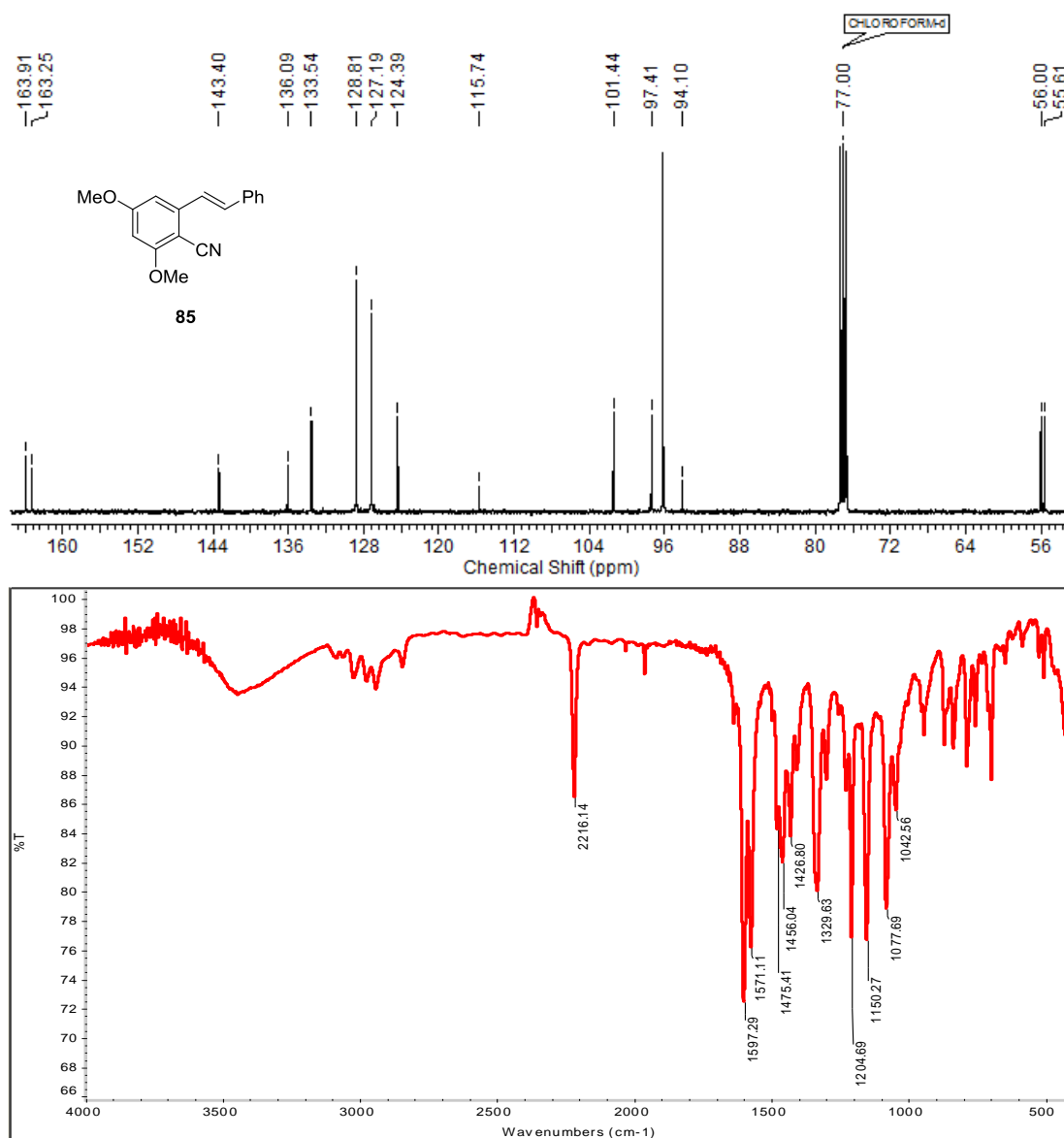
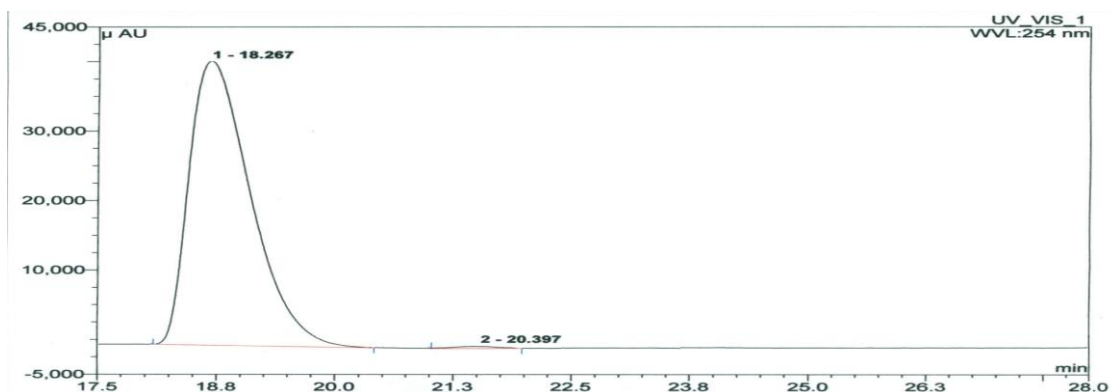
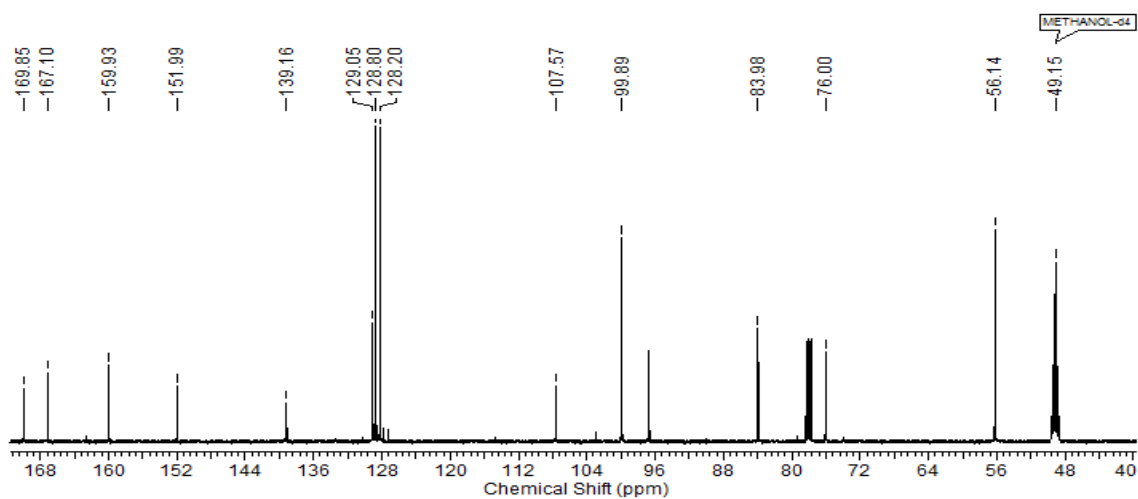
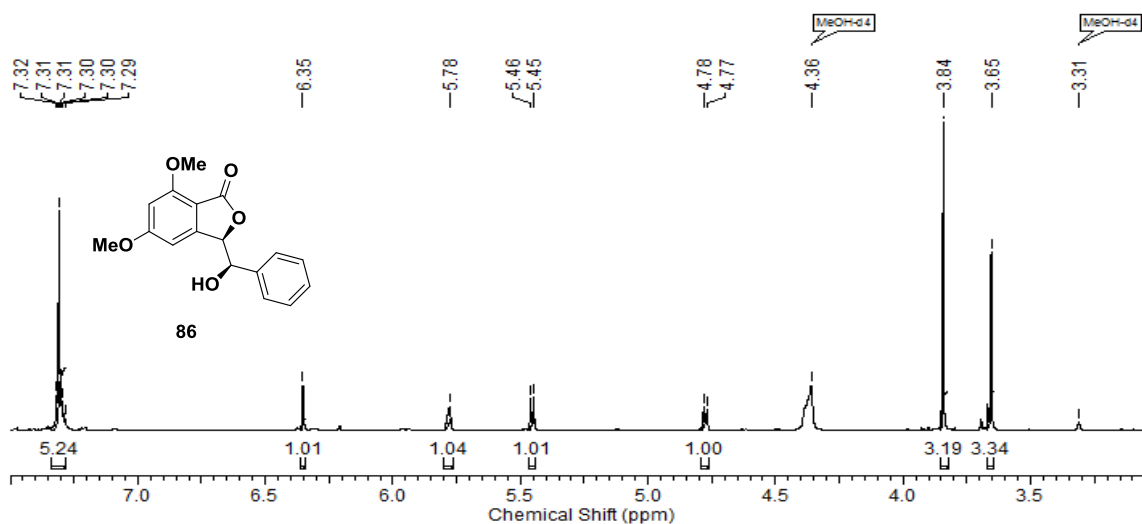


Fig. 16: ^{13}C NMR and IR spectra of *o*-cyanostilbene derivative **85**

o-Cyanostilbene derivative **85** was then subjected to CN-assisted one-pot oxidative cyclization using AD-mix- β process to give chiral phthalide **86** in 93% yield and 99% ee. The ^1H , ^{13}C NMR and IR spectra of **86** confirmed the formation of phthalide (Fig. 17). Thus, the methine protons attached to lactone moiety and hydroxyl group resonated at δ 4.77 (d, $J = 6.4$ Hz, 1H) and 5.45 (d, $J = 6.4$ Hz, 1H) respectively in its ^1H NMR spectrum. It was further substantiated by the appearance of carbonyl carbon [-(C=O)-O-] signal at δ 169.8 in its ^{13}C NMR spectrum. Its IR spectrum also exhibited a characteristic lactone carbonyl absorption band at 1752 cm^{-1} .



No	Ret. Time min	Height μ AU	Area μ AU* min	Rel. Area %	Amount	Type
1	18.27	40895.238	30456.124	99.36	n. a.	BMB
2	20.40	2196.930	195.138	0.64	n. a.	BMB

Fig. 17: ^1H , ^{13}C NMR spectra and HPLC chromatogram of phthalide **86**

The enantiomeric excess of chiral phthalide **86** was determined to be 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 18.27 min (99.36%) and 20.40 min (0.64%) (**Fig. 17**).

Finally, demethylation of chiral phthalide **66** was achieved with BBr₃ in CH₂Cl₂ that afforded (-)-matteucen C (**62**) in 69% yield with 99% ee. The optical rotation of the target molecule was found to be $[\alpha]_{25}^D$ -54.16 (*c* 1.0, CH₃OH). The formation of (-)-matteucen C (**62**) was confirmed by the appearance of two doublets at δ 4.94 (t, *J* = 4.8 Hz, 1H) and 5.45 (d, *J* = 4.0 Hz, 1H) in its ¹H NMR spectrum corresponding to the methine protons attached to benzylic hydroxyl group and lactone moiety respectively (**Fig. 18**). The corresponding methine carbons resonated at δ 72.6 and 81.7 in its ¹³C NMR spectrum. The spectral data of (-)-matteucen C (**62**) were in complete agreement with the reported values.³⁴

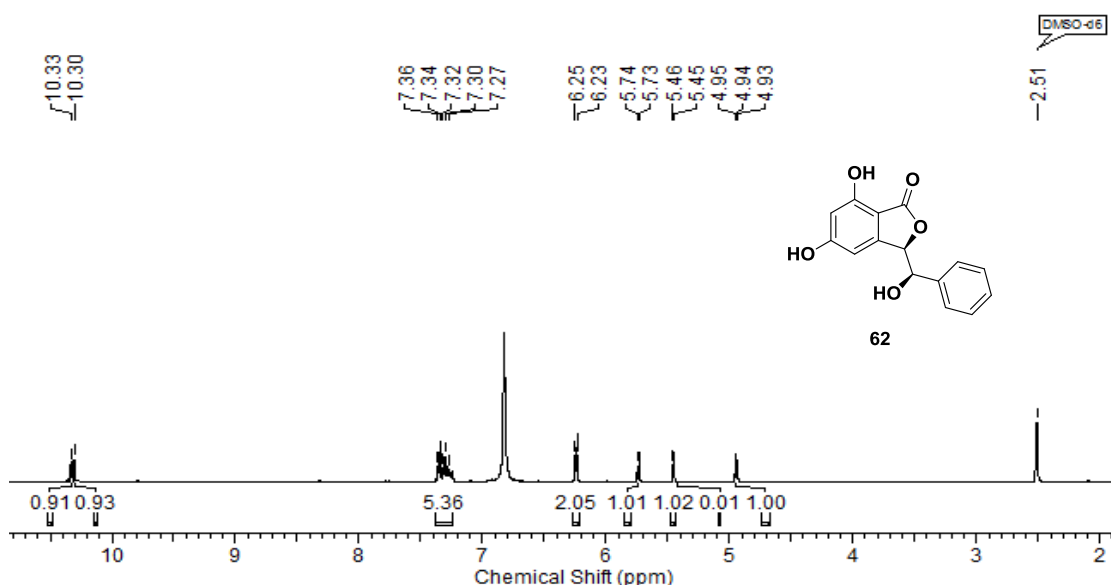
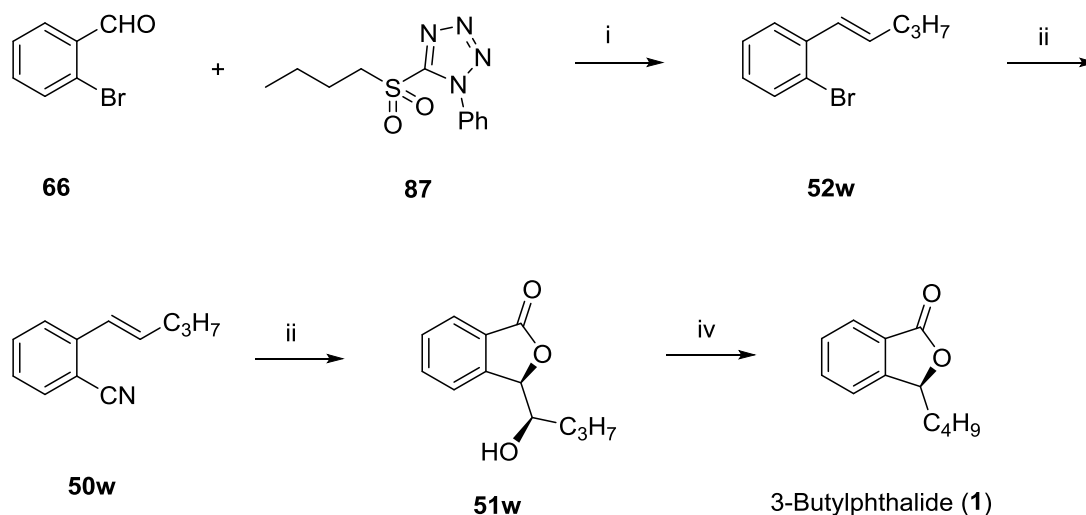


Fig. 18: ¹H NMR spectrum of (-)-matteucen C (**62**)

(b) Facile Enantioselective Synthesis of 3-Butylphthalide (1)

The synthetic scheme for 3-butylphthalide (**1**), wherein Os-catalyzed CN-assisted oxidative cyclization of *o*-cyanostyrene **50w** constitutes a key step for the introduction of chirality, is shown in **Scheme 26**.



Scheme 26: (i) NaHMDS, THF, -78 °C to 25 °C, 14 h, 82%; (ii) CuCN (3.5 equiv), dry DMF, reflux, 14 h, 87%; (iii) AD-mix- β , *tert*-BuOH:THF:H₂O (0.5:0.5:1), 25 °C, 7 h, 93%; (iv) (a) 1,1-thiocarbonyldiimidazole, DMAP (10 mol%), CH₂Cl₂, 18 h; (b) AIBN (10 mol%), Bu₃SnH, toluene, reflux, 15 min, 86%.

The present synthesis of 3-butylphthalide (**1**) commenced from commercially available *o*-bromobenzaldehyde (**66**), which on subjecting to Julia-Kocienski olefination⁵² with butyl sulfone **87** gave (*E*)-2-bromoalkene **52w** in 82% yield. The formation of (*E*)-2-bromoalkene **52w** was confirmed from its ¹H NMR spectrum, which showed a doublet at δ 6.69 (d, J = 15.6 Hz, 1H) and a triplet of doublet at δ 6.15 (td, J = 6.9, 15.6 Hz, 1H) corresponding to *trans*-olefinic protons of alkene **52w**; It was further confirmed by its ¹³C NMR spectrum, which displayed typical carbon signals at δ 127.4 and 128.8 corresponding to olefinic carbons (**Fig. 19**).

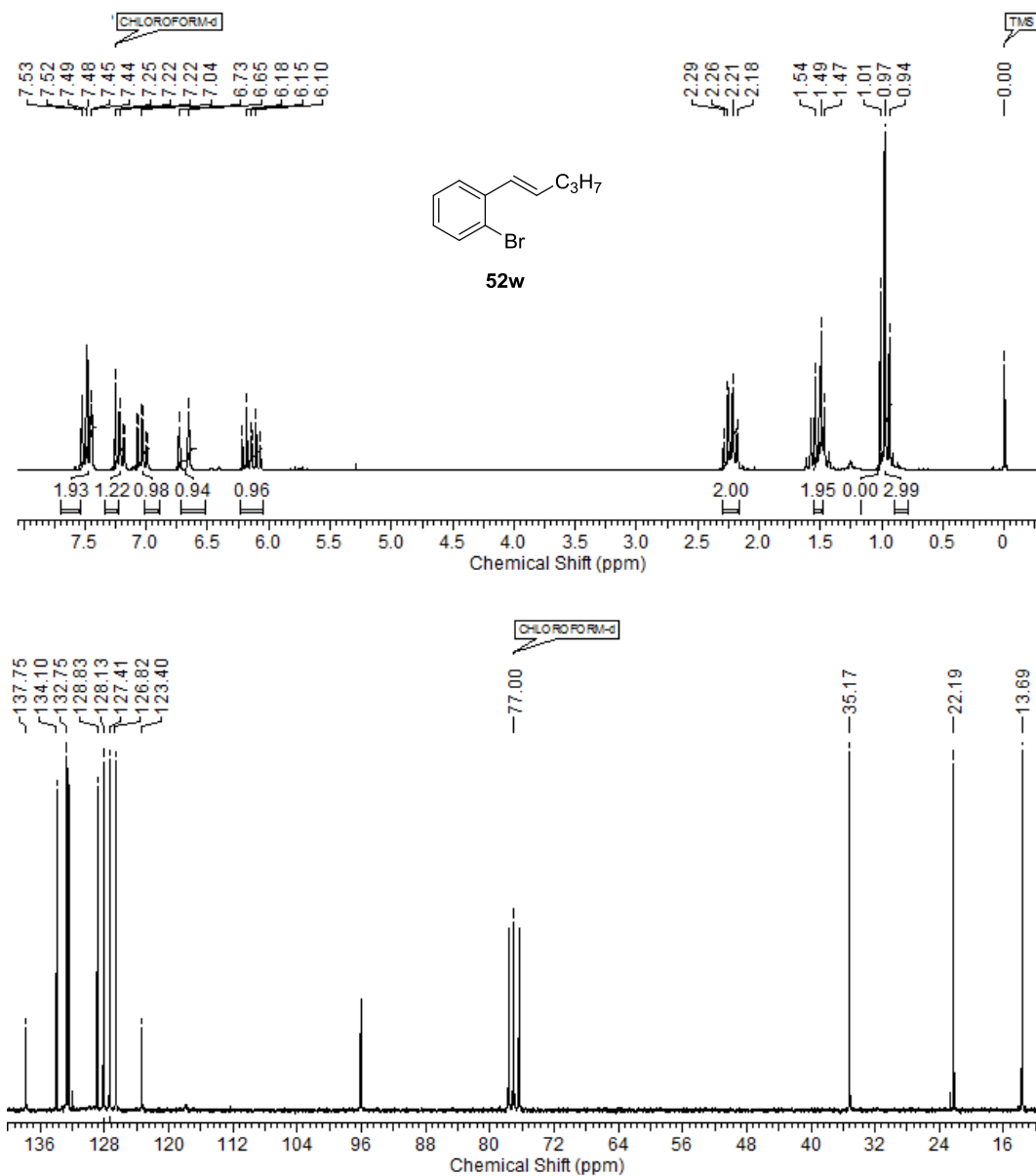


Fig. 19: ¹H and ¹³C NMR spectra of (*E*)-2-bromoalkene **52w**

(*E*)-2-Bromoalkene **52w** was then converted to *o*-cyanoalkene derivative **50w** using Rosenmund-von Braun reaction [CuCN, DMF, reflux, 87%]. Its ¹H NMR spectrum showed characteristic doublet at δ 6.74 (d, $J = 15.3$ Hz, 1H) and a triplet of doublet at δ 6.43 (dt, $J = 6.8, 15.3$ Hz, 1H), corresponding to *trans*-olefinic protons of alkene **50w**. Its ¹³C NMR spectrum showed a characteristic carbon signal for CN carbon at δ 117.9 confirming the formation of *o*-cyanoalkene **50w** (Fig. 20). Its IR spectrum displayed a sharp CN stretching vibrational frequency at 2219 cm^{-1} .

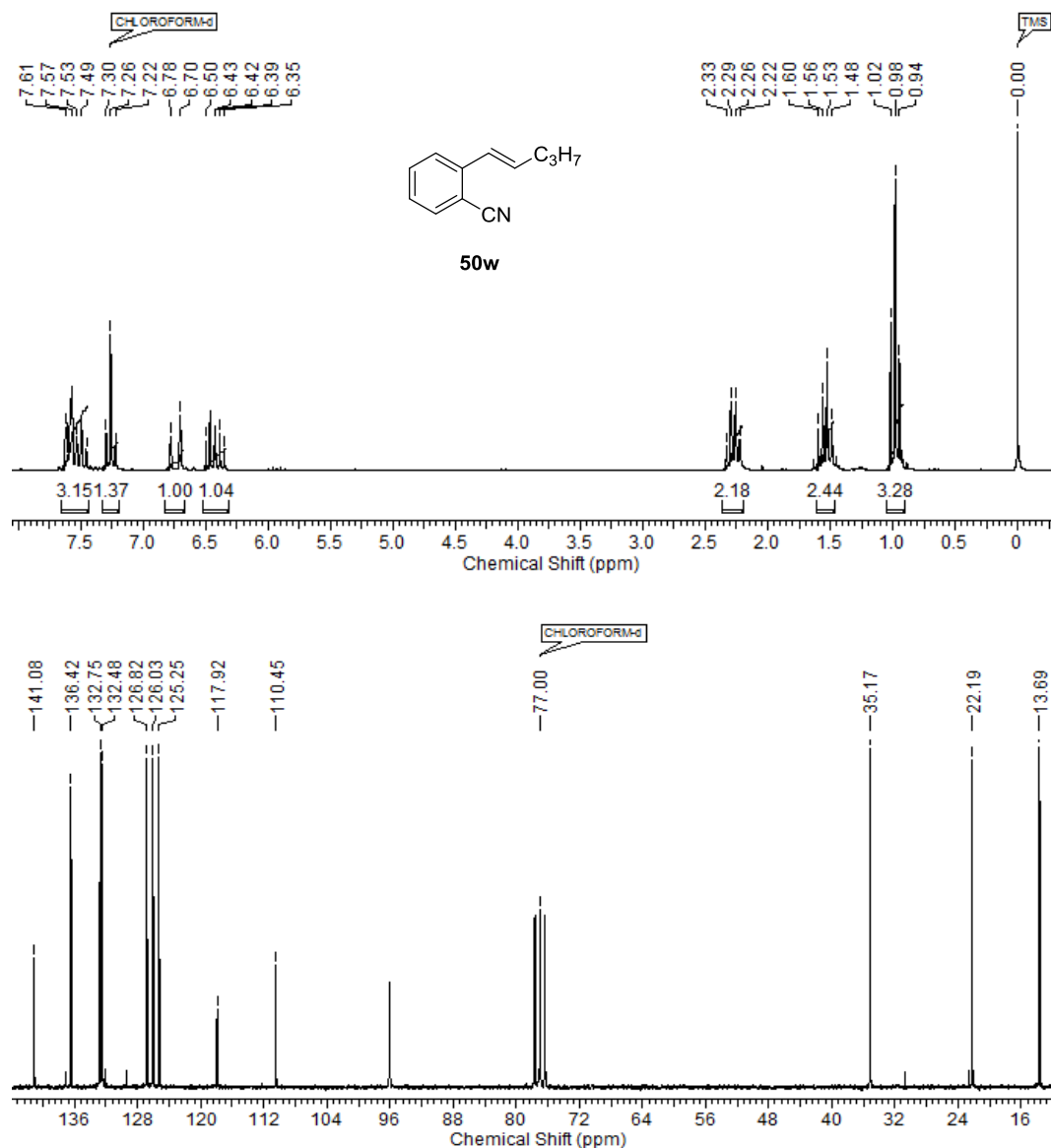


Fig. 20: ¹H and ¹³C NMR spectra of *o*-cyanoalkene **50w**

o-Cyanoalkene **50w** was then subjected to CN-assisted one-pot oxidative cyclization using AD-mix- β process to give chiral phthalide **51w** in 93% yield and 97% ee. The ¹H, ¹³C NMR and IR spectra of **51w** confirmed the formation of phthalide **51w** (Fig. 21). The methine proton attached to lactone moiety resonated at δ 5.40 (d, $J = 3.6$ Hz, 1H) in its ¹H NMR spectrum. It was further substantiated by the appearance of carbonyl carbon [-(C=O)-O-] signal at δ 170.5 in its ¹³C NMR spectrum. Its IR spectrum exhibited a characteristic lactone carbonyl absorption band at 1752 cm^{-1} .

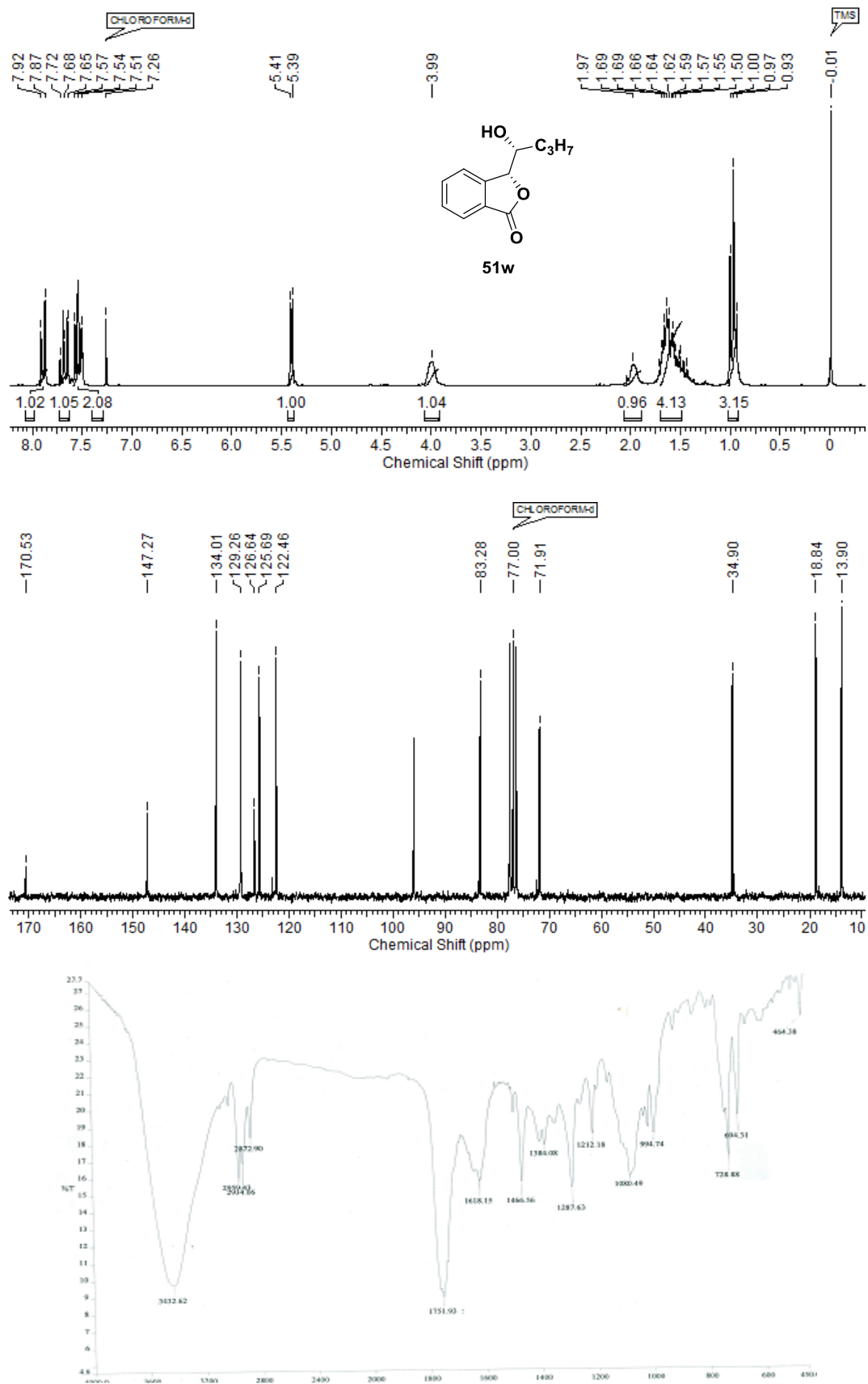


Fig. 21: ¹H, ¹³C NMR and IR spectra of phthalide **51w**

The optical purity of chiral phthalide **51w** was determined to be 97% ee from ^1H NMR analysis of the corresponding Mosher's ester **88** (Fig. 22) (see experimental section for details).

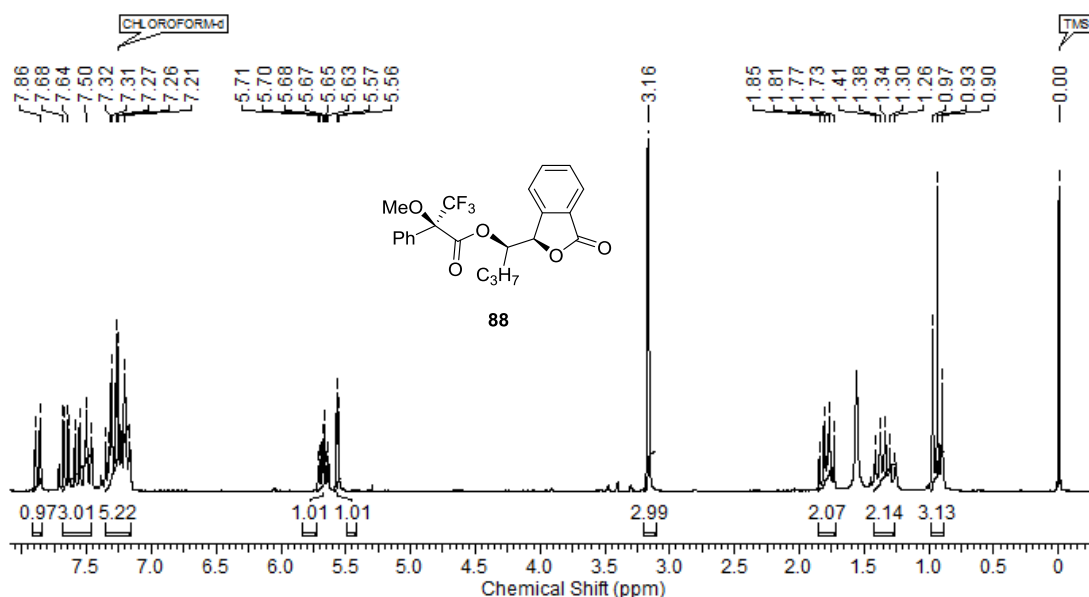


Fig. 22: ^1H NMR spectrum of Mosher's ester **88**

Finally, Barton-McCombie protocol⁵³ was utilized for deoxygenation of alcohol **51w**, in a two-step reaction sequence: (i) xanthate formation of alcohol **51w** (1,1-thiocarbonyldiimidazole, DMAP); (ii) followed by reduction of formed xanthate (Bu_3SnH , AIBN) afforded 3-butylphthalide (**1**) in 86% yield. The ^1H NMR spectrum of 3-butylphthalide (**1**) showed typical proton signals at δ 5.46 (dd, $J = 4.3, 7.9$ Hz, 1H), 1.68-1.86 (m, 1H) and 1.97-2.07 (m, 1H) corresponding to the methine proton attached to lactone moiety and homobenzylic protons respectively. Its ^{13}C NMR spectrum showed a characteristic carbonyl resonance of lactone [$-(\text{C}=\text{O})-\text{O}-$] at δ 170.3 (Fig. 23). The ee of 3-butylphthalide (**1**) was found to be 97% based on comparison of its optical rotation with the reported value $\{[\alpha]_{25}^{\text{D}} -67.0$ (c 1.15,

CHCl_3); lit.⁴⁹ $[\alpha]_{25}^D -64.7$ (c 1.06, CHCl_3). The spectral data of 3-butylphthalide (**1**) were in complete agreement with the reported values.⁴⁶⁻⁵¹

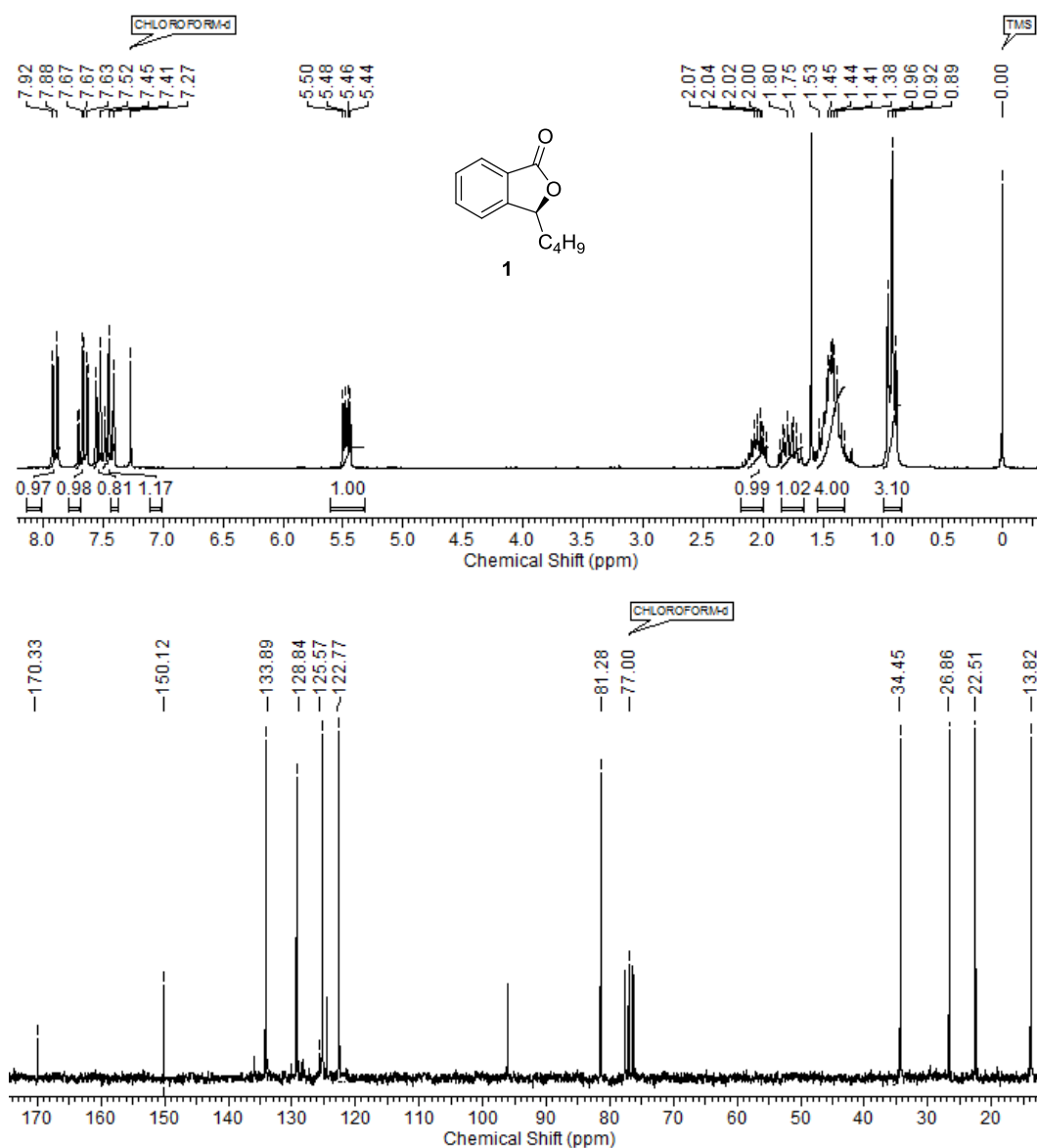


Fig. 23: ^1H and ^{13}C NMR spectra of 3-butylphthalide (**1**)

2.2.5 Conclusion

A short and an efficient enantioselective synthesis of (-)-matteucen C (**62**) and 3-butylphthalide (**1**) has been achieved in four linear steps (44 % overall yield, 99% ee and 57% overall yield, 97% ee respectively). Synthesis of (-)-matteucen C (**62**) was achieved for the first time, thereby confirming its structural and stereochemical

assignments. The CN-assisted one-pot oxidative cyclization of *o*-cyanoalkene derivatives **85** and **50w** was used as the key reaction, which proceeded to give high enantioselectivity.

2.2.6 Experimental Section

(*E*)-2-Bromo-1, 5-dimethoxy-3-styrylbenzene (**84**)

To a stirred solution of benzyltriphenylphosphonium iodide (2.1 g, 4.5 mmol) in THF was added *n*-butyllithium in hexane (2.8 mL, 4.5 mmol). The solution was stirred for 30 min at 0 °C followed by the addition of 2-bromobenzaldehyde (**83**) (1 g, 4.1 mmol) in THF dropwise *via* syringe at the same temperature and the reaction mixture was allowed to stir for 90 min at 25 °C (monitored by TLC). It was then cooled to 0 °C, diluted with sat. NH₄Cl (25 mL) and EtOAc (25 mL). The organic layer was separated and the aqueous layer extracted with EtOAc (2 x 20 mL). The combined organic extracts were washed with brine and dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to give crude product, which was purified by column chromatography [silica gel (230-400 mesh) and pet. ether:EtOAc (90:10) as an eluent] affording (*E*)-2-bromostyrene derivative **84** (1.12 g) as a gum.

Yield: 86%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 669, 769, 1216, 1384, 1468, 1580, 2098, 3020; **¹H NMR** (200 MHz, CDCl₃): δ 3.86 (s, 3H), 3.89 (s, 3H), 6.42 (d, *J* = 2.7 Hz, 1H), 6.99 (d, *J* = 16.1 Hz, 1H), 7.26-7.40 (m, 3H), 7.52 (d, *J* = 16.1 Hz, 1H), 7.53-7.55 (m, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 55.3, 56.1, 98.9, 102.4, 105.1, 126.8, 127.9, 128.0, 128.6, 131.5, 136.9, 138.5, 156.7, 159.4; **Analysis:** C₁₆H₁₅BrO₂ requires C, 60.21; H, 4.74; found: C, 60.08; H, 4.59%.

2, 4-Dimethoxy-6-styrylbenzonitrile (**85**)

o-Bromostilbene derivative **84** (1 g, 3.1 mmol) was taken up in dry DMF (10 mL) and CuCN (0.83 g, 9.3 mmol) was added and the mixture refluxed under N₂ for 18 h (monitored by TLC). The reaction mixture was then cooled to 25 °C and then diluted with water (30 mL) and EtOAc (25 mL). The organic layer was separated and the aqueous layer extracted with EtOAc (3 x 20 mL). The combined organic extracts were washed with brine and dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to give crude product, which was purified by column chromatography [silica gel (230-400 mesh) and pet. ether:EtOAc (7:3) as an eluent] to give *o*-cyanostilbene derivative **85** (0.7 g).

Yield: 83%; colorless solid; **mp:** 147-148 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 694, 831, 953, 1045, 1073, 1150, 1203, 1326, 1460, 1570, 1595, 2216; **¹H NMR** (400 MHz, CDCl₃): δ 3.90 (s, 6H), 6.34 (d, *J* = 2.3 Hz, 1H), 6.80 (d, *J* = 2.3 Hz, 1H), 7.20 (d, *J* = 16.4 Hz, 1H), 7.26-7.30 (m, 1H), 7.32-7.38 (m, 3H), 7.55 (d, *J* = 7.4 Hz, 2H); **¹³C NMR** (100 MHz, CDCl₃): δ 55.6, 56.0, 94.1, 97.4, 101.4, 115.7, 124.4, 127.2, 128.8, 133.5, 136.1, 143.4, 163.2, 163.9; **Analysis:** C₁₇H₁₅NO₂ requires C, 76.96; H, 5.70; N, 5.28; found: C, 76.92; H, 5.68; N, 5.24%.

(*R*)-3-((*R*)-Hydroxy(phenyl)methyl)-5,7-dimethoxyisobenzofuran-1-(3*H*)-one (**86**)

A 50 mL RB flask was charged with K₃Fe(CN)₆ (1 g, 3 mmol), K₂CO₃ (414 mg, 3 mmol), *tert*-BuOH (2.5 mL), THF (2.5 mL) and H₂O (5 mL) and stirred for 10 min. Subsequently, (DHQD)₂PHAL (8 mg, 1 mol%) and K₂OsO₄·2H₂O (2 mg, 0.5 mol%) were added and the stirring continued for additional 30 min. To this reaction mixture, *o*-cyanostilbene derivative **85** (265 mg, 1 mmol) was added and allowed to stir for 7 h at 25 °C. After completion of reaction (as monitored by TLC), sodium bisulphite (1 g) was added slowly at 0 °C. The organic layer was separated, aqueous layer extracted

with ethyl acetate (3 x 10 ml) and the combined organic layers washed with brine (15 mL), dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to yield the crude product. Flash column chromatographic purification [silica gel (230-400 mesh) and pet. ether:EtOAc (7:3) as an eluent] gave **86** (221 mg).

Yield: 93%; colorless solid; **mp:** 170-172 °C; $[\alpha]_{25}^D$ -77.56 (*c* 1.15, CHCl₃); 99% ee from chiral HPLC analysis (Chiracel OJ-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 18.27 min (99.36%) and 20.40 min (0.64%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 698, 759, 947, 1041, 1077, 1204, 1336, 1461, 1625, 1754, 2981, 3018, 3444; **¹H NMR** (500 MHz, CD₃OD): δ 3.61 (s, 3H), 3.84 (s, 3H), 4.77 (d, *J* = 6.4 Hz, 1H), 5.45 (d, *J* = 6.4 Hz, 1H), 5.78 (d, *J* = 1.7 Hz, 1H), 6.35 (d, *J* = 1.7 Hz, 1H), 7.29-7.32 (m, 5H); **¹³C NMR** (125 MHz, CDCl₃+CD₃OD): δ 56.1, 76.0, 83.9, 99.9, 102.9, 107.6, 128.2, 128.8, 129.0, 139.2, 152.0, 159.9, 167.1, 169.8; **HRMS** (ESI) *m/z*. calcd for C₁₇H₁₆O₅ [M + Na]⁺: 323.0890, found: 323.0850; **Analysis:** C₁₇H₁₆O₅ requires C, 67.99; H, 5.37; found: C, 67.85; H, 5.29%.

(R)-5, 7-Dihydroxy-3-((R)-hydroxy(phenyl)methyl)isobenzofuran-1(3H)-one:
[(-)-Matteucen C] (62)

To a solution of phthalide **86** (0.17 mmol, 50 mg) in CH₂Cl₂ (5 mL) at - 78 °C was added BBr₃ (1.36 mL, 1.36 mmol, 1 M in CH₂Cl₂) over 10 min. The reaction mixture was allowed to warm to 25 °C and then stirred for 24 h. It was quenched with sat. aq. NaHCO₃ (5 mL). The aqueous layer washed with CH₂Cl₂ and extracted with ethyl acetate (3 x10 mL). The combined organic layers were washed with brine (10 mL) and dried over anhyd. Na₂SO₄. After concentration, the crude product was purified by silica gel column chromatography to give **62** (31 mg).

Yield: 68%; colorless powder; **mp:** 135-147 °C; $[\alpha]_{25}^D$ -54.16 (*c* 1.0, CH₃OH); **IR** (CHCl₃, cm⁻¹): ν_{\max} 691, 710, 1169, 1615, 1684, 1725, 3364; **¹H NMR** (500 MHz,

DMSO- d_6): δ 4.94 (t, $J = 4.8$ Hz, 1H), 5.44 (d, $J = 4.0$ Hz, 1H), 5.73 (d, $J = 4.8$ Hz, 1H), 6.23 (d, $J = 1.8$ Hz, 1H), 6.25 (d, $J = 1.8$ Hz, 1H), 7.27-7.36 (m, 5H), 10.30 (s, 1H), 10.33 (s, 1H); ^{13}C NMR (125 MHz, DMSO- d_6): δ 72.6, 81.7, 101.1, 102.3, 104.2, 126.9, 127.3, 127.6, 140.7, 151.5, 157.6, 163.9, 167.7; **Analysis:** $\text{C}_{15}\text{H}_{12}\text{O}_5$ requires C, 66.17; H, 4.44; found: C, 66.09; H, 4.39%.

(E)-1-Bromo-2-(pent-1-enyl)benzene (52w)

To a stirred solution of sulfone **87** (1 g, 3.78 mmol) in dry THF (20 mL) at -78 °C under N_2 atmosphere was added drop-wise NaHMDS (4.15 mL, 4.15 mmol), the mixture was stirred for 30 min followed by addition of neat *o*-bromobenzaldehyde (**66**) (0.3 mL, 5.6 mmol). After being stirred for 3 h at -78 °C, the mixture was allowed to warm slowly to 25 °C and stirred for 10 h, finally quenched with H_2O . The aqueous layer was extracted with EtOAc (3x25 mL). The combined organic extracts were dried over anhyd. Na_2SO_4 and concentrated under reduced pressure to get the crude alkene, which was purified by column chromatography on silica gel with pet. ether:EtOAc (95:5) to give pure (*E*)-*o*-bromoalkene **52w** (0.700 g, 82%).

Yield: 82%; colorless oil; **IR** (CHCl_3 , cm^{-1}): ν_{max} 671, 757, 929, 1025, 1216, 1435, 1465, 1588, 1647, 2870, 2929; ^1H NMR (200 MHz, CDCl_3): δ 0.97 (t, $J = 7.3$ Hz, 3H), 1.47-1.54 (m, 2H), 2.18-2.29 (m, 2H), 6.15 (td, $J = 6.9, 15.6$ Hz, 1H), 6.69 (d, $J = 15.6$, 1H), 7.04 (dt, $J = 1.8, 7.6$ Hz, 1H), 7.22 (t, $J = 7.6$ Hz, 1H), 7.44-7.53 (m, 2H); ^{13}C NMR (50 MHz, CDCl_3): δ 13.7, 22.2, 35.2, 123.4, 126.8, 127.4, 128.1, 128.8, 132.8, 134.1, 137.7; **Analysis** for $\text{C}_{11}\text{H}_{13}\text{Br}$ requires: C, 58.69; H, 5.82%; found: C, 58.47; H, 5.69%.

(E)-2-(Pent-1-en-1-yl)benzotrile (50w)

o-Bromoalkene **52w** (0.7 g, 3.1 mmol) was taken up in dry DMF (10 mL) and CuCN (0.83 g, 9.3 mmol) was added and the mixture refluxed under N_2 for 14 h (monitored

by TLC). The reaction mixture was then cooled to 25 °C and then diluted with water (30 mL) and EtOAc (25 mL). The organic layer was separated and the aqueous layer was extracted with EtOAc (3 x 20 mL). The combined organic extracts were dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to get the crude *o*-cyanoalkene, which was purified by column chromatography on silica gel with pet. ether:EtOAc (80:20) to give pure *o*-cyanoalkene **50w** (0.461 g, 87%).

Yield: 87%, colorless solid; **mp:** 126-128 °C; **IR** (CHCl₃, cm⁻¹): ν_{\max} 756, 857, 974, 1037, 1095, 1184, 1202, 1216, 1251, 1275, 1291, 1319, 1347, 1368, 1393, 1444, 1477, 1573, 1607, 1640, 1716, 2219, 2984, 3023; **¹H NMR** (200 MHz, CDCl₃): δ 0.98 (t, *J* = 7.3 Hz, 3H), 1.48-1.60 (m, 2H), 2.22-2.33 (m, 2H), 6.43 (dt, *J* = 6.8, 15.3 Hz, 1H), 6.74 (d, *J* = 15.3 Hz, 1H), 7.25 (dd, *J* = 15.3, 1.4 Hz, 1H), 7.45-7.62 (m, 3H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.7, 22.2, 35.2, 110.5, 117.9, 125.2, 126.0, 126.8, 132.5, 132.7, 136.4, 141.1; **Analysis:** C₁₂H₁₃N requires C, 84.17; H, 7.65; N, 8.18; found: C, 84.14; H, 7.61; N, 8.15%.

(R)-3-((R)-1-Hydroxybutyl)isobenzofuran-1(3H)-one (51w)

A 50 mL RB flask was charged with K₃Fe(CN)₆ (1 g, 3 mmol), K₂CO₃ (414 mg, 3 mmol), *tert*-BuOH (2.5 mL), THF (2.5 mL) and H₂O (5 mL) and stirred for 10 min. Subsequently, (DHQD)₂PHAL (8 mg, 1 mol%) and K₂OsO₄·2H₂O (2 mg, 0.5 mol%) were added and the stirring continued for additional 30 min. To this reaction mixture, *o*-cyanoalkene **50w** (172 mg, 1 mmol) was added and allowed to stir for 7 h at 25 °C. After completion of reaction (as monitored by TLC), sodium bisulphite (1 g) was added slowly at 0 °C. The organic layer was separated, aqueous layer extracted with ethyl acetate (3 x 10 ml) and the combined organic layers washed with brine (15 mL), dried over anhyd. Na₂SO₄ and concentrated under reduced pressure to yield the crude

product. Flash column chromatographic purification [silica gel (230-400 mesh) and pet. ether:EtOAc (75:25) as an eluent] gave phthalide **51w** (0.191 g, 93%).

Yield: 93%; colorless solid; **mp:** 103-109 °C; $[\alpha]_{25}^D$ -76.89 (*c* 1, CHCl₃); 97% ee by Mosher's ester analysis; **IR** (CHCl₃, cm⁻¹): ν_{\max} 694, 728, 1080, 1212, 1287, 1467, 1618, 1752, 2873, 2959, 3433; **¹H NMR** (200 MHz, CDCl₃): δ 0.93 (t, *J* = 6.8 Hz, 3H), 1.44-1.72 (m, 4H), 1.97 (br s, 1H), 3.99 (br s, 1H), 5.40 (d, *J* = 3.6 Hz, 1H), 7.51-7.57 (m, 2H), 7.65-7.73 (m, 1H), 7.87-7.92 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 13.9, 18.8, 34.9, 71.9, 83.2, 122.4, 125.6, 126.6, 129.2, 134.0, 147.2, 170.5; **Analysis:** C₁₂H₁₄O₃ requires C, 69.88; H, 6.84; found: C, 69.82; H 6.81%.

Mosher's ester of (R)-3-((R)-1-Hydroxybutyl)isobenzofuran-1(3H)-one (88)

A two-neck 10 mL flask with septum was charged with (38 mg, 0.18 mmol) *N,N'*-dicyclohexylcarbodiimide (DCC), catalytic amount of 4-dimethylaminopyridine (DMAP) and CH₂Cl₂ (2 mL) under argon atmosphere. The flask was allowed to cool at 0 °C for 10 min and a solution of alcohol **51w** (34 mg, 0.16 mmol) in CH₂Cl₂ (2 mL) was introduced through a syringe. It was allowed to stir for additional 10 min, followed by dropwise addition of (*R*)- α -methoxy- α -trifluoromethylphenyl acetic acid (42 mg, 0.176 mmol) in CH₂Cl₂ (2 mL). The reaction mixture was then stirred at 0 °C for additional one hour and then at room temperature for overnight. The reaction mixture was diluted with CH₂Cl₂ (50 mL), washed with saturated NaHCO₃ solution (50 mL), dried over anhyd. Na₂SO₄ and then concentrated under reduced pressure to get Mosher's ester **88** (53 mg, 80%) as a thick syrup.

Yield: 80%; $[\alpha]_{25}^D$ -42.5 (*c* 0.4, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 650, 737, 1016, 1154, 1243, 1410, 1496, 1644, 1754, 2255, 2851, 2930, 2951, 3069, 3156; **¹H NMR** (400 MHz, CDCl₃): δ 0.93 (t, *J* = 6.8 Hz, 3H), 1.26-1.41 (m, 2H), 1.73-1.85 (m, 2H), 3.16 (s, 3H), 5.56 (d, *J* = 3.6 Hz, 1H), 5.63-5.71 (m, 1H), 7.17-7.35 (m, 5H), 7.46-7.52 (m,

2H), 7.67 (td, $J = 1.13, 7.4$ Hz, 1H), 7.89 (d, $J = 7.6$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 14.1, 19.3, 33.8, 37.5, 49.9, 55.2, 80.1, 92.8 (d, $J = 36.3$ Hz), 123.8 (d, $J = 271.4$ Hz), 125.8, 127.5, 128.1, 129.1, 131.9, 137.2, 148.1, 165.2, 170.6; **Analysis:** $\text{C}_{23}\text{H}_{23}\text{F}_3\text{O}_4$ requires C, 65.71; H, 5.51; found: C, 65.62; H, 5.34%.

(S)-3-Butylisobenzofuran-1(3H)-one: [3-Butylphthalide] (1)

Under a nitrogen atmosphere, 1,1-thiocarbonyldiimidazole (233 mg, 1.818 mmol) was added to a solution of **51w** (250 mg, 1.212 mmol) and DMAP (22 mg, 0.121 mmol) in CH_2Cl_2 (10 mL). After stirring for 18 h at room temperature, solvent was removed *in vacuo*. To this were added AIBN (20 mg, 0.121 mmol) and tributyltin hydride (1.6 mL, 6.06 mmol) in toluene (15 mL) and the mixture was refluxed for 15 min. It was then diluted with ethyl acetate and successively washed with water and brine. The organic layer was dried with anhyd. Na_2SO_4 and concentrated *in vacuo*. The residue was chromatographed twice on silica gel (pet. ether:EtOAc 80:20) to give butylphthalide (163 mg, 86%).

Yield: 86%; colourless oil; $[\alpha]_{25}^{\text{D}} -67.0$ (c 1.15, CHCl_3); lit.⁴⁹ $[\alpha]_{25}^{\text{D}} -64.7$ (c 1.06, CHCl_3); **IR** (CHCl_3): 780, 1346, 1465, 1526, 1716, 2932 cm^{-1} ; **^1H NMR** (200 MHz, CDCl_3): δ 0.88-0.95 (m, 3H), 1.32-1.53 (m, 4H), 1.68-1.86 (m, 1H), 1.97-2.07 (m, 1H), 5.46 (dd, $J = 4.3, 7.9$ Hz, 1H), 7.42 (dd, $J = 1.13, 7.6$ Hz, 1H), 7.52 (t, $J = 7.4$ Hz, 1H), 7.66 (td, $J = 1.13, 7.4$ Hz, 1H), 7.90 (d, $J = 7.6$ Hz, 1H); **^{13}C NMR** (CDCl_3): δ 13.8, 22.5, 26.9, 34.4, 81.3, 121.8, 125.5, 128.9, 133.9, 150.1, 170.3; **Analysis:** $\text{C}_{12}\text{H}_{14}\text{O}_2$ requires C, 75.76; H, 7.42; found: C, 75.72; H, 7.39%.

2.2.7 References

1. For recent reviews of synthesis and biological applications of phthalides, see: (a) Lin, G.; Chan, S. S. -K.; Chung, H. -S.; Li, S. L. *Stud. Nat. Prod. Chem.* **2005**, 611; (b) Beck, J. J.; Chou, S. C. *J. Nat. Prod.* **2007**, *70*, 891; (c) Xioang, M. J.; Li, Z. H. *Curr. Org. Chem.* **2007**, *11*, 833; (d) Liu, J.; Li, F.; Kim, E. L.; Li, J. L.; Hong, J.; Bae, K. S.; Chung, H. Y.; Kim, H. S.; Jung, J. H. *J. Nat. Prod.* **2011**, *74*, 1826.
2. (a) Barton, D. H. R.; De Vries, J. X. *J. Chem. Soc.* **1963**, 1916; (b) Xu, Z.; Hu, G. Y.; Tan, G. S. *Chin. J. Modern Med.* **2004**, 93; (c) Sato, H.; Yorozu, H.; Yamaoka, S. *Biomed. Res.* **1993**, *14*, 385; (d) Wang, X. W. *Drugs Future* **2000**, *25*, 16.
3. Yoganathan, K.; Rossant, C.; Ng, S.; Huang, Y.; Butler, M. S.; Buss, A. D. *J. Nat. Prod.* **2003**, *66*, 1116.
4. Chatterjee, P.; Franklin, M. R. *Drug Metab. Dispos.* **2003**, *31*, 1391.
5. Zhou, J.; Gupta, K.; Yao, J.; Ye, K.; Panda, D.; Giannakakaou, P.; Joshi, H. C. *J. Biol. Chem.* **2002**, *277*, 39777.
6. (a) Palermo, J. A.; Rodriguez, B.; Maria, F.; Spagnuolo, C.; Seldes, A. M. *J. Org. Chem.* **2000**, *65*, 4482; (b) Li, J.; Li, L.; Si, Y.; Jiang, X.; Guo, L.; Che, Y. *Org. Lett.* **2011**, *13*, 2670.
7. (a) Barton, D. H. R.; de vries, J. H. *J. Chem. Soc.* **1963**, 1916; (b) Xu, Z.; Hu, G. Y.; Tan, G. S. *Chin. J. Modern Med.* **2004**, 93; (c) Yu, S.; You, S.; Chen, H. *Yaoxue Xuebao* **1984**, *101*, 486; (d) Sato, H.; Yorozu, H.; Yamaoka, S. *Biomed. Res.* **1993**, *14*, 385; (e) Wang, X. *Drugs Future* **2000**, *25*, 16; (f) Xu, Z.; Hu, G. Y.; Tan, G. S. *Chin. J. Modern Med.* **2004**, 93; (g) Ramachandran, P. V.; Guang-ming C.; Brown, H. C. *Tetrahedron Lett.* **1996**, *37*, 2205.
8. Ohkuma, T.; Kitamura, M.; Noyori, R. *Tetrahedron Lett.* **1990**, *31*, 5509.
9. Watanabe, M.; Hashimoto, N.; Araki, S.; Butsugan, Y. *J. Org. Chem.* **1992**, *57*, 742.
10. Lei, J.-G.; Hong, R.; Yuan, S.-G.; Lin, G.-Q. *Synlett* **2002**, 927.
11. Ohezeki, T.; Mori, K. *Biosci. Biotechnol. Biochem.* **2003**, *67*, 2584.
12. (a) Tanaka, K.; Nishida, G.; Wada, A.; Noguchi, K. *Angew. Chem., Int. Ed.* **2004**, *43*, 6510; (b) Tanaka, K.; Osaka, T.; Noguchi, K.; Hirano, M. *Org. Lett.* **2007**, *9*, 1307
13. Lipka, E.; Vacher, M-P.; Vaccher, C.; Len, C. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 501.
14. Chang, H. T.; Jaganmohan, M.; Cheng, C. H. *Chem. Eur. J.* **2007**, *13*, 4356
15. Zhang, B.; Xu, M. H.; Lin, G. Q. *Org. Lett.* **2009**, *11*, 4712.
16. Phan, D. H. T.; Kim, B.; Dong V. M. *J. Am. Chem. Soc.* **2009**, *131*, 15608.
17. Zhang, H.; Zhang, S.; Liu, L.; Luo, G.; Duan, W.; Wang, W. *J. Org. Chem.* **2010**, *75*, 368.
18. Mangas-Sánchez, J.; Busto, E.; Gotor-Fernández, V.; Gotor, V. *Org. Lett.* **2012**, *14*, 1444.
19. (a) Jacobsen, E. N.; Marko, I.; France, M. B.; Svendsen, J. S.; Sharpless, K. B. *J. Am. Chem. Soc.* **1989**, *111*, 737; (b) Kolb, H. C.; Anderson, P. G.; Bennani, Y. L.; Crispino, G. A.; Jeong, K. S.; Kwong, H. L.; Sharpless, K. B. *J. Am. Chem. Soc.* **1993**, *115*, 12226.
20. Hentges, S. G.; Sharpless, K. B. *J. Am. Chem. Soc.* **1980**, *102*, 4263.
21. (a) Criegee, R. *Justus Liebigs Ann. Chem.* **1996**, 522, 75; (b) Criegee, R.; Marchand, B.; Wannowias, H. *Justus Liegs. Ann. Chem.* **1942**, 550, 99; (c) Sharpless, K. B.; Teranishi, A. Y.;

- Backwall, J. E. *J. Am. Chem. Soc.* **1977**, *99*, 3120; (d) Jorgensen, K. A.; Schioett, B. *Chem. Rev.* **1990**, *90*, 1483.
22. Gawley, R. A.; Aube, J. *Principles of Asymmetric Synthesis* Elsevier Science (Oxford), **1996**, Vol. 14, (Chap. 8), pp 314.
23. Sharpless, K. B.; Amereg, W.; Bennani, Y. L.; Crispino, G. A.; Hartung, J.; Jeong, K. S.; Kwong, H. L.; Morikawa, K.; Wang, Z. M.; Xu, D.; Zhang, X. L. *J. Org. Chem.* **1992**, *57*, 2768.
24. Amereg, W.; Bennani, Y. L.; Chadha, R. K.; Crispino, G. A.; Davis, W. D.; Hartung, J.; Jeong, K. S.; Ogino, Y.; Shibata, T.; Sharpless, K. B. *J. Org. Chem.* **1993**, *58*, 844.
25. (a) Jagdale, A. R.; Reddy, R. S.; Sudalai, A. *Org. Lett.* **2009**, *11*, 803; (b) Jagdale, A. R.; Reddy, R. S.; Sudalai, A. *Tetrahedron: Asymmetry* **2009**, *20*, 335.
26. (a) Bower, J. F.; Szeto, P.; Gallagher, T. *Chem Commun.* **2005**, 5793; (b) Zhai, H.; Luo, S.; Ye, C.; Ma, Y. *J. Org. Chem.* **2003**, *68*, 8268; (c) Fuchs, J. R.; Funk, R. L. *Org. Lett.* **2001**, *3*, 3923. (d) Katoh, M.; Inoue, H.; Suzuki, A.; Honda, T. *Synlett* **2005**, *18*, 2820; (e) Bower, J. F.; Szeto, P.; Gallagher, T. *J. Org. Biomol. Chem.* **2007**, *5*, 143.
27. Uchida, K.; Watanabe, H.; Usui, T.; Osada, H.; Kitahara, T. *Heterocycles* **1998**, *48*, 2049.
28. (a) Gensler, W. *J. Chem. Rev.*, **1957**, *57*, 191; (b) Julia, M.; Paris, J.-M. *Tetrahedron Lett.* **1973**, *14*, 4833.
29. (a) Dupau, P.; Epple, R.; Thomas, A. A.; Fokin, V. V.; Sharpless, K. B. *Adv. Synth. Catal.* **2002**, *344*, 421; (b) Fujita, M.; Yoshida, Y.; Miyata, K.; Wakisaka, A.; Sudimura, T. *Angew. Chem. Int. Ed.* **2010**, *49*, 7068.
30. (a) Li, G.; Chang, H.-T.; Sharpless, K. B. *Angew. Chem. Int. Ed.* **1996**, *35*, 451; (b) Sharpless, K. B.; Rudolph, J.; Sennhenn, P. C.; Vlaar, C. P. *Angew. Chem. Int. Ed.* **1996**, *35*, 2810.
31. Deubel, D. V.; Muniz, K. *Chem. Eur. J.* **2004**, *10*, 2475.
32. Baldwin, J. E. *J. Chem. Soc., Chem. Commun.* **1976**, 734.
33. (a) Flora of China Editorial Committee, "Flora Reipublicate Popularis Sinicae," Vol. 4, Science Press, Beijing, **2006**, pp. 159; (b) Jiangsu Institute of Botany, "Xinhua Herbal Schem," Shanghai Scientific and Technologic Press, Shanghai, **1990**, pp.684.
34. Shao, P.; Zhang, W.; Li, B.; Jiao, W. H.; Wu, L. J.; Yao, X. S. *Chem. Pharm. Bull.* **2010**, *58*, 1650.
35. Hankey, G. J. *J. Thromb. Haemost.* **2005**, *3*, 1638.
36. Donnan, G. A.; Fisher, M.; Macleod, M.; Davis, S. M. *Lancet* **2008**, *371*, 1612.
37. (a) Markus, H. S. *Hum. Mol. Genet.* **2011**, *20*, 124; (b) Kraft, P.; Schwarz, T.; Meijers, J. C.; Stoll, G.; Kleinschnitz, C. *PLoS One* **2010**, *5*, e11658; (c) Andelario, J. E. *Curr. Opin. Invest. Drugs* **2009**, *10*, 644.
38. (a) Mackman, N. *Nature* **2008**, *451*, 914; (b) Zhang, Y.; Wang, L.; Li, J.; Wang, X. L. *J. Pharmacol. Exp. Ther.* **2006**, *317*, 973.
39. (a) Huang, J. Z.; Chen, Y. Z.; Su, M. *Neurosci. Lett.* **2010**, *475*, 89; (b) Li, L.; Zhang, B.; Tao, Y. *Brain Res.* **2009**, *1290*, 91.
40. Liu, C. L.; Liao, S. J.; Zeng, J. S. *J. Neurol. Sci.* **2007**, *260*, 106.
41. (a) Zhang, T.; Jia, W.; Sun, X. *Neurol. Res.* **2010**, *32*, 390; (b) Li, J.; Li, Y.; Ogle, M. *Brain Res.*

- 2010**, 1359, 216.
42. Xu, H. L.; Feng, Y. P. *Acta Pharmacol Sin.* **2000**, 21, 433.
43. (a) Ying, P.; Jing, S.; Stephanie, H.; Alyssa, N. N.; Weiming, X.; Yipu, F.; Xiaoliang, W.; Cynthia, A. L. *J. Neurosci.*, **2010**, 30, 8180; (b) Yaping, H.; Yanhong, D.; Jing, X.; Nan, M.; Chunfeng, S.; Wenbin, L.; Peiyuan, L. V. *Neural Reg. Res.* **2013**, 8, 1733.
44. (a) Dubois, M. F.; Hebert, R. *Neuroepidemiology* **2001**, 20, 179; (b) Yokota, O.; Sasaki, K.; Fujisawa, Y. *Eur. J. Neurol.* **2005**, 12, 782; (c) Suh, G. H.; Shah, A. *Acta Psychiatr Scand*, **2001**, 104, 4.
45. Takabashi, H.; Tsubuki, T.; Higashiyama, K. *Chem. Pharm. Bull.* **1991**, 39, 3136.
46. Soai, K.; Hori, K.; Karahara, M. *Tetrahedron: Asymmetry* **1991**, 2, 253.
47. Matsui, S.; Uejima, A.; Suzuki, Y.; Tanakab, K. *J. Chem. Soc. Perkin Trans 1*, **1993**, 701.
48. Kitayama, T. *Tetrahedron: Asymmetry* **1997**, 8, 3765.
49. Kosaka, M.; Sekigughi, S.; Naito, J.; Uemure, J.; Kuwahara, S.; Watanabe, J.; Harada, N.; Hiroi, K. *Chirality* **2005**, 17, 218.
50. Coric, I.; Muller, S.; List, B. *J. AM. Chem. Soc.* **2010**, 132, 17370.
51. Reddy, R. S.; Kiran, I. N. C.; Sudalai A. *Org. Biomol. Chem.* **2012**, 10, 3655.
52. Blakemore, P. R.; Cole, W. J.; Kocienski, P. J.; Morley, A. *Synlett*, **1998**, 26.
53. Liao, C. C.; Liu, W.C. *Chem. Commun.* **1999**, 117.

CHAPTER 3

A New Concise Method for the Synthesis of Chiral 3-Amino-1,2-Diols, (+)-*epi*-Cytosazone and Formal Synthesis of *N*-Thiolated-2-Oxazolidinone via Proline-Catalyzed α -Amino oxylation of β -Aminoaldehydes

Section I:

Proline-Catalyzed α -Aminooxylation of β -Aminoaldehydes: A Highly Stereoselective Synthesis of 3-Amino-1,2-Alkane Diols

3.1.1 Introduction

The enantiomerically pure *syn*- and *anti*-3-amino-1,2-alkane diols are ubiquitous substructures associated with biologically active natural products.¹ They are important and versatile ‘building blocks’ for asymmetric synthesis of bioactive pharmaceuticals and complex bioactive molecules. Some representative examples of biologically active and pharmacologically relevant therapeutic agents, wherein 3-amino-1,2-alkane diols play a vital role as key intermediates in their synthesis are shown in **Fig. 1**. For example, taxol (**1**) is a mitotic inhibitor drug used in cancer chemotherapy and AIDS-related Kaposi's sarcoma.²

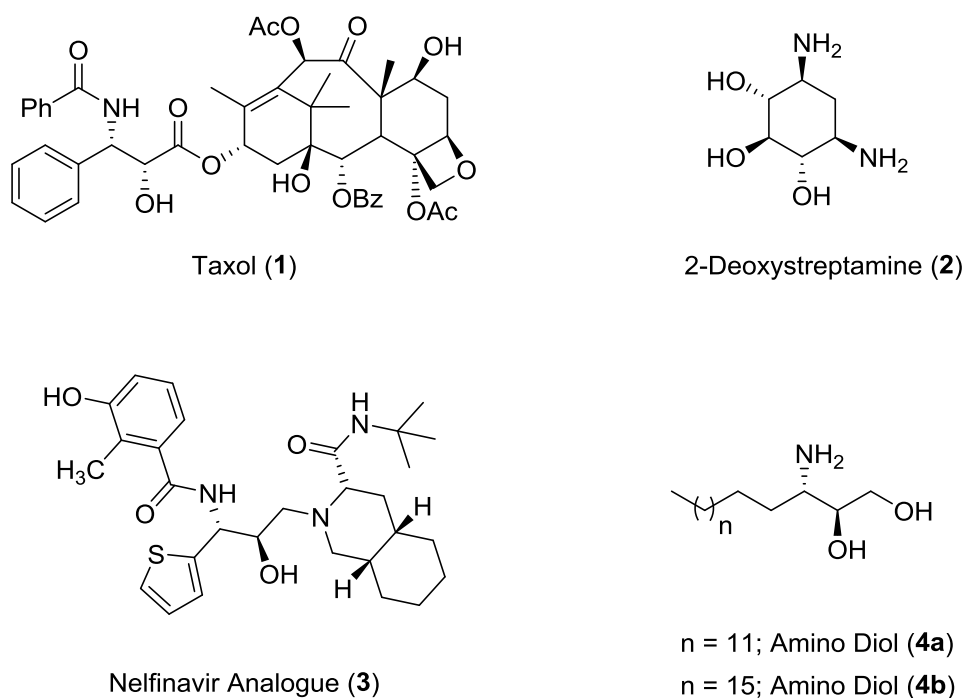
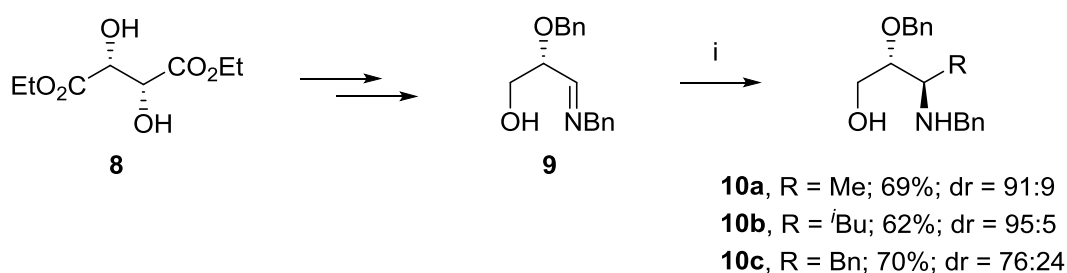


Fig. 1: Structures of bioactive 3-amino-1,2-alkane diols

Jagers's approach (1994)^{7a}

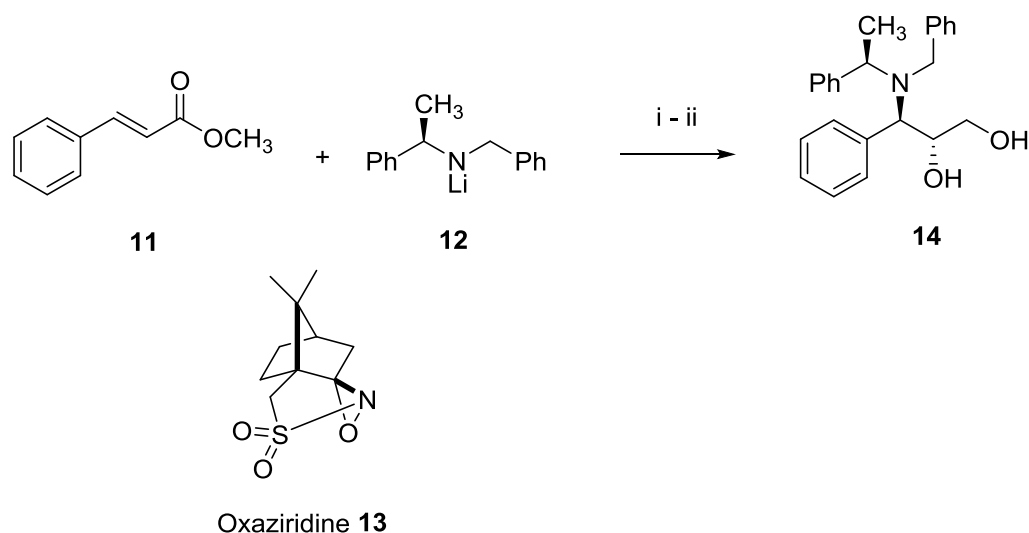
Jagers *et al.* have achieved the synthesis of optically active 3-amino-1,2-alkane diols **10a-c** using a chiral pool approach commencing from (+)-diethyl tartrate (**8**), which was converted to glycerinealdimine **9** using simple standard transformations.^{7b} Stereoselective Grignard addition of alkyl magnesium halides onto glycerinealdimine **9** in the presence of CeCl₃ provided *anti*-amino diols **10a-c** as major diastereomers (de = 52-90%) in 62-70% yield (**Scheme 2**).



Scheme 2: (i) RMgX, CeCl₃, THF, 0 °C to 25 °C, 12 h.

Bunnage's approach (1994)⁸

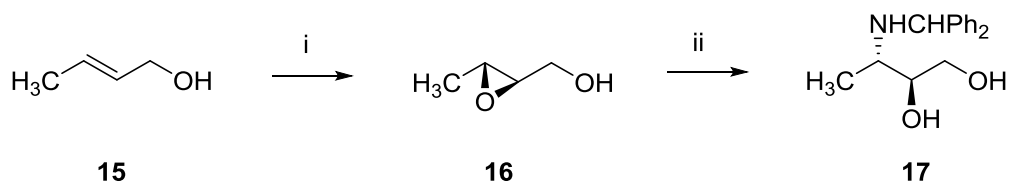
Bunnage *et al.* have reported a useful method for the synthesis of 3-amino-1,2-alkane diol **14** using diastereoselective conjugate addition of amine source with enoate acceptor **11** as key step. Thus, tandem diastereoselective conjugate addition of lithium N-benzylamide **12** with methyl cinnamate **11**, followed by the electrophilic hydroxylation of the resultant β -amino enolates with camphorsulfonyl oxaziridine **13** resulted in β -amino- α -hydroxyester. Subsequent reduction of ester with LiAlH₄ provided *anti*-amino diol **14** with good diastereoselectivity (> 90% d.e.) and moderate yield (**Scheme 3**).



Scheme 3: (i) THF, $-78\text{ }^{\circ}\text{C}$, 2 h then oxaziridine (**13**), $-78\text{ }^{\circ}\text{C}$ to $0\text{ }^{\circ}\text{C}$, 1 h, 43%; (ii) LiAlH_4 , THF, $0\text{ }^{\circ}\text{C}$ to $25\text{ }^{\circ}\text{C}$, 2 h, 72%.

Riera's approach (1995)⁹

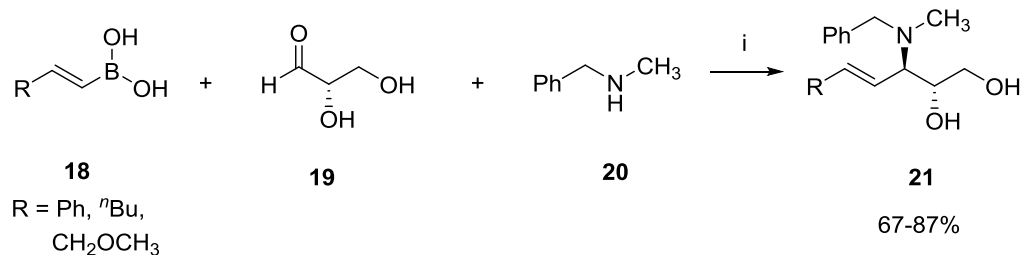
Riera *et al.* have used Sharpless asymmetric epoxidation as the key reaction for the introduction of chirality. Thus, commercially available (*E*)-crotyl alcohol **15** was subjected to Sharpless asymmetric epoxidation with D-(-)-DIPT to give epoxy alcohol **16**. $\text{Ti}(\text{O}^i\text{Pr})_4$ mediated regioselective ring opening of chiral epoxy alcohol **16** by primary amine provided (2*S*, 3*S*)-3-benzhydrylamino-1,2-butanediol (**17**) in 68% yield with 92% ee (**Scheme 4**). Regioselective opening of chiral epoxides with amine source have been extensively used for many bioactive molecule syntheses, especially antiHIV Protease inhibitors.



Scheme 4: (i) D-(-)-DIPT, $\text{Ti}(\text{O}^i\text{Pr})_4$, *tert*-BuOOH, CH_2Cl_2 , $-25\text{ }^{\circ}\text{C}$; (ii) Ph_2CHNH_2 , $\text{Ti}(\text{O}^i\text{Pr})_4$, CH_2Cl_2 , $25\text{ }^{\circ}\text{C}$, 24 h, 68% (over two steps).

Petasis's approach (1998)¹⁰

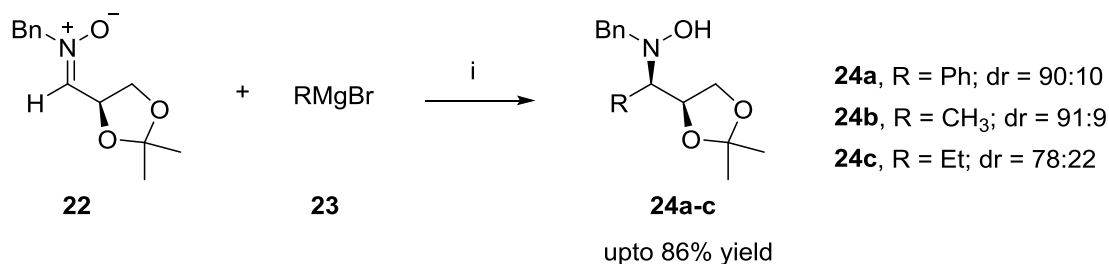
Petasis *et al.* have described the enantioselective synthesis of 3-amino-1,2-alkane diols using chiral pool approach. This method involves one-step three component variant of Mannich reaction involving organoboronic acids **18**, α -hydroxy aldehyde **19** and an amine **20** under milder reaction conditions that gave directly the corresponding *anti*-3-amino-1,2-alkane diols **21** in 99% de and ee (**Scheme 5**).



Scheme 5: (i) EtOH, sealed tube, 25 °C, 24 h.

Merino's approach (1998)¹¹

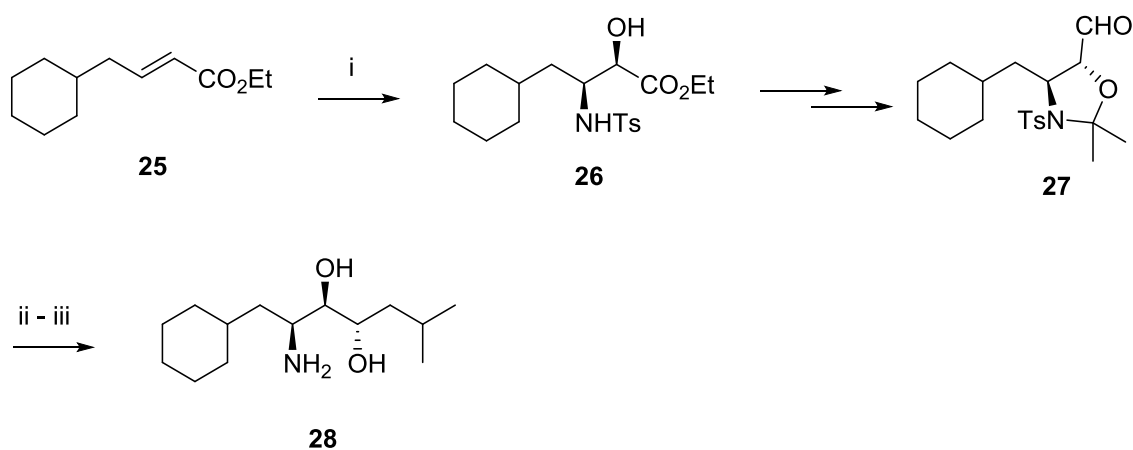
Merino *et al.* have reported the synthesis of *syn*-3-amino-1,2-alkane diols **24** using a chiral pool approach commencing from glyceraldehyde nitron **22**, which was derived from D-glyceraldehyde by condensation with N-benzyl hydroxylamine. Stereoselective Grignard addition of alkyl magnesium halides **23** onto glyceraldehyde nitron **22** in the presence of ZnBr₂ provided *syn*-amino diols **24a-c** as major diastereomer (de = 66-82%) in 72-86% yield (**Scheme 6**).



Scheme 6: (i) ZnBr₂, Et₂O, -60 °C, 6 h.

Chandrasekhar's approach (1999)¹²

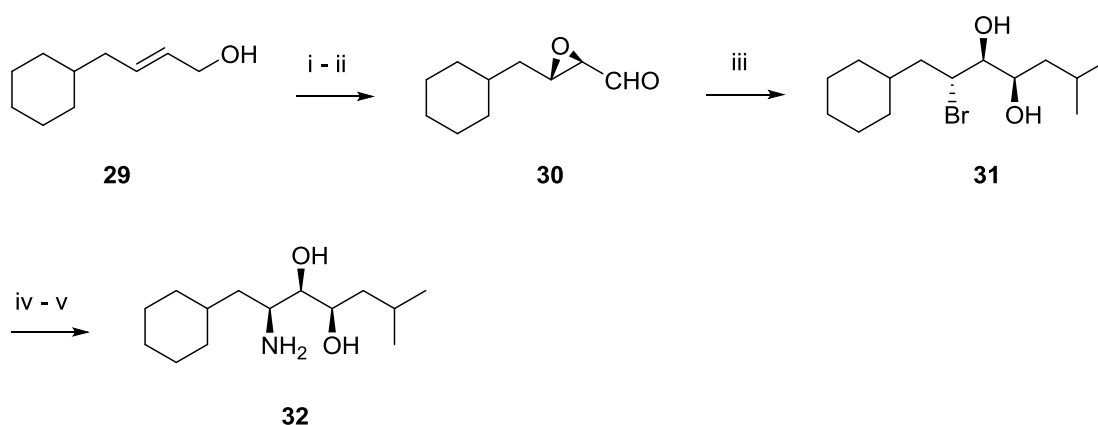
Chandrasekhar *et al.* have reported the synthesis of Abbott amino diol **28** using Sharpless asymmetric aminohydroxylation as the key step. Thus, α,β -unsaturated ester **25** was subjected to asymmetric aminohydroxylation using catalytic amount of quinine derived ligand (DHQ)₂PHAL, K₂OsO₂(OH)₄ and commercially available Chloramine-T to provide amino alcohol **26** in 65% yield and 89% ee. Acetonide protection of amino alcohol **26** followed by reduction of ester afforded aldehyde **27**, which on reaction with ⁱBuMgBr followed by deprotection of acetonide with HCl gave *syn,anti*-amino diol **28** as major diastereomer (dr = 80:20) (**Scheme 7**). Sharpless asymmetric aminohydroxylation has been extensively used for the introduction of chiral amino alcohols in the synthesis of many bioactive molecules. But the major drawback with this method is the poor regioselectivity of the newly introduced amino alcohol functionality.



Scheme 7: (i) chloroamine-T, K₂[OsO₂(OH)₄], (DHQ)₂PHAL, *tert*-BuOH:H₂O (1:1), 12 h, 25 °C, 65%; (ii) ⁱBuMgBr, THF, 0 °C to 25 °C, 6 h, 72%; (iii) 6 N HCl, CH₃OH, 25 °C, 4 h, 52%.

Righi's approach (2000)¹³

Righi *et al.* have used Sharpless asymmetric epoxidation as the key reaction for the synthesis of 3-amino-1,2-alkane diol **32**. Thus, allyl alcohol **29** was subjected to Sharpless asymmetric epoxidation using L-(+)-DET to give epoxy alcohol, which on subsequent oxidation provided epoxy aldehyde **30** in 68% yield with 93% ee. One-pot ring opening/organometallic addition of α,β -epoxy aldehyde **30** with *t*BuMgBr in the presence of MgBr₂.Et₂O afforded *syn*-diol **31** in stereocontrolled manner. Subsequent substitution of the bromine with azide, followed by catalytic hydrogenation to the amino group, led to *syn,syn*-3-amino-1,2-alkane diol **32** in 63% yield (over two steps) (Scheme 8).

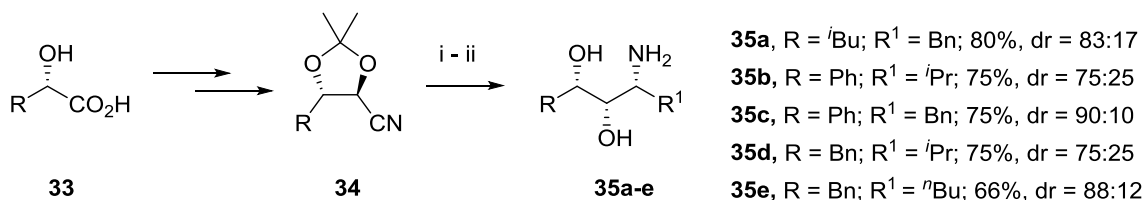


Scheme 8: (i) L-(+)-DET, Ti(O^{*i*}Pr)₄, *tert*-BuOOH, CH₂Cl₂, -25 °C, 24 h; (ii) TEMPO, PhI(OAc)₂, dry CH₂Cl₂, 25 °C, 1 h, 72% (over two steps); (iii) MgBr₂.Et₂O, CH₂Cl₂, *t*BuMgBr, THF, -50 °C, 24 h, 65%; (iv) NaN₃, DMF, 40 °C, 24 h, 71%; (v) 10% Pd/C, H₂ (50 psi), EtOAc, 24 h, 89%.

Larcheveque's approach (2000)^{14a}

Larcheveque *et al.* have reported the synthesis of *syn,syn*-3-amino-1,2-alkane diols **35a-e** using a chiral pool approach commencing from acetone protected *syn*-2,3-dihydroxynitriles **34**, which were derived from optically active 2-hydroxy acids **33**

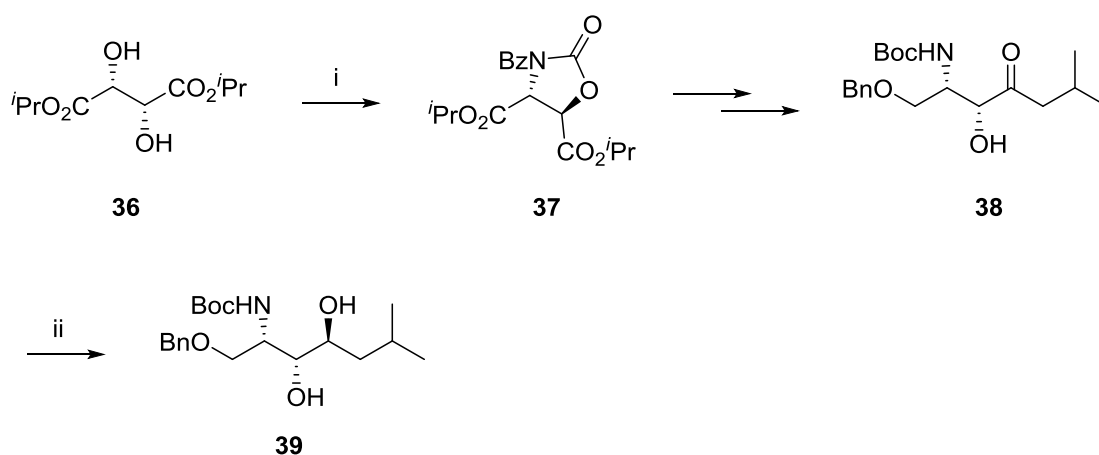
using simple standard transformation.^{14b} Stereoselective Grignard addition of alkyl magnesium bromide onto dihydroxynitriles **34**, followed by NaBH₄ reduction of the resulting imine, afforded the corresponding *syn*-amino diols **35a-e** as major diastereomer (de = 50-80%) in 66-80% yield (**Scheme 9**).



Scheme 9: (i) R¹MgBr, Et₂O, -15 °C, 6 h then NaBH₄, CH₃OH, 25 °C, 14 h; (ii) 2 N HCl, CH₃OH, H₂O, 25 °C, 3 h.

Ko's approach (2003)¹⁵

Ko *et al.* have reported the synthesis of *syn,anti*-3-amino-1,2-alkane diol **39** using a chiral pool approach.



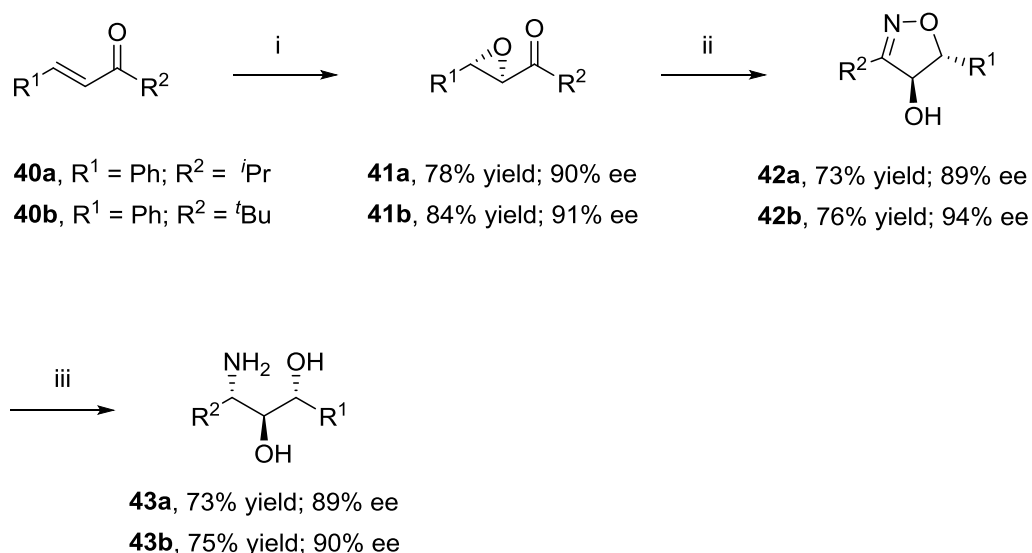
Scheme 10: (i) Bu₂SnO, CH₂Cl₂, reflux, Dean-Stark, 4 h, then BzNCS, Et₃N, Bu₄NBr, reflux, 2 h, 76%; (ii) NaBH(OAc)₃, CH₃CN:*n*-hexane (1.2:1), 3 h, 84%.

(+)-Diisopropyl tartrate **36** was converted to protected amino alcohol **37** in 76% yield using a three-step reaction sequence in one pot: (i) activation of diol **36** via tin

ketalization; (ii) iminocarbonate formation by treatment of tinketal with BzNCS; (iii) rearrangement of iminocarbonate under nucleophilic condition. Protected amino alcohol **37** was then converted to ketone **38** using simple standard transformations, where regioselective reduction of ester and Weinreb's amide formation served as key steps. Diastereoselective reduction of the carbonyl group **38** with NaBH(OAc)₃ afforded *syn,anti*-3-amino-1,2-alkane diol **39** in 8.5:1 diastereoselectivity and 84% yield (Scheme 10).

Bickley's approach (2003)¹⁶

Bickley *et al.* have used L-leucine-based chiral epoxidation as the key reaction for synthesis of *anti,anti*-3-amino-1,2-alkane diols **43a-b**.



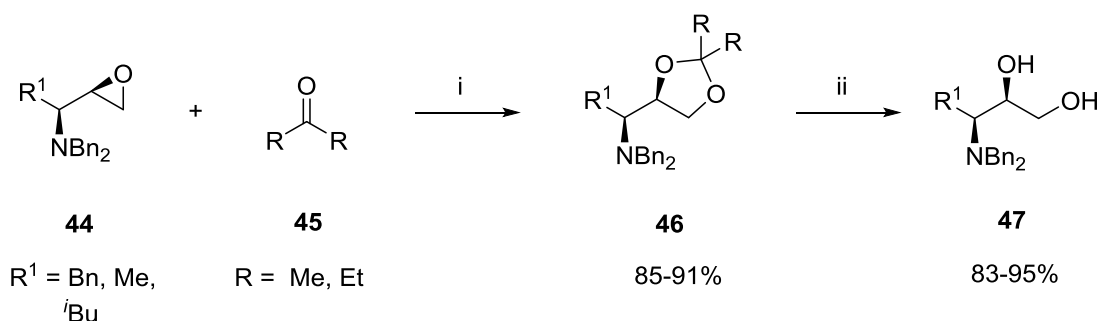
Scheme 11: (i) poly-L-leucine, THF, NH₂CONH₂.H₂O₂, DBU, 25 °C, 24 h; (ii) NH₂OH. HCl, EtOH, pyridine, reflux, 14 h; (iii) LiAlH₄, THF, 0 °C to 25 °C, 2 h.

Thus, enones **40a-b** were epoxidised using urea-hydrogen peroxide complex in the presence of poly-L-leucine supported on silica to yield epoxides **41a-b** in 90% ee. Dihydroisoxazoles **42a-b** were obtained in good yields by treatment of epoxides **41a-**

b with hydroxylamine. Subsequent diastereoselective reduction of dihydroisoxazoles **42a-b** with LiAlH_4 afforded *anti,anti*-3-amino-1,2-alkane diols **43a-b** in good yields (73-75%) with 90% ee (**Scheme 11**).

Concellon's approach (2005)¹⁷

Concellon *et al.* have reported the synthesis of *syn*-3-amino-1,2-alkane diol **47** using a chiral pool approach commencing from enantiopure *syn*-2-(1-aminoalkyl)epoxides **44**. Thus, epoxides **44** were treated with different ketones **45** in the presence of $\text{BF}_3 \cdot \text{Et}_2\text{O}$ to provide the corresponding 4-(1-aminoalkyl)-1,3-dioxolanes **46** in high yields and without epimerization. Finally, deprotection of 1,3-dioxolanes **46** with HCl afforded *syn*-3-amino-1,2-alkane diol **47** (**Scheme 12**).



Scheme 12: (i) $\text{BF}_3 \cdot \text{Et}_2\text{O}$, CH_2Cl_2 , 0 °C, 1 h; (ii) 1N HCl, CH_3CN , reflux, 1 h.

3.1.3 Present Work

3.1.3.1 Objective

As can be seen from the above discussion, the reported methods are quite effective. However, there are certain serious short-comings associated with them such as: (i) dependence on chiral pool resources; (ii) expensive chiral ligands, catalysts and reagents; (iii) multi-step reaction sequences; (iv) lack of broader substrate scope; (v) lack of higher enantio- and diastereoselectivity and (vi) use of protection and

deprotection of various functional groups involved in the synthesis limiting the overall yield of the process, particularly unsuitable for atom economic synthesis. In this regard, a simple metal-free procedure to obtain chiral 3-amino-1,2-alkane diol derivatives in high enantio- and diastereoselectivity is highly desirable. In this section, a highly stereoselective, one-pot procedure for obtaining chiral 3-amino-1,2-alkane diols using proline-catalyzed α -aminooxylation of β -aminoaldehydes is described. Since the method involves organocatalysis, especially proline-catalysed (i) Mannich reaction of acetaldehyde¹⁸ and (ii) α -aminooxylation of aldehydes¹⁹ for introducing stereogenicity into the prochiral molecule, a brief account of each of them is described below.

3.1.3.2 Organocatalysis

The broad utility of synthetic chiral molecules as single-enantiomer pharmaceuticals, in electronic and optical devices, as components in polymers with novel properties, and as probes of biological function, has made asymmetric catalysis a prominent area of investigation. Until a few years ago, it was generally established that transition metal complexes and enzymes were the two main classes of very efficient asymmetric catalysts. Synthetic chemists have hardly used small organic molecules as catalysts throughout the last century, even though some of the very first asymmetric catalysts were purely organic molecules. Simple organic molecules can be highly effective enantioselective catalysts for a variety of important organic transformations.²⁰ This rediscovery has initiated an explosive growth of research activities in organocatalysis both in industry and in academia. Organocatalysis is the catalysis of chemical transformations using a purely organic molecule, which is composed of mainly carbon, hydrogen, nitrogen, sulfur and phosphorus, and does not contain any metals. The advantages of organocatalysts include their lack of sensitivity to moisture and

oxygen, their ready availability, low cost, and low toxicity, which confers a huge direct benefit in the production of pharmaceutical intermediates when compared with transition metal catalysts. Organic molecules not only have ease of manipulation and a “green” advantage but also can be very efficient catalysts. Asymmetric organocatalysis may begin to catch up with the spectacular advancements of enantioselective transition metal catalysis. In particular, L-proline (**48**) has been defined as a “universal catalyst” because of its high utility in a variety of asymmetric organic transformations. Proline is the only natural amino acid with a secondary amine functionality, which raises the pKa value and better nucleophilicity as compared to other amino acids. It can act as a nucleophile to carbonyl groups (iminium intermediate) or Michael acceptors (enamines).

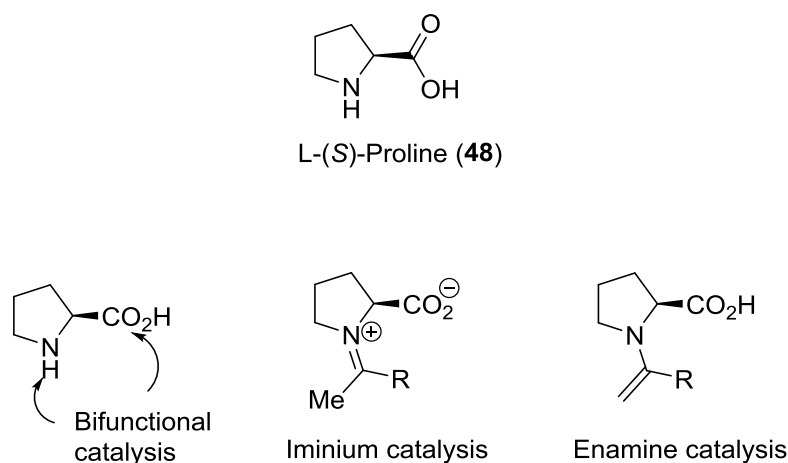


Fig. 2: Modes of proline catalysis

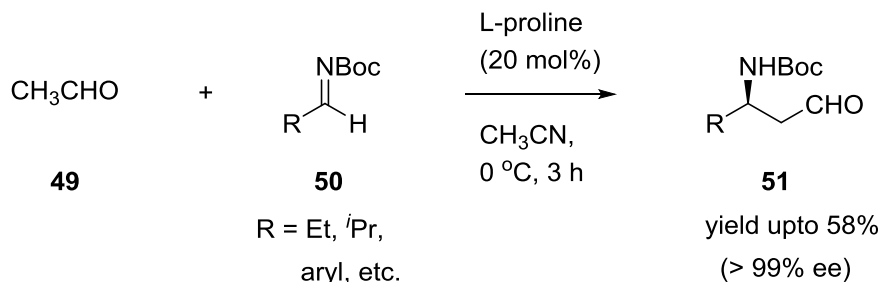
It can be regarded as a bifunctional catalyst as the secondary amine acts as Lewis base and the acid group acts as Bronsted acid (**Fig. 2**). The high stereoselectivity in the proline-catalyzed reactions is possibly due to its formation of organized transition states with many hydrogen bonding frameworks. Proline is not the only molecule to promote catalysis, but it still seems to be one of the best in the diversity of

transformations. It is known to catalyze aldol,²¹ Diels-Alder,²² Michael addition,²³ Mannich¹⁸ and α -functionalization²⁴ among many other organic transformations.²⁵

3.1.3.3 Proline-catalyzed Mannich reaction of acetaldehyde

Mannich reaction is enormously useful for the construction of nitrogenous molecules.²⁶ The increasing popularity of the Mannich reaction has been fueled by the ubiquitous nature of nitrogen in drugs and natural products as well as by the potential of this multi-component reaction to generate diversity. Only a handful of catalytic asymmetric Mannich reactions have been reported.²⁷ The proline-catalysed Mannich reaction has evolved into a broadly useful transformation that has been applied to the synthesis of natural products, pharmaceuticals, and several classes of chiral amino acids.²⁸ Very recently, N-Boc-imines **50** have been introduced to the proline-catalysed Mannich reaction,²⁹ significantly widening the already large substrate scope and utility of this process. However, acetaldehyde, which would be a particularly useful nucleophile in this reaction, was not used under proline-catalysed Mannich reactions conditions till recent years due to several problems associated with the potential use of acetaldehyde, such as: (i) acetaldehyde rapidly reacting with itself *via* aldol condensation, forming coloured oligomers and polymers if treated with proline and (ii) hypothetical acetaldehyde Mannich products, themselves α -unbranched aldehydes, may undergo further reaction with an additional imine equivalent or eliminate to form the corresponding unsaturated aldehydes. It was recently found that these potential side reactions can be suppressed if a higher excess of acetaldehyde (5-10 equivalents) is used.¹⁸ Thus, when N-Boc-imines **50** were treated with acetaldehyde **49** in the presence of L-proline (20 mol%) in CH₃CN at 0 °C the desired β -aminoaldehydes **51** were obtained in extremely high enantioselectivities (> 99%) and reasonable yields (**Scheme 13**). The β -amino aldehyde products **51** formed with very high

enantioselectivity are highly attractive precursors of chiral β -amino acids, which play a key role in investigations of β -peptides and pharmaceuticals.³⁰

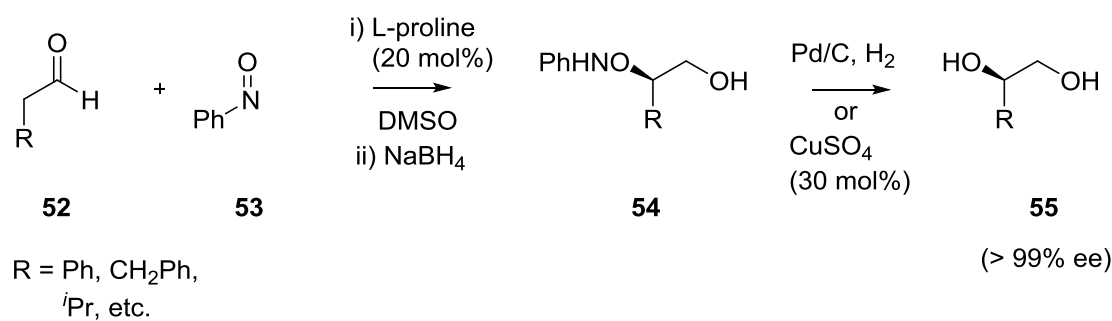


Scheme 13: Mannich reaction of acetaldehyde

3.1.3.4 Proline-catalyzed α -Aminooxylation of Aldehydes

Optically active α -hydroxy aldehydes and ketones are important intermediates in organic synthesis as they are the direct precursors to 1,2-diols. Because of this utility many methods have been developed for their preparation. The more prominent, well-established methods of enantioselective α -oxygenations include the use of Davis oxaziridine,^{31a} Sharpless dihydroxylation of enol ethers,^{31b} manganese-salen epoxidation of enol ethers,^{31c} and Shi' epoxidation of enol ethers.^{31d} It is only rather recently that direct catalytic, asymmetric variants have been reported.³² Most of these methods, however, require multiple manipulations and there is no direct method, nor catalytic asymmetric version for their synthesis from the corresponding aldehyde.

Proline is equally efficient for α -functionalization²⁴ of aldehydes and ketones. When an aldehyde **52** without substitution at α -position was reacted with nitrosobenzene **53** in presence of L-proline in DMSO at ambient temperature, aminooxylation of the aldehyde takes place at α -position. The aminooxyl moiety **54** undergoes hydrogenolysis with Pd/C, H₂ or CuSO₄ to give the corresponding diols **55** in very high enantioselectivities (**Scheme 14**).



Scheme 14: α -aminoxylation of aldehydes

The mechanism of the α -aminoxylation reaction is shown in **Fig. 3**. The observed enantioselectivity of the catalytic α -aminoxylation of aldehydes can be rationalized by invoking an enamine mechanism operating through a chair like transition state, wherein the *Si* face of an (*E*)-enamine formed from the aldehyde and L-proline approaches the less hindered oxygen atom of nitrosobenzene to provide a chiral α -aminoxyaldehyde with *R* configuration. Since proline is commercially available in both enantiopure forms, a one-pot sequential catalytic α -aminoxylation of aldehydes followed by *in situ* reduction with NaBH₄ affords *R*- or *S*- configured 1,2-diol units (the secondary alcohol “protected” by an *O*-amino group) with excellent enantioselectivities and in good yields.

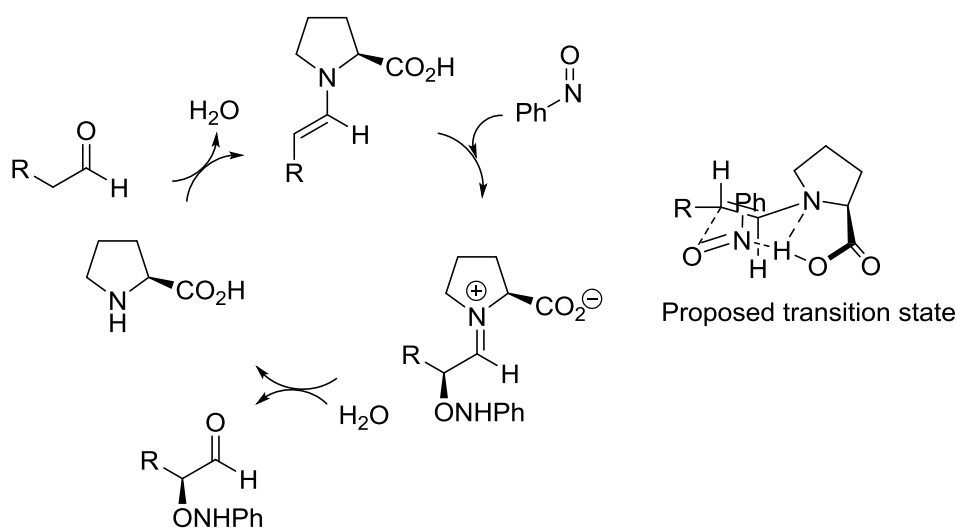
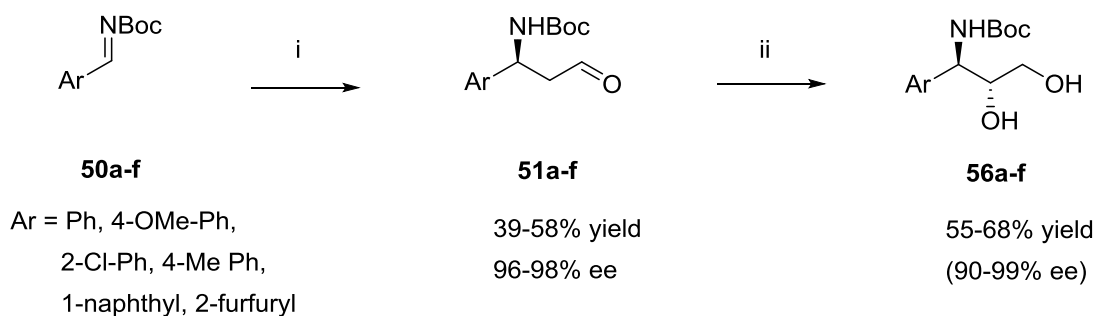


Fig. 3: Proposed mechanism of the α -aminoxylation reaction

3.1.4 Results and Discussion

Considering the high biological importance of chiral 3-amino-1,2-diols, which are also key intermediates in many drugs, we became interested in synthesizing them in one-pot reaction. Based on the literature knowledge of proline-catalyzed Mannich reaction (**Scheme 13**) and α -functionalization of aldehydes (**Scheme 14**), we hypothesized that one pot-synthesis of 3-amino-1,2-alkanediol **56** should be possible through sequential Mannich reaction of Boc-protected benzaldimine **50** with acetaldehyde **49** via List's protocol using L-proline providing β -aminoaldehyde **51** *in situ* followed by addition of PhNO and reduction with NaBH₄ in the same pot. Accordingly, we performed several experiments to identify a suitable reaction condition for this sequential α -aminoxylation reaction in one-pot, such as the variation of solvents (CH₃CN, DMSO), temperature, oxygen source (PhNO, benzoyl peroxide) and so on. Unfortunately, we ended up with complex reaction mixtures, often achieving a maximum yield of only 12% of the required product. We then reasoned that low yields may be due to unwanted side reaction of PhNO with the undesired products formed in the Mannich reaction of the first step. To overcome this, we decided to separate pure β -aminoaldehydes **51** after Mannich reaction and then subject it to α -aminoxylation process subsequently (**Scheme 15**).



Scheme 15: (i) CH₃CHO, L-proline (20 mol%), CH₃CN, 0 °C, 3 h; (ii) (a) PhNO (0.8 equiv), L-proline (20 mol%), CH₃CN, -10 °C, 20 h then NaBH₄ CH₃OH, 10 min; (b) Cu(OAc)₂·H₂O (15 mol%), CH₃OH, 25 °C, 10 h.

Thus, pure β -aminoaldehydes **51a-f**, the starting materials for α -aminoxylation were efficiently prepared from the corresponding Boc-protected arylaldimines **50a-f** following literature protocol (L-proline, CH_3CN , 0 °C). The formation of β -aminoaldehydes **51a-f** was confirmed by ^1H and ^{13}C NMR spectroscopy as follows.

Example 1: The ^1H NMR spectrum of β -aminoaldehyde **51e** showed a typical triplet at δ 9.81 (t, $J = 2.2$ Hz, 1H) and a doublet at δ 3.09 (d, $J = 6.5$ Hz, 2H) corresponding to aldehydic and homobenzylic methylene protons respectively. Its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 195.4 and 200.2 due to carbamate and aldehydic carbonyl carbons respectively (**Fig. 4**).

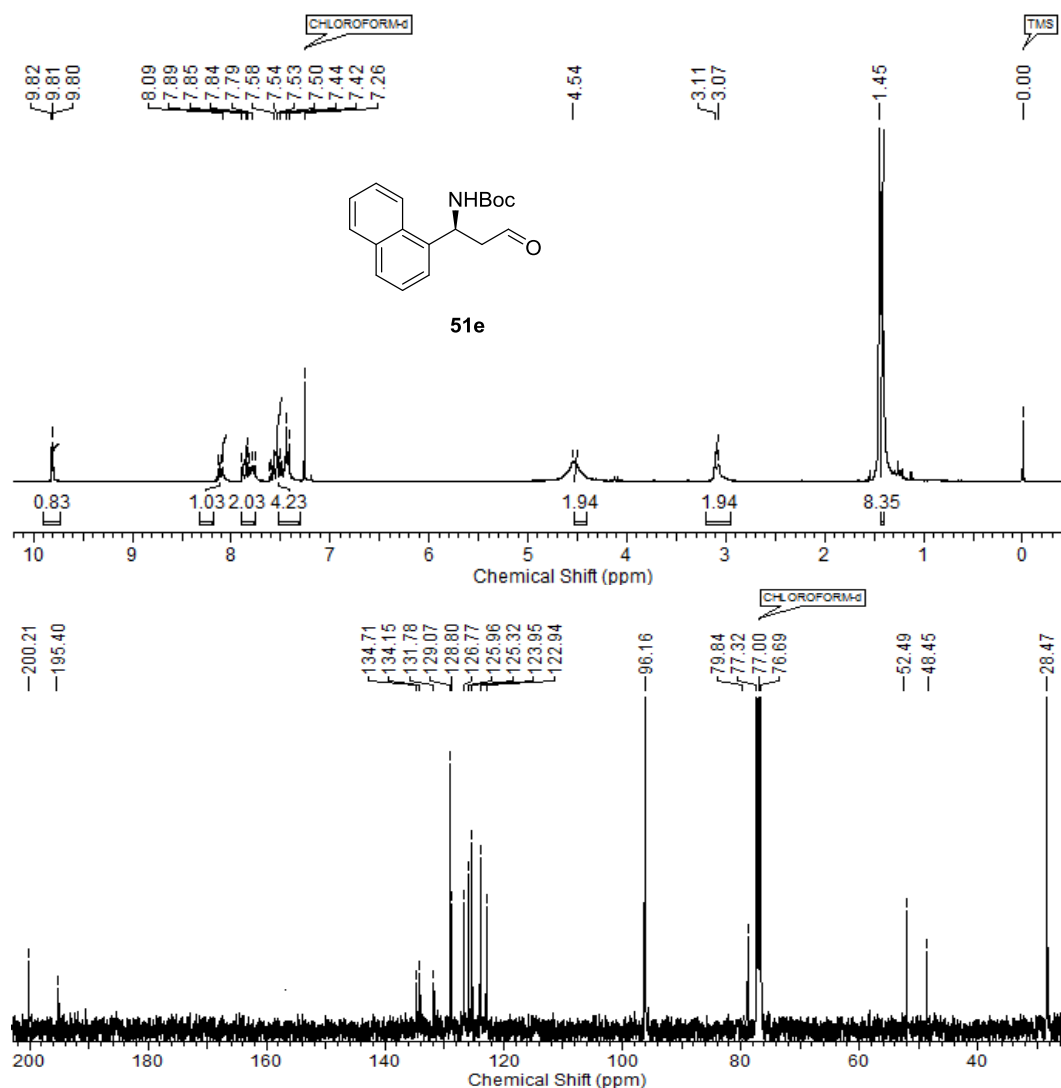


Fig. 4: ^1H and ^{13}C NMR spectra of β -aminoaldehyde **51e**

Example 2: The ^1H NMR spectrum of β -aminoaldehyde **51a** showed typical proton signals at δ 9.73 (t, $J = 1.7$ Hz, 1H) and δ 2.83-2.96 (m, 2H) corresponding to aldehydic and homobenzylic methylene protons respectively. Its ^{13}C NMR showed two characteristic carbon signals at δ 199.8 and 193.3 corresponding to carbamate and aldehyde carbonyl carbons respectively. Its IR spectrum also exhibited a characteristic strong $\text{C}=\text{O}$ absorption band at 1692 cm^{-1} due to carbamate and aldehydic carbonyl group (**Fig. 5**).

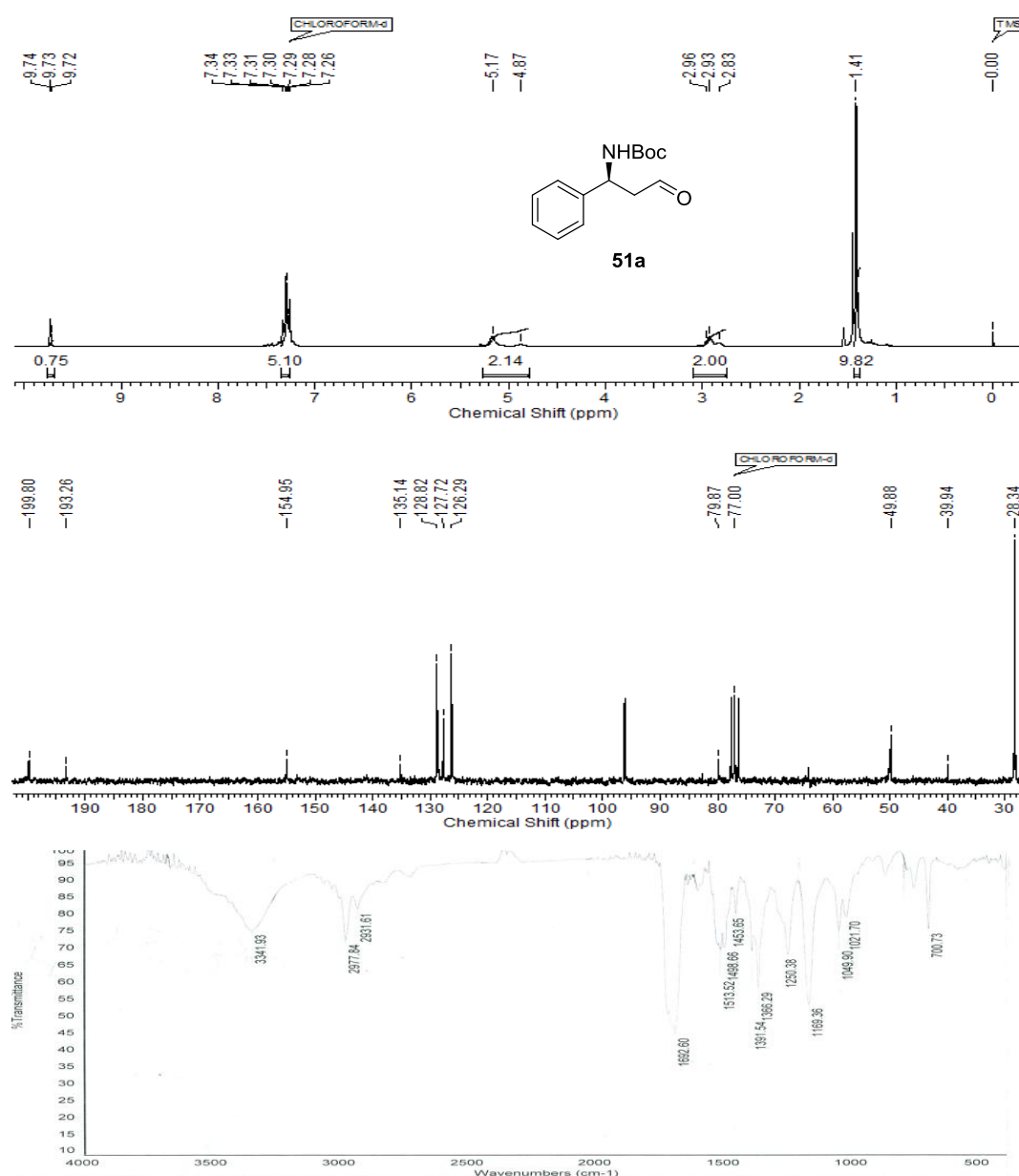


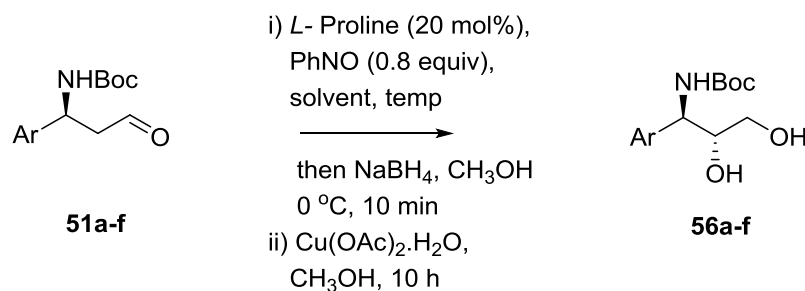
Fig. 5: ^1H , ^{13}C NMR and IR spectra of β -aminoaldehyde **51a**

We observed a drastic improvement in the yield of **56a**, when pure β -aminoaldehyde **51a** was subjected to α -aminooxylation separately. The reaction proceeded through α -aminoxy aldehyde, which was *in situ* reduced with NaBH₄. This was followed by subsequent reduction of the crude aminoxy product with Cu(OAc)₂·H₂O providing a single diastereomer of *anti*-3-amino-1,2-diol **56a** in 50% yield and 98% ee (**Table 1, Entry 1**).

Encouraged by the result, we became interested to improve the yield further by optimizing the reaction conditions such as variation of solvents and temperatures for the α -aminooxylation step using β -aminoaldehyde **51a** as the model substrate. Among the solvents screened, CH₃CN was found to give a maximum yield as compared to DMSO, CHCl₃, CH₂Cl₂ and THF. When the temperature was maintained at -10 °C, high yields and high enantiomeric excess were obtained (**Table 1, Entry 2**). However, further increase or decrease in the temperature to either 0 °C or -20 °C had a deleterious effect on the yield.

To determine the scope and generality of the reaction, a variety of β -amino aldehydes **51a-f** were subjected to α -aminooxylation process. In every case, the reaction proceeded smoothly to give a single diastereomer of *anti*-3-amino-1,2-diols **56a-f**, yields ranging from 55-68% with the intact of excellent enantioselectivity. For instance, substrates having electron-rich (entry 9) or electron-deficient (entry 11) substituent on the aromatic ring including 1-naphthyl ring system (entry 12) and heteroaryl (entry 13) gave the desired *anti*-3-amino-1,2-diols in excellent stereoselectivity.

A major advantage of this strategy is that all the four stereoisomers of 3-amino-1,2-diols can be prepared by choosing a suitable L or D-proline for Mannich and subsequent aminooxylation reactions.

Table 1: L-proline-catalyzed α -aminoxylation of β -aminoaldehydes: studies on optimization and substrate scope

entry	substrates	solvent	temp	time	products	yield	dr	%
	51a-f (Ar)		(°C)	(h)	56a-f	(%) ^a	(%) ^b	ee
1	Ph	CH ₃ CN	-20	22	56a	50	>99	93 ^d
2	Ph	CH₃CN	-10	15	56a	68	>99	93^d
3	Ph	CH ₃ CN	0	15	56a	53	>99	90 ^d
4	Ph	DMSO	25	0.5	56a	55	>99	93 ^d
5	Ph	DMF	25	1	56a	17	>99	nd
6	Ph	CHCl ₃	25	24	56a	30	>99	nd
7	Ph	CH ₂ Cl ₂	25	20	56a	40	>99	91 ^d
8	Ph	THF	25	24	56a	16	>99	nd
9	4-OMe-Ph	CH ₃ CN	-10	15	56b	65	>99	92 ^d
10	4-Me-Ph	CH ₃ CN	-10	15	56c	63	>99	95 ^c
11	2-Cl-Ph	CH ₃ CN	-10	15	56d	55	>99	>99 ^c
12	1-naphthyl	CH ₃ CN	-10	15	56e	62	>99	92 ^c
13	2-furfuryl	CH ₃ CN	-10	15	56f	58	>99	90 ^c

^aIsolated yield after column chromatographic purification. ^bdetermined based on ¹H NMR spectrum.

^cdetermined from HPLC analysis. ^dby comparing with specific rotation reported in the literature.

The formation of all *anti*-3-amino-1,2-diols **56a-f** was established unambiguously from their corresponding ^1H & ^{13}C NMR, IR and HRMS spectral data. Their optical purity was established from their chiral HPLC analyses.

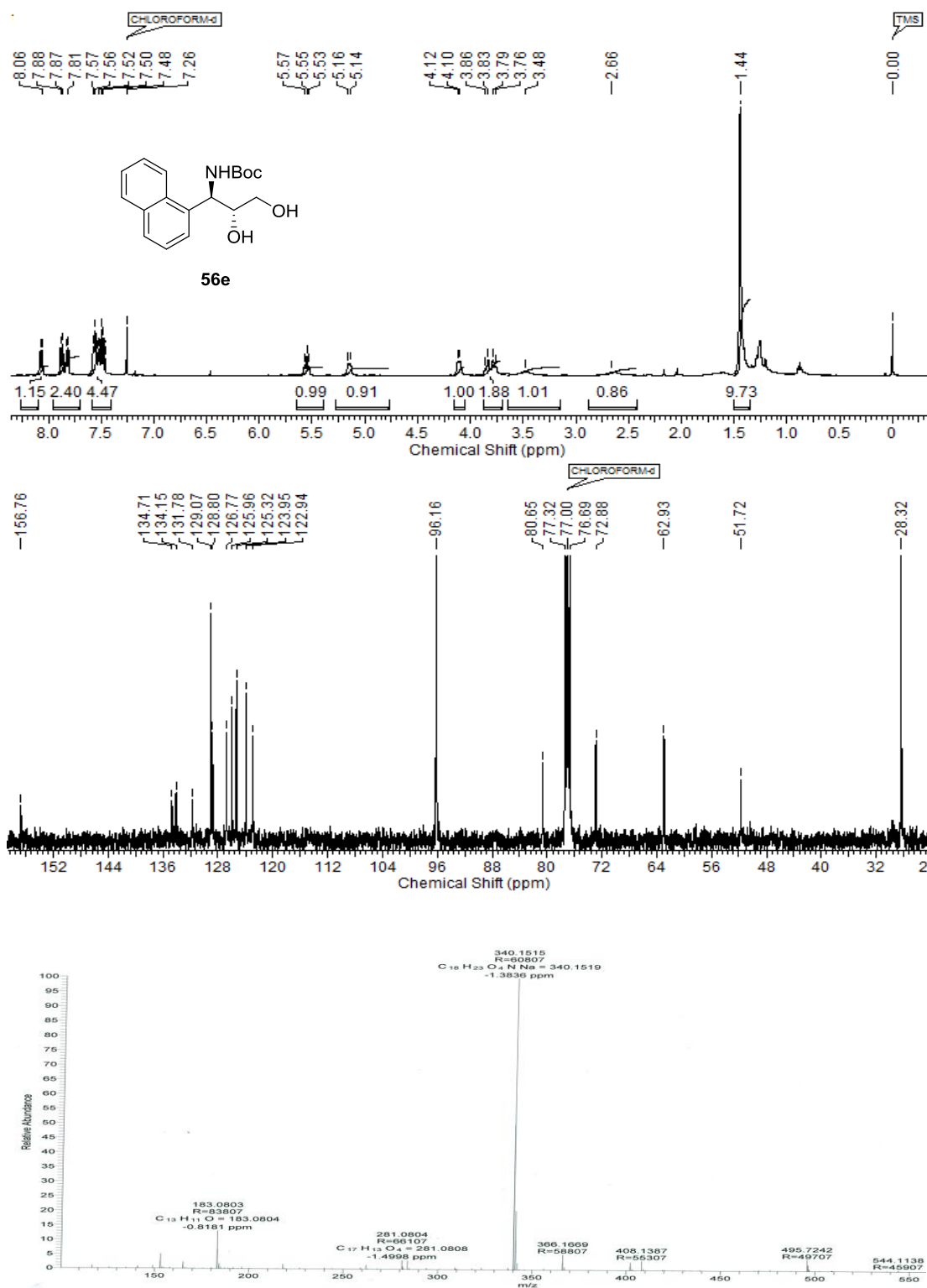
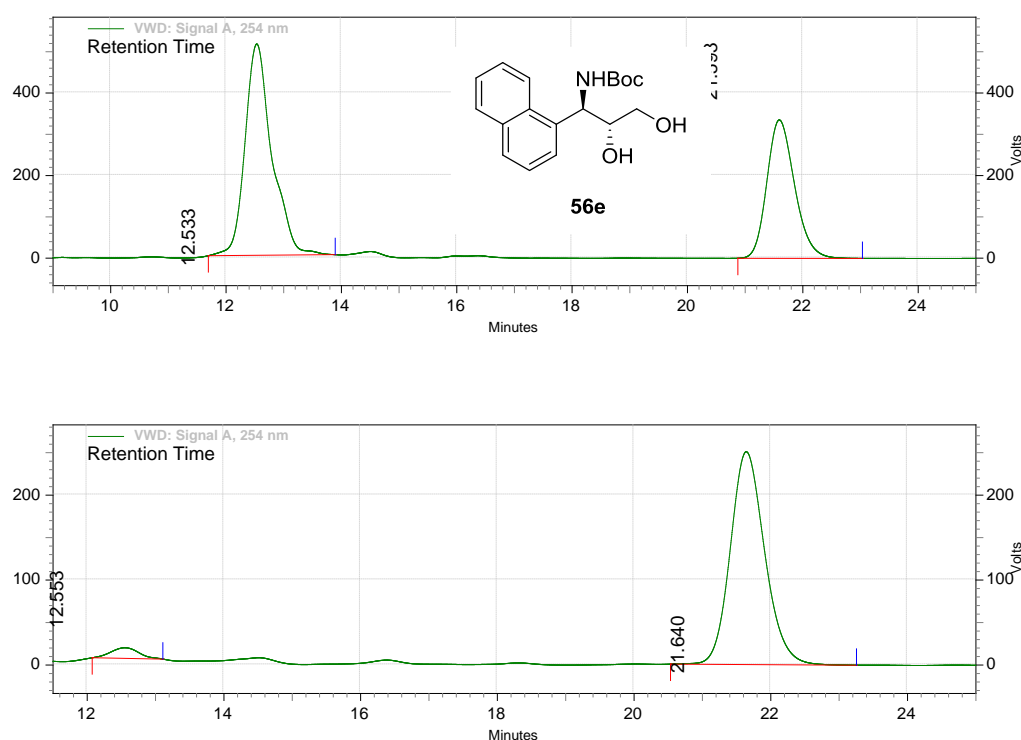


Fig. 6: ^1H , ^{13}C NMR and mass spectra of *anti*-3-amino-1,2-diol **56e**

Example 1: The ^1H NMR spectrum of *anti*-3-amino-1,2-diol **56e** showed two typical signals at δ 2.66 (br s, 1H) and 3.48 (br s, 1H) corresponding to hydroxyl protons while the other signal at δ 3.81 (dd, $J = 11.8, 28.9$ Hz, 2H) for methylene ($-\text{CH}_2\text{-OH}$) protons; Its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 62.9 and 72.9 attributed to methylene and methine carbons attached to hydroxyl groups respectively. The mass spectrum also confirmed the formation of *anti*-3-amino-1,2-diols **56e** (Fig. 6).



Retention Time	Area	Area %	Height	Height %
12.553	5989186	3.92	209816	4.75
21.640	146796171	96.08	4207363	95.25

Fig. 7: HPLC chromatogram of *anti*-3-amino-1,2-diol **56e**

The optical purity of *anti*-3-amino-1,2-diol **56e** was determined to be 92% ee from chiral HPLC analysis (Chiracel AD-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 12.55 min (3.92%) and 21.64 min (96.1%) (Fig.7).

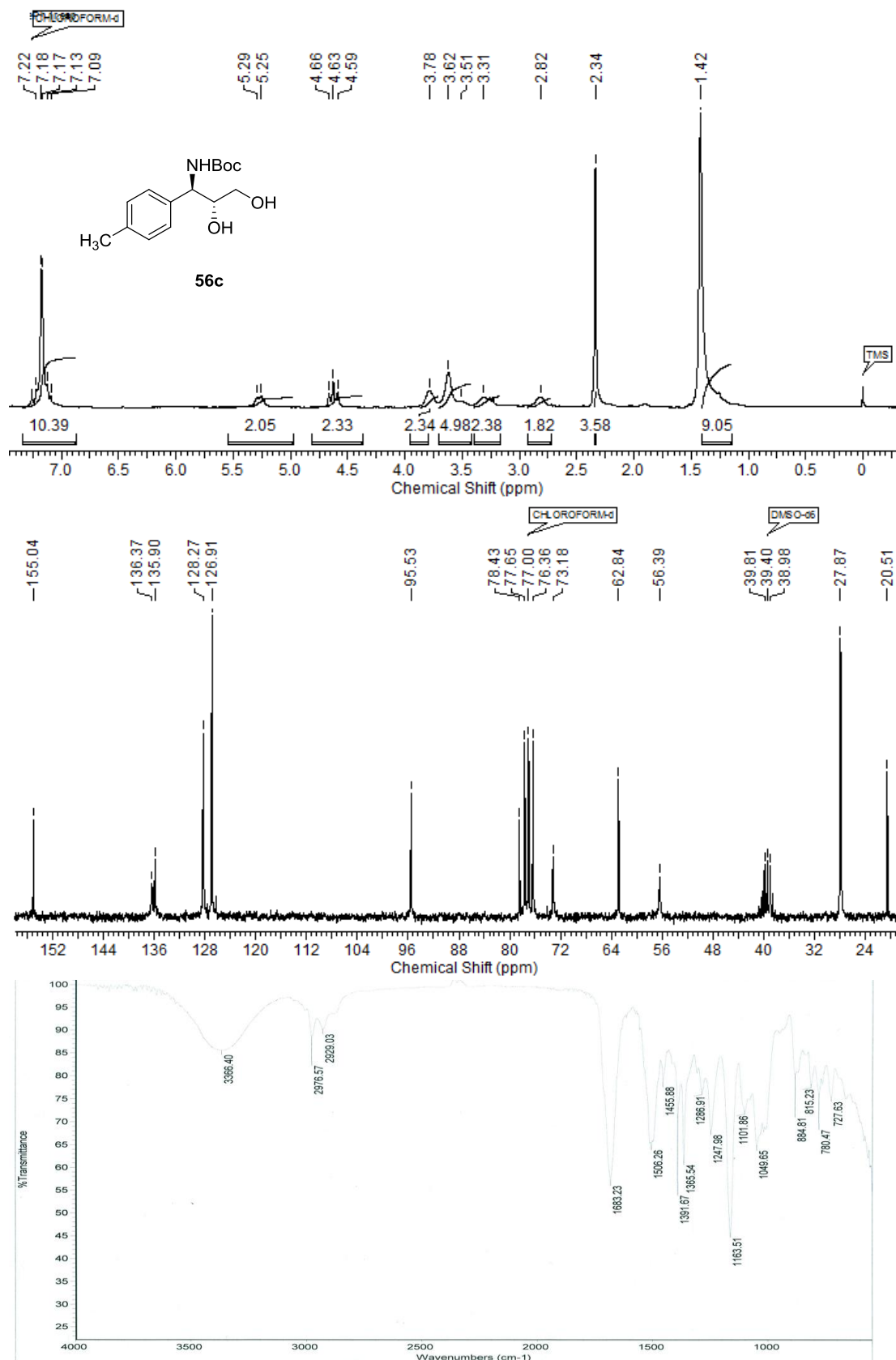


Fig. 8: ^1H , ^{13}C NMR and mass spectra of *anti*-3-amino-1,2-diol **56c**

Example 2: The ^1H NMR spectrum of *anti*-3-amino-1,2-diol **56c** showed two typical signals at δ 2.82 (br s, 1H) and 3.31 (br s, 1H) corresponding to hydroxyl protons while the other signal at δ 2.31 (s, 3H) for methyl protons attached to aromatic ring. Its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 62.8 and 73.2 due to methylene and methine carbons attached to hydroxyl groups respectively. Its IR spectrum displayed a characteristic broad absorption band at 3366 cm^{-1} indicating the presence of OH functional group (**Fig. 8**).

The absolute configuration of the newly generated chiral center was assigned on the basis of the previously established configuration of α -aminoxylation of aldehydes.¹⁹ The *anti*-stereochemistry in *anti*-3-amino-1,2-diol **56c** is, however, unambiguously proven from X-ray crystallographic analysis (**Fig. 9**).

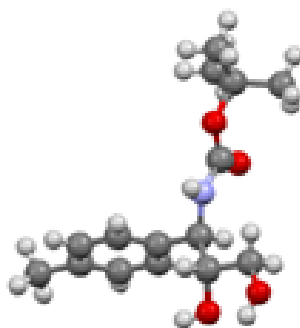


Fig. 9: ORTEP diagram of *anti*-3-amino-1,2-diol **56c**

3.1.5 Conclusion

A highly stereoselective route to 3-amino-1,2-diols using proline as catalyst in a single transformation starting from β -aminoaldehydes has been developed. This reaction is also practical in the sense that (i) all the four stereoisomers of 3-amino-1,2-diols can be prepared by choosing a suitable proline as catalyst for Mannich and subsequent aminooxylation reactions; (ii) products were obtained in high optical purities (single diastereomer with 90-99% ee) and moderate yields; (iii) showed broad

substrate scope and good functional group tolerance. We believe that this strategy will find applications in the field of asymmetric synthesis of bioactive molecules owing to the flexible nature of the synthesis and the ready availability of proline catalysts in both enantiomeric forms.

3.1.6 Experimental Section

General experimental procedure for the preparation of β -aminoaldehydes (**51a-f**)

To a stirred solution of aryl N-Boc-imine **50a-f** (1.4 mmol) and redistilled acetaldehyde **49** (0.39 mL, 7 mmol) in CH₃CN (15 mL) at 0 °C was added L-proline (0.032 g, 20 mol%) and the mixture stirred further at 0 °C for 3 h. After the completion of reaction (monitored by TLC), it was quenched with water and extracted with Et₂O (3 x 50 mL). The combined organic layers were washed with brine, dried over anhyd. Na₂SO₄, filtered and concentrated under reduced pressure to give the crude aldehyde. Flash column chromatographic purification [silica gel (230-400 mesh) and pet. ether:EtOAc as an eluent] gave β -aminoaldehydes **51a-f**.

(*S*)-*tert*-Butyl (3-oxo-1-phenylpropyl)carbamate (**51a**)

Yield: 55%; pale yellow solid; **mp:** 91-94 °C, (lit.³³ **mp:** 92-93.5 °C); $[\alpha]_{25}^D$ -30.10 (*c* 1.15, CHCl₃); lit.³³ $[\alpha]_{25}^D$ +29.0 (*c* 1.4, CHCl₃) for its antipode; **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 1021, 1049, 1169, 1250, 1369, 1391, 1498, 1513, 1692, 2977, 3341; **¹H NMR** (200 MHz, CDCl₃): δ 1.41 (s, 9H), 2.83-2.96 (m, 2H), 4.87 (br s, 1H), 5.17 (br s, 1H), 7.26-7.34 (m, 5H), 9.73 (t, *J* = 1.7 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.3, 39.9, 49.9, 79.9, 126.3, 127.7, 128.8, 135.2, 155.0, 193.3, 199.8; **Analysis:** C₁₄H₁₉NO₃ requires C, 67.45; H, 7.68; N, 5.62; found: C, 67.32; H, 7.41; N, 5.46%.

(*S*)-*tert*-Butyl (1-(4-methoxyphenyl)-3-oxopropyl)carbamate (**51b**)

Yield: 56%; pale yellow liquid; $[\alpha]_{25}^D$ -32.90 (*c* 1.1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 690, 1021, 1050, 1170, 1246, 1378, 1387, 1463, 1520, 1689, 2850, 2924, 2978, 3346;

¹H NMR (200 MHz, CDCl₃): δ 1.41 (s, 9H), 2.80-2.94 (m, 2H), 3.78 (s, 3H), 4.90 (br s, 1H), 5.13 (br s, 1H), 6.85 (d, *J* = 8.6 Hz, 2H), 7.21 (d, *J* = 8.6 Hz, 2H), 9.72 (t, *J* = 1.6 Hz, 1H); **¹³C NMR** (100 MHz, CDCl₃): δ 28.3, 39.8, 49.8, 55.1, 79.6, 114.1, 127.5, 134.9, 154.9, 159.0, 193.4, 200.0; **Analysis**: C₁₅H₂₁NO₄ requires C, 64.50; H, 7.58; N, 5.01; found: C, 64.32; H, 7.38; N, 5.06%.

(*S*)-tert-Butyl (3-oxo-1-(*p*-tolyl)propyl)carbamate (51c)

Yield: 58%; pale yellow liquid; [α]₂₅^D -36.28 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 1026, 1054, 1172, 1242, 1384, 1467, 1518, 1692, 2900, 2982, 3341; **¹H NMR** (200 MHz, CDCl₃): δ 1.41 (s, 9H), 2.33 (s, 3H), 2.83-2.96 (m, 2H), 5.15 (br s, 1H), 5.17 (br s, 1H), 7.13-7.26 (m, 4H), 9.73 (t, *J* = 1.7 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 21.3, 28.4, 50.8, 53.1, 79.6, 125.1, 128.8, 136.9, 140.5, 193.4, 202.2; **Analysis**: C₁₅H₂₁NO₃ requires C, 68.42; H, 8.04; N, 5.32; found: C, 68.26; H, 8.01; N, 5.26%.

(*S*)-tert-Butyl (1-(2-chlorophenyl)-3-oxopropyl)carbamate (51d)

Yield: 49%; yellow liquid; [α]₂₅^D -12.32 (*c* 1.6, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 696, 1032, 1061, 1168, 1256, 1376, 1459, 1522, 1698, 2851, 2868, 3347; **¹H NMR** (200 MHz, CDCl₃): δ 1.45 (s, 9H), 2.86-2.98 (m, 2H), 4.54 (br s, 2H), 5.26 (br s, 1H), 7.21-7.28 (m, 2H), 7.32-7.40 (m, 2H), 9.73 (t, *J* = 1.5 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.4, 46.4, 51.5, 79.4, 126.6, 128.3, 132.2, 136.5, 193.4, 199.7; **Analysis**: C₁₄H₁₈ClNO₃ requires C, 59.26; H, 6.39; N, 4.94; found: C, 59.12; H, 6.19; N, 4.82%.

(*S*)-tert-Butyl (1-(naphthalen-1-yl)-3-oxopropyl)carbamate (51e)

Yield: 44%; yellow liquid; [α]₂₅^D -18.71 (*c* 1.9, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 700, 1024, 1053, 1174, 1259, 1398, 1481, 1501, 1626, 2979, 3412; **¹H NMR** (200 MHz, CDCl₃): δ 1.45 (s, 9H), 3.09 (d, *J* = 6.5 Hz, 2H), 4.54 (br s, 2H), 7.42-7.61 (m, 4H), 7.76-7.89 (m, 2H), 8.11 (d, *J* = 8.1 Hz, 1H), 9.81 (t, *J* = 2.2 Hz, 1H); **¹³C NMR** (50

MHz, CDCl₃): δ 28.4, 48.4, 52.5, 79.8, 122.9, 123.9, 125.3, 125.9, 126.7, 128.8, 129.1, 131.8, 134.2, 134.7, 195.4, 200.2; **Analysis:** C₁₈H₂₁NO₃ requires C, 72.22; H, 7.07; N, 4.68; found: C, 72.12; H, 7.03; N, 4.47%.

(S)-tert-Butyl (1-(furan-2-yl)-3-oxopropyl)carbamate (51f)

Yield: 39%; yellow liquid; $[\alpha]_{25}^D$ -18.71 (*c* 1.9, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 690, 1084, 1062, 1174, 1259, 1391, 1467, 1501, 1683, 2824, 2841, 3421; **¹H NMR** (200 MHz, CDCl₃): δ 1.44 (s, 9H), 2.91-3.02 (m, 2H), 5.11 (br s, 1H), 5.27 (br s, 1H), 6.30-6.34 (m, 2H), 7.36 (br s, 2H), 9.77 (t, *J* = 2.1 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.4, 50.1, 51.5, 79.4, 108.3, 110.6, 142.3, 151.8, 193.6, 200.7; **Analysis:** C₁₂H₁₄NO₄ requires C, 60.24; H, 7.16; N, 5.85; found: C, 60.12; H, 7.03; N, 5.76%.

General experimental procedure for the preparation of 3-amino-1,2-alkane diols (56a-f)

To a stirred precooled (-10 °C) acetonitrile (25 mL) solution of β -aminoaldehydes **51a-f** (17 mmol) and nitrosobenzene (1.45 g, 13.6 mmol) was added L-proline (0.039 g, 20 mol%). The reaction mixture was allowed to stir at the same temperature for 20 h followed by the addition of MeOH (10 mL) and NaBH₄ (0.97 g, 25 mmol) to the reaction mixture, which was stirred for further 10 min. After addition of phosphate buffer, the resulting mixture was extracted with EtOAc (3 \times 30 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude aminoxy alcohol, which was directly taken up for the next step without purification. To a MeOH (25 mL) solution of the above crude aminoxyalcohol was added Cu(OAc)₂.H₂O (0.501 g, 2.6 mmol) at 25 °C and the reaction mixture was allowed to stir for 10 h at that temperature. After addition of phosphate buffer, the resulting mixture was extracted with CHCl₃ (3 \times 30 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then

purified by column chromatography over silica gel using pet. ether:EtOAc to give 3-amino-1,2-alkane diols **56a-f**.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-phenyl-1,2-propanediol (56a)

Yield: 68%; colourless solid; **mp:** 106-109 °C, (lit.³⁴ **mp:** 107-108 °C); $[\alpha]_{25}^D$ -40.96 (*c* 1.32, CHCl₃); lit.³⁴ $[\alpha]_{25}^D$ +42.90 (*c* 1.4, CHCl₃) for its antipode; **IR** (CHCl₃, cm⁻¹): ν_{\max} 790, 1025, 1056, 1124, 1161, 1369, 1400, 1495, 1696, 2930, 2986, 3370; **¹H NMR** (400 MHz, CDCl₃): δ 1.43 (s, 9H), 2.81 (br s, 1H), 3.28 (br s, 1H), 3.64 (br s, 2H), 3.82 (br s, 1H), 4.66-4.71 (m, 1H), 5.29 (br s, 1H), 7.28-7.38 (m, 5H); **¹³C NMR** (100 MHz, CDCl₃): δ 28.3, 56.7, 63.2, 73.9, 80.1, 127.4, 127.7, 128.6, 139.0, 156.1; **HRMS** (ESI) *m/z* calcd for C₁₄H₂₁NO₄ [M + Na]⁺: 290.1368, found: 290.1377; **Analysis:** C₁₄H₂₁NO₄ requires C, 62.90; H, 7.92; N, 5.24; found: C, 62.78; H, 7.81; N, 5.12%.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-(*p*-methoxyphenyl)-1,2-propanediol (56b)

Yield: 65%; colorless solid; **mp:** 114-116 °C, (lit.³⁵ **mp:** 116-118 °C); $[\alpha]_{25}^D$ -49.43 (*c* 1.0, CHCl₃); lit.³⁵ $[\alpha]_{25}^D$ -50.2 (*c* 0.5, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 669, 757, 831, 927, 1035, 1167, 1216, 1368, 1585, 1612, 1701, 2400, 2839, 2981, 3019, 3438, 3682; **¹H NMR** (200 MHz, CDCl₃): δ 1.34 (s, 9H), 3.11 (br s, 2H), 3.54 (br s, 2H), 3.71 (br s, 4H), 4.55 (br s, 1H), 5.29 (br s, 1H), 6.78 (d, *J* = 6.71 Hz, 2H), 7.14 (d, *J* = 6.71 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.3, 55.2, 56.1, 63.2, 74.1, 80.1, 114.7, 128.5, 131.1, 156.2, 159.2; **HRMS** (ESI) *m/z* calcd for C₁₅H₂₃NO₅ [M + Na]⁺: 320.1473, found: 320.1469; **Analysis:** C₁₅H₂₃NO₅ requires C, 60.59; H, 7.80; N, 4.71; found: C, 60.29; H, 7.62; N, 4.69%.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-(*p*-tolyl)-1,2-propanediol (56c)

Yield: 63%; colorless solid recrystallized from CHCl₃; **mp:** 126-129 °C; $[\alpha]_{25}^D$ -57.87 (*c* 2.7, CHCl₃); 95% ee from chiral HPLC analysis (Chiracel AD-H, *n*-hexane/*i*PrOH,

90:10, 0.5 mL/min) retention time 21.6 min (97%) and 27.1 min (2%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 727, 780, 815, 884, 1049, 1101, 1163, 1247, 1287, 1365, 1391, 1506, 1683, 2929, 2976, 3366; **^1H NMR** (200 MHz, CDCl_3): δ 1.42 (s, 9H), 2.31(s, 3H), 2.82 (br s, 1H), 3.31 (br s, 1H), 3.51-3.62 (m, 2H), 3.78 (br s, 1H), 4.59-4.66 (m, 1H), 5.27 (d, $J = 6.7$ Hz, 1H), 7.09-7.22 (m, 4H); **^{13}C NMR** (50 MHz, CDCl_3 +DMSO- d_6): δ 20.5, 27.9, 56.4, 62.8, 73.2, 78.4, 127.0, 128.3, 135.9, 136.4, 155.0; **HRMS** (ESI) m/z calcd for $\text{C}_{15}\text{H}_{23}\text{NO}_4$ [$\text{M} + \text{Na}$] $^+$: 304.1519, found: 304.1514; **Analysis**: $\text{C}_{15}\text{H}_{23}\text{NO}_4$ requires C, 64.04; H, 8.24; N, 4.98; found: C, 63.91; H, 8.12; N, 4.93%.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-(o-chlorophenyl)-1,2-propanediol (56d)

Yield: 55%; colorless gum; $[\alpha]_{25}^{\text{D}} -7.50$ (c 0.32, CHCl_3); 99% ee from chiral HPLC analysis (Chiracel AS-H, *n*-hexane/*i*PrOH, 95:05, 0.5 mL/min) retention time 25.9 min (0.5%) and 28.8 min (99.5%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 702, 704, 1036, 1164, 1264, 1367, 1393, 1498, 1694, 2928, 3420; **^1H NMR** (400 MHz, CDCl_3): δ 1.43 (s, 9H), 2.97 (br s, 1H), 3.39 (br s, 1H), 3.64-3.79 (m, 2H), 3.98-3.99 (m, 1H), 5.13-5.17 (m, 1H), 5.56 (br s, 1H), 7.20-7.29 (m, 2H), 7.34-7.42 (m, 2H); **^{13}C NMR** (100 MHz, CDCl_3): δ 28.3, 54.1, 62.8, 72.2, 80.4, 127.1, 128.7, 129.0, 130.1, 133.8, 136.6, 156.3; **HRMS** (ESI) m/z calcd for $\text{C}_{14}\text{H}_{20}\text{ClNO}_4$ [$\text{M} + \text{Na}$] $^+$: 324.0973, found: 324.0970; **Analysis**: $\text{C}_{14}\text{H}_{20}\text{ClNO}_4$ requires C, 55.72; H, 6.68; N, 4.64; found: C, 55.56; H, 6.48; N, 4.47%.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-(1-naphthyl)-1,2-propanediol (56e)

Yield: 62%; colorless gum; $[\alpha]_{25}^{\text{D}} -13.00$ (c 0.2, CHCl_3); 92% ee from chiral HPLC analysis (Chiracel AD-H, *n*-hexane/*i*PrOH, 90:10, 0.5 mL/min) retention time 12.55 min (3.92%) and 21.64 min (96.1%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 774, 1019, 1038, 1121, 1159, 1351, 1408, 1499, 1687, 2921, 2991, 3382; **^1H NMR** (400 MHz, CDCl_3): δ 1.44 (s, 9H), 2.66 (br s, 1H), 3.48 (br s, 1H), 3.81 (dd, $J = 11.8, 28.9$ Hz, 2H), 4.24 (d, $J =$

8.3 Hz, 1H), 5.15 (d, $J = 8.3$ Hz, 1H), 5.53-5.57 (m, 1H), 7.46-7.57 (m, 4H), 7.82 (d, $J = 8.1$ Hz, 1H), 7.88 (d, $J = 7.8$ Hz, 1H), 8.07 (d, $J = 8.3$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 28.3, 51.7, 62.9, 72.9, 80.6, 122.9, 123.9, 125.3, 125.9, 126.8, 128.8, 129.1, 131.8, 134.2, 134.7, 156.8; **HRMS** (ESI) m/z calcd for $\text{C}_{18}\text{H}_{23}\text{NO}_4$ $[\text{M} + \text{Na}]^+$: 340.1519, found: 340.1515; **Analysis**: $\text{C}_{18}\text{H}_{23}\text{NO}_4$ requires C, 68.12; H, 7.30; N, 4.41; found: C, 67.93; H, 7.12; N, 4.31%.

(2R, 3R)-3-(tert-Butoxycarbonylamino)-3-(furfuryl)-1,2-propanediol (56f)

Yield: 58%; colorless gum; $[\alpha]_{25}^{\text{D}}$ -44.32 (c 0.46, CHCl_3); 90% ee from chiral HPLC analysis (Chiracel AS-H, n -hexane/ i PrOH, 90:10, 0.5 mL/min) retention time 13.68 min (4.98%) and 14.84 min (95.02%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 784, 1089, 1066, 1178, 1263, 1398, 1469, 1508, 1699, 2824, 2841, 3384, 3421; **^1H NMR** (500 MHz, CDCl_3): δ 1.44 (s, 9H), 2.98 (br s, 2H), 3.67 (s, 1H), 3.81 (br s, 1.02), 4.76-4.79 (m, 1H), 5.29 (br s, 1H), 6.32-6.34 (m, 2H), 7.37 (br s, 1H); ^{13}C NMR (125 MHz, CDCl_3): δ 28.3, 50.7, 62.8, 73.1, 80.6, 108.1, 110.5, 142.2, 151.7, 156.2; **HRMS** (ESI) m/z calcd for $\text{C}_{12}\text{H}_{19}\text{NO}_5$ $[\text{M} + \text{Na}]^+$: 280.1169, found: 280.1177; **Analysis**: $\text{C}_{12}\text{H}_{19}\text{NO}_5$ requires C, 56.02; H, 7.44; N, 5.44; found: C, 56.07; H, 7.36; N, 5.32%.

Section II:

Enantioselective Synthesis of Cytokine Modulator (+)-*epi*-Cytosaxone and Formal Synthesis of Antibacterial *N*-Thiolated-2-Oxazolidinone

3.2.1 Introduction

In 1998, Osada had reported the isolation of (4*R*, 5*R*)-5-(hydroxymethyl)-4-(4-methoxyphenyl)-1,3-oxazolidinone [(-)-**57**, generic name cytosaxone],³⁶ which was shown to possess high cytokine modulator activity by acting on the Th2 cells.³⁷ Its *trans*-diastereoisomer namely (+)-*epi*-cytosaxone (**58**) showed highly potential biological activity to allergens. Because of these biological properties, several total syntheses of (+)-*epi*-cytosaxone (**58**) and its epimer have been reported.³⁸ Prompted by the first positive biological results, many researchers have also reported the preparation of *cis*- and *trans*-isocytosaxones which are structural isomers of cytosaxone (**57**) and its *trans*-epimer **58** (Fig 10).³⁹

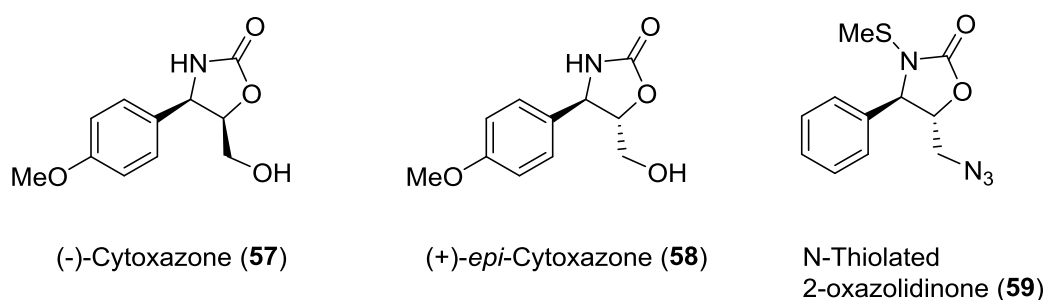


Fig. 10: Structures of cytosaxone epimers (**57** & **58**) and *N*-thiolated 2-oxazolidinone (**59**)

The problem of bacterial drug resistance has reached a crisis level such that successful treatment of antibiotic-resistant infections in hospitals and health care centers can no longer be taken for granted. Infections caused by methicillin-resistant *Staphylococcus*

aureus (MRSA) are becoming particularly difficult to treat with conventional antibiotics such as penicillin, leading to a sharp rise in clinical complications and deaths. The need for new antibacterial agents and protocols for treating MRSA infections is becoming extremely serious. Oxazolidinones have already been recognized for their favorable pharmacological properties and are the only new class of antibacterial drugs introduced into clinical use in the last three decades.⁴⁰ Recent studies have shown that *N*-thiolated-2-oxazolidinone (**59**) possesses antibacterial activity against methicillin-resistant *Staphylococcus aureus* and *Bacillus anthracis*.⁴¹

3.2.2.1 Pharmacology of Cytosaxone Epimers

It is well established that the induction of humoral or cellular response is influenced by the development of distinct subsets of CD4⁺ T cells.⁴² The Th1 cell subset produces predominantly IL-2, GM-CSF, INF- γ , and TNF- β , (type 1 cytokines) and is involved in delayed-type hypersensitivity reactions, whereas the Th2 cell subset secretes IL-4, IL-5, IL-6, IL-10, and IL-13 (type 2 cytokines), which are important factors for β cell growth and differentiation to Ig secretion. The imbalance of cytokine production by CD4⁺ T cells leads to a wide variety of immunological disorders, *i.e.* allergy, progressive lymphoproliferation, and severe immunodeficiency.⁴³ Skin and lung biopsies from allergic patients indicate that the pivotal cells in the allergic site are the Th2 cells.⁴⁴ Treatments effectively suppressing the function or the differentiation of these allergen-specific Th2 cells will most likely provide efficient ways to intervene in Ig-mediated allergic diseases. In the course of screening for chemical immunomodulators that inhibit the type 2 cytokine productions in Th2 cells, it was found that cytosaxone containing a 2-oxazolidinone ring, which is rare in microbial metabolites, as a novel cytokine modulator produced by *Streptomyces sp.*

Cytosaxzones show a cytokine-modulating activity by inhibiting the signaling pathway of Th2 cells, but not Th1 cells.

3.2.2.2 Pharmacology of *N*-Thiolated-2-oxazolidinone

N-Thiolated-2-oxazolidinone (**59**), like their β -lactam counterparts,⁴⁵ react covalently with their biological target through transfer of the organothio side chain as shown in **Fig 11**. Further studies to assess the mode of action of this antiMRSA, antiBacillus compound, and to identify their cellular targets, are of current interest.

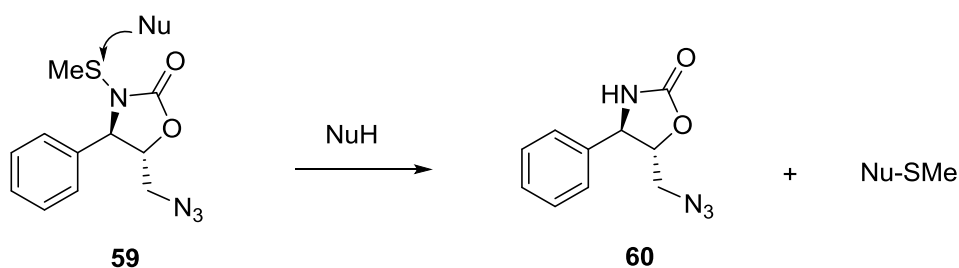


Fig 11: Pharmacology of *N*-thiolated-2-oxazolidinone (**59**)

3.2.3 Review of Literature

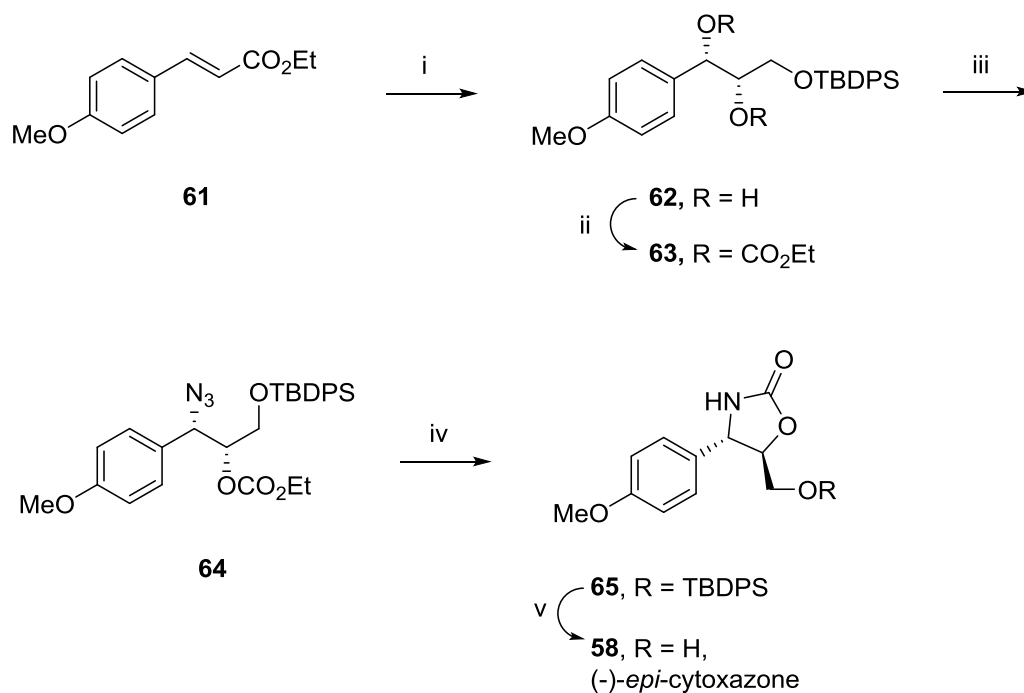
(a) Review of Literature for (+)-*epi*-Cytosaxzone (**58**)

Literature search revealed that there are several reports available for the synthesis of (+)-*epi*-cytosaxzone (**58**), which are described below.

Nakata's approach (1999)⁴⁶

Nakata *et al.* have achieved the synthesis of (-)-*epi*-cytosaxzone (**58**) using Sharpless asymmetric dihydroxylation as the key step. Thus, *p*-methoxycinnamate **61** was subjected to Sharpless asymmetric dihydroxylation to give chiral diol **62** in 93% yield and 99% ee. (4*S*, 5*S*)-Diethylcarbonate **63**, prepared from diol **62**, was treated with TMSN₃ (6 equiv) for achieving stereoselective azidation in the presence of TMSOTf that afforded the desired α -azide **64** (dr = 6:1). The α -azide **64** was then treated under Staudinger reaction condition (PPh₃, THF, H₂O, 50 °C) to give 2-oxazolidinone **65**,

which was converted to 4-*epi*-cytosazone (**58**) in 99% yield on reaction with tetrabutylammonium fluoride (**Scheme 16**).

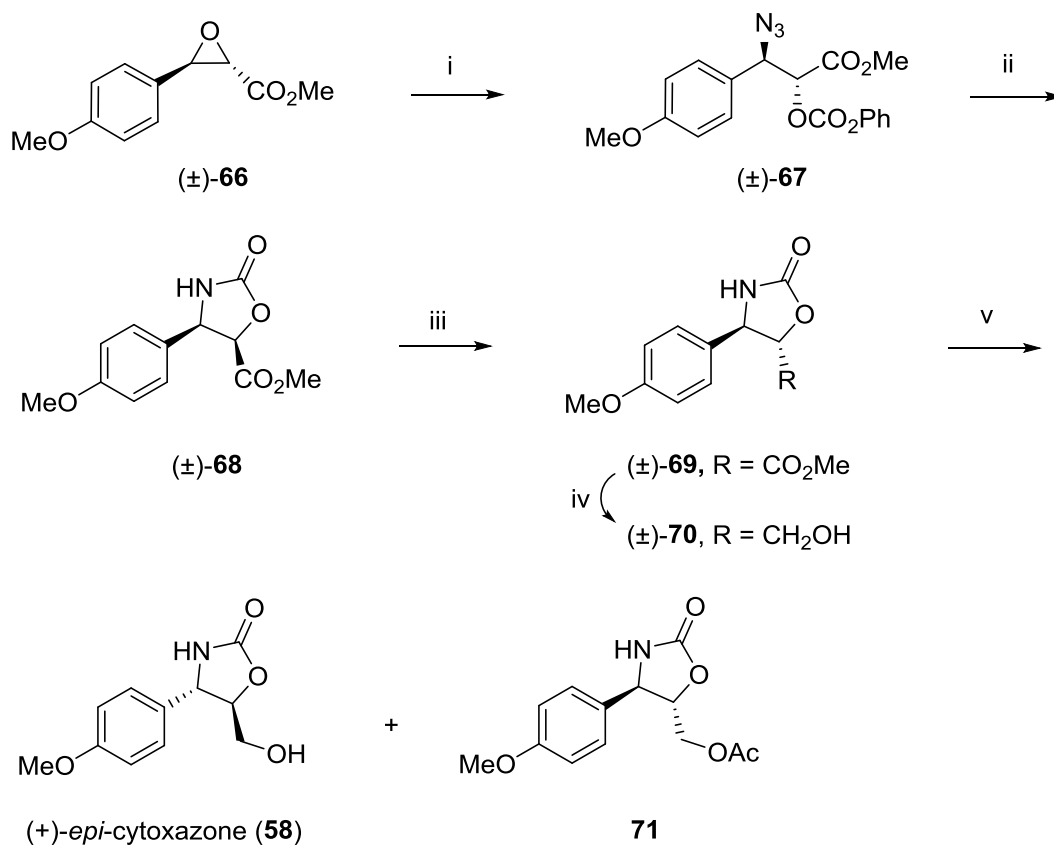


Scheme 16: (i) (a) AD-mix- α , *tert*-BuOH: H₂O (1:1), 25 °C, 93 %, 99% ee; (b) NaBH₄, THF, 0 °C, 66%; (c) TBDPSCl, imid., DMF, 0 °C, 99%; (ii) ClCO₂Et, pyridine, CH₂Cl₂, 0 °C, 92%; (iii) TMSN₃, TMSOTf, CH₃CN, -43 °C, 99%; (iv) PPh₃, THF, H₂O, 50 °C, 100%; (v) ⁿBu₄NF, THF, 0 °C, 99%.

Sunjic's approach (2001)⁴⁷

Sunjic *et al.* have reported the synthesis of (-)-*epi*-cytosazone (**58**) by employing the enzymatic kinetic resolution as the key reaction, starting from the glycidic ester (±)-**66**. Nucleophilic ring opening of epoxide (±)-**66** with NaN₃, followed by protection of the alcohol coupled with intramolecular cyclization gave the ester (±)-**68**. Epimerization at C(5) in oxazolidinone (±)-**68** using potassium hydroxide followed by esterification with methyl iodide gave ester (±)-**69**. Reduction of ester (±)-**69** with NaBH₄ in the presence of CaCl₂ and the subsequent kinetic resolution using lipase

from *Candida antarctica* in SOL-Gel-AK afforded (+)-*epi*-cytosaxone (**58**) in 49% yield and 87.3% ee (**Scheme 17**).

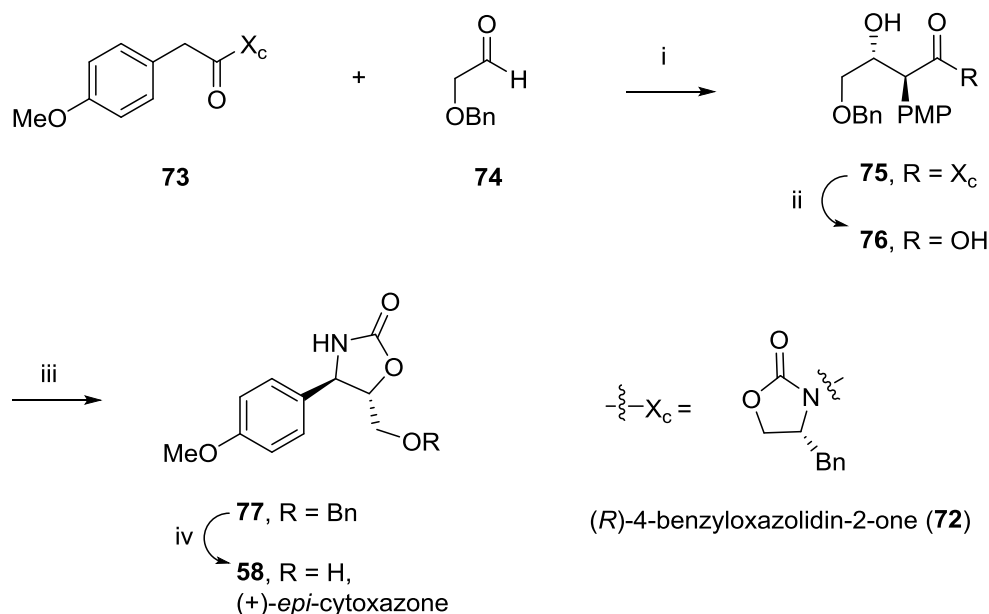


Scheme 17: (i) aq. NaN₃, dioxane, 50 °C, 3 h, 56%; (ii) ClCO₂Ph, CH₂Cl₂, -5 °C, 1 h, 100%; (iii) (a) KOH, EtOH, reflux, 1 h; (b) MeI, K₂CO₃, DMF, 25 °C, 16 h, 46%; (iv) NaBH₄, CaCl₂, absolute EtOH, 25 °C, 20 min, 82%; (v) CAL in SOL-Gel-AK, vinyl acetate, 30 °C, 49%.

Carter's approach (2003)⁴⁸

Carter *et al.* have made use of the Evans' *anti*-selective aldol approach as the key reaction for the synthesis of (+)-*epi*-cytosaxone (**58**). Thus, acylation of the commercially available (*R*)-oxazolidin-2-one **72** with 4-methoxyphenylacetic acid afforded imide **73**. The reaction of dibutylboryl enolate of **73** with pre-complexed solution of benzyloxyacetaldehyde **74** and 0.5 equiv of SnCl₄ provided the *anti*-aldol

75 (dr = 3:1). Removal of the chiral auxiliary from **75** provided the corresponding acid **76**, which was transformed into the oxazolidinone **77** in a one-pot 3 step procedure: (i) acyl azide formation, (ii) Curtius rearrangement and (iii) isocyanate trapping. Oxazolidinone **77** was debenzylated using Pearlman's catalyst to provide (+)-*epi*-cytoxazone (**58**) (Scheme 18).

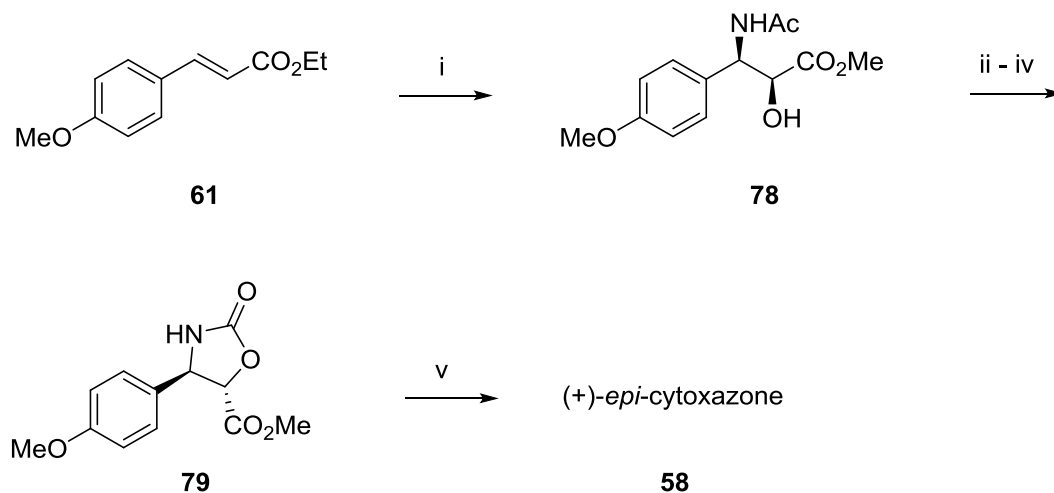


Scheme 18: (i) Bu₂BOTf, ^tPr₂EtN, 0 °C, 30 min, then BnOCH₂CHO precomplexed with 0.5 equiv SnCl₄, -78 °C, 3 h, 64%; (ii) H₂O₂, LiOH, THF:H₂O (4:1), 0 °C, 1 h, 99%; (iii) (PhO)₂PON₃, CH₂Cl₂, 23 °C, 40 min, 45 °C, 12 h, 61%; (iv) H₂ (1 atm), Pd(OH)₂, CH₃OH, 23 °C, 24 h, 84%.

Saicic's approach (2004)⁴⁹

Saicic's approach was based on the Sharpless' asymmetric aminohydroxylation reaction, starting from methyl *p*-methoxycinnamate **61** in five steps with 36% overall yield. *p*-Methoxycinnamate **61** was thus subjected to Sharpless aminohydroxylation with catalytic amount of K₂OsO₂(OH)₄, (DHQD)₂PHAL and BrNHAc as amine source to provide *syn*-amino alcohol **78** in 72% yield and 98% ee. Hydrolysis of aminoalcohol **78** under acidic medium gave hydroxy amino acid, which on

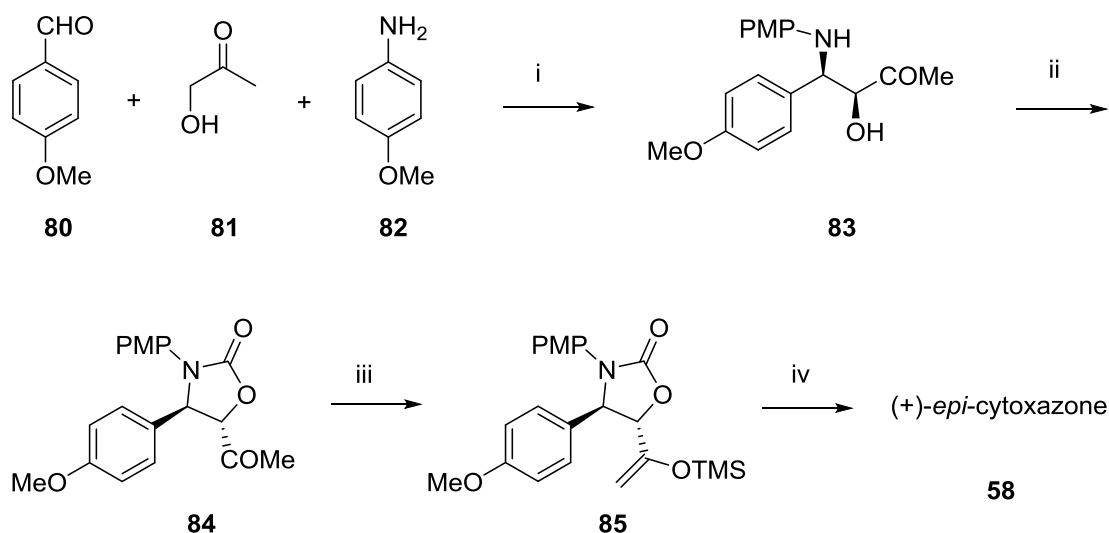
subsequent treatment with diphosgene, followed by esterification with diazomethane provided ester **79**. Reduction of ester **79** with sodium borohydride gave (+)-*epi*-cytosaxone (**58**) in 80% yield (**Scheme 19**).



Scheme 19: (i) $K_2[OsO_2(OH)_4]$ (4 mol %), $BrNHAc$, $(DHQD)_2PHAL$ (1 mol%), $LiOH$, H_2O , *tert*- $BuOH$, 4 °C, 20 h, 72%; (ii) 10% HCl , reflux, 4 h; (iii) $ClCO_2CCl_3$, $NaOH$, H_2O , 0 °C; (iv) CH_2N_2 , THF , 63%; (v) $NaBH_4$, THF , 0 °C, 80%.

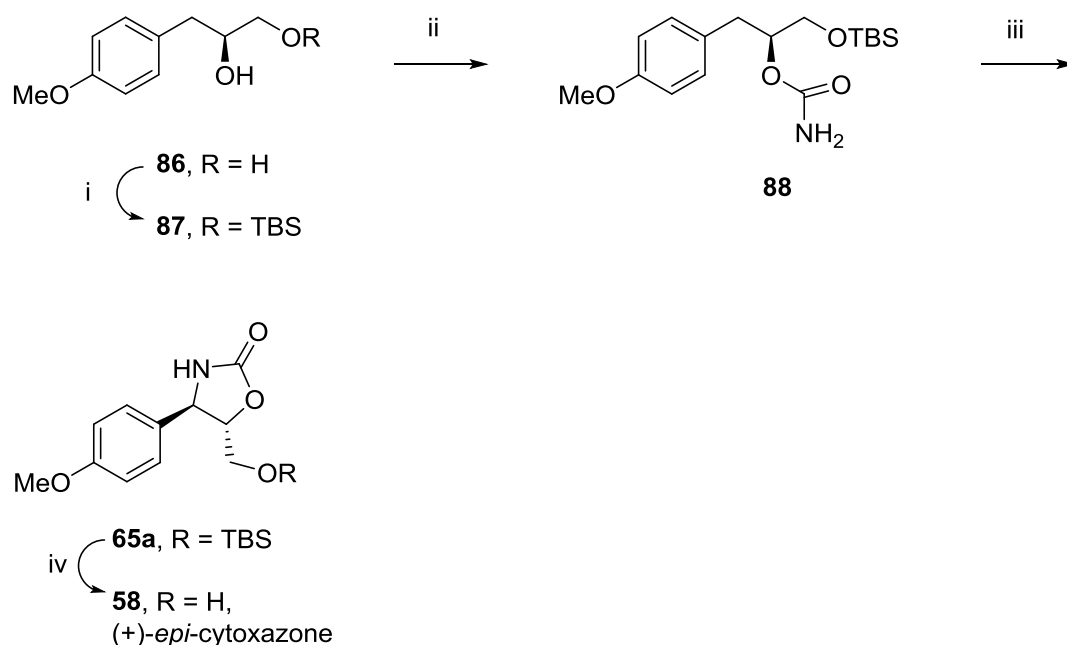
Sudalai's approach (2006), (2007) and (2010)^{50,51,52}

Sudalai *et al.* have developed a simple method for the enantioselective synthesis of (+)-*epi*-cytosaxone (**58**) using L-proline-catalyzed asymmetric Mannich reaction. Thus, the key intermediate *syn*-amino alcohol **83** was obtained from L-proline-catalyzed asymmetric Mannich reaction of 4-methoxybenzaldehyde (**80**), hydroxyacetone **81** and *p*-anisidine **82** in 76% yield with *syn/anti* ratio (2:1). Amino alcohol **83** was then protected with triphosgene to give oxazolidinone **84** in 82% yield. The *in situ* generated silyl enol ether **85** was then subjected to ozonolysis without purification. Reductive work up of ozonide and PMP deprotection with CAN provided (+)-*epi*-cytosaxone (**58**) in 59% yield and 81% ee (**Scheme 20**).



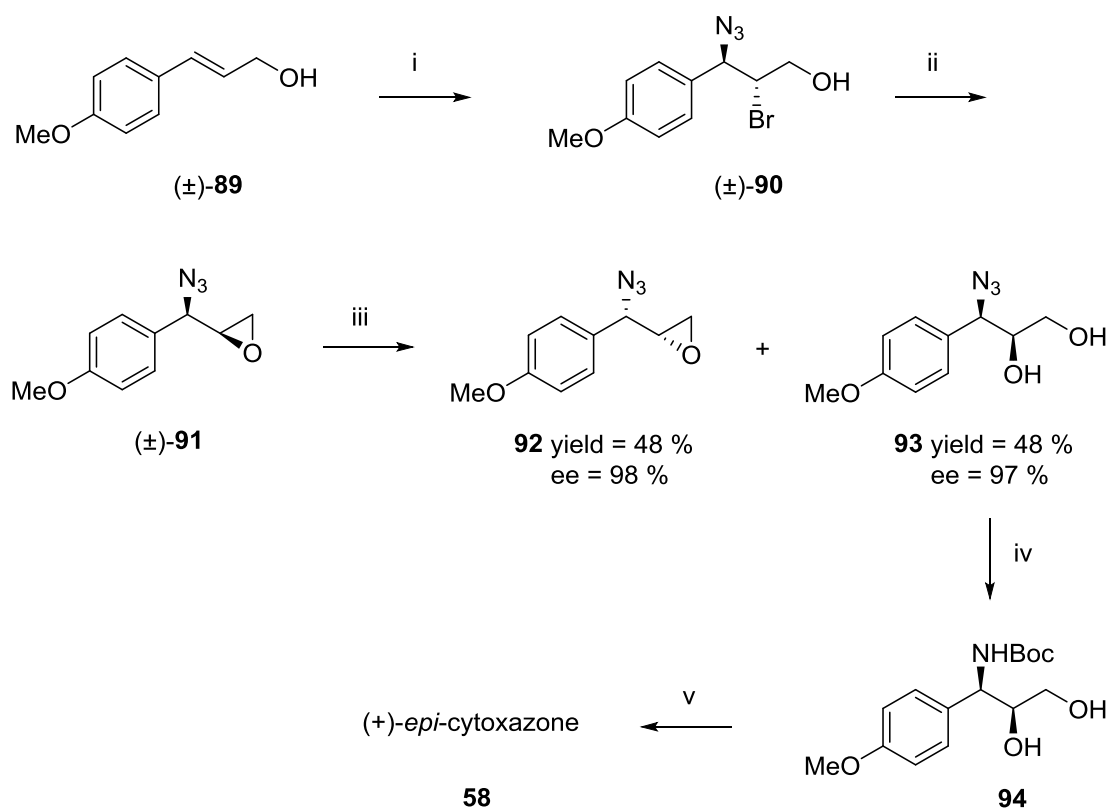
Scheme 20: (i) L-proline, DMSO, 25 °C, 24 h, 76%; (ii) triphosgene, Et₃N, CH₂Cl₂, -10 to 25 °C, 82%; (iii) LiHMDS, TMSCl, THF, -78 °C; (iv) (a) O₃, PPh₃, CH₂Cl₂, -78 °C; (b) NaBH₄, CH₃OH, 25 °C; (c) CAN, CH₃CN, 5 h, 59% (in three steps), 81% ee.

Sudalai *et al.* have also developed another useful route for the enantioselective synthesis of (+)-*epi*-cytosazone (**58**) commencing from the diol **86**, which was obtained by two different routes: (i) hydrolytic kinetic resolution and (ii) proline-catalyzed α -aminoxylation. Primary alcohol of diol **86** was selectively protected with TBSCl to give the secondary alcohol **87**, which was then converted to carbamate **88** in 92% yield using reported conditions (trichloroacetyl isocyanate, CH₂Cl₂, then K₂CO₃, MeOH, H₂O). Carbamate **88** underwent C-H insertion on treatment with catalytic amount of Rh₂(OAc)₄ (2 mol%), PhI(OAc)₂ and MgO in CH₂Cl₂ at 40 °C to afford the corresponding oxazolidinone **65a** with *syn* diastereoselectivity (5.5:1) in 87% combined yield. The *syn*-diastereomer, namely oxazolidinone **65a**, was readily separated by column chromatography. Finally, deprotection of the TBS group using TBAF in THF furnished (+)-*epi*-cytosazone (**58**) in 92% yield (**Scheme 21**).



Scheme 21: (i) TBSCl, imid., CH₂Cl₂, 25 °C, 98%; (ii) trichloroacetyl isocyanate, CH₂Cl₂, 0 °C to 25 °C, 2 h, then K₂CO₃, CH₃OH, H₂O, 0 °C to 25 °C, 12 h, 92%; (iii) Rh₂(OAc)₄ (2 mol%), PhI(OAc)₂, MgO, CH₂Cl₂, 40 °C, 87%, *syn:anti* (5.5:1); (iv) TBAF, THF, 92%.

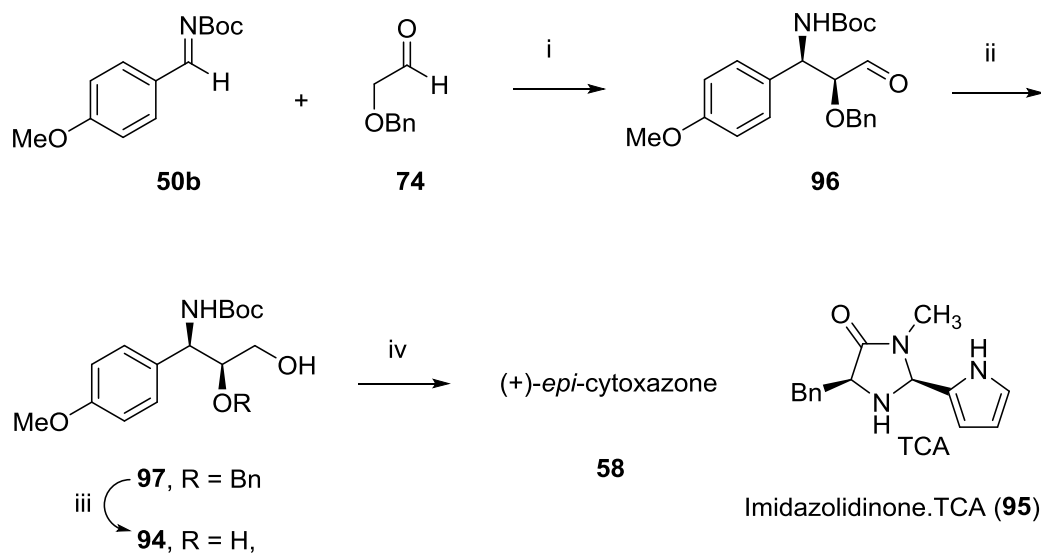
In yet another approach Sudalai *et al.* have employed two stereo-centered hydrolytic kinetic resolution of *syn*-azido epoxide **91** as the key step. Here, the synthesis commenced from 4-methoxycinnamyl alcohol **89**, which on azidobromination with NBS and NaN₃ afforded (±)-*anti*-bromo azide **90**. (±)-*syn*-Azido epoxide **91** was obtained from (±)-*anti*-bromo azide **90** under basic conditions and subjected to two stereo-centered hydrolytic kinetic resolution to give chiral *syn*-azido epoxide **92** and the corresponding diol **93**. Chiral azido diol **93** was then subjected to one pot azide reduction followed by Boc protection to give N-Boc amino diol **94**. Regioselective intramolecular cyclization of amino diol **94** with NaH gave (+)-*epi*-cytosazone (**58**) in 43% yield and 97% ee (**Scheme 22**).



Scheme 22: (i) NBS, NaN₃, CH₃CN:H₂O (4:1), 0 °C, 75%; (ii) LiOH, THF:H₂O (4:1), 0 °C to 25 °C, 3 h, 76%; (iii) (*R,R*)-Co(salen)OAc (0.5 mol%), THF, H₂O (0.5 equiv), 0 °C, 12 h; (iv) polymethylhydro siloxane (PMHS), 10 % Pd/C, (Boc)₂O, EtOH, 25 °C, 4 h, 95 %; (v) NaH, THF, 25 °C, 3 h, 96%.

Kim's approach (2008)⁵³

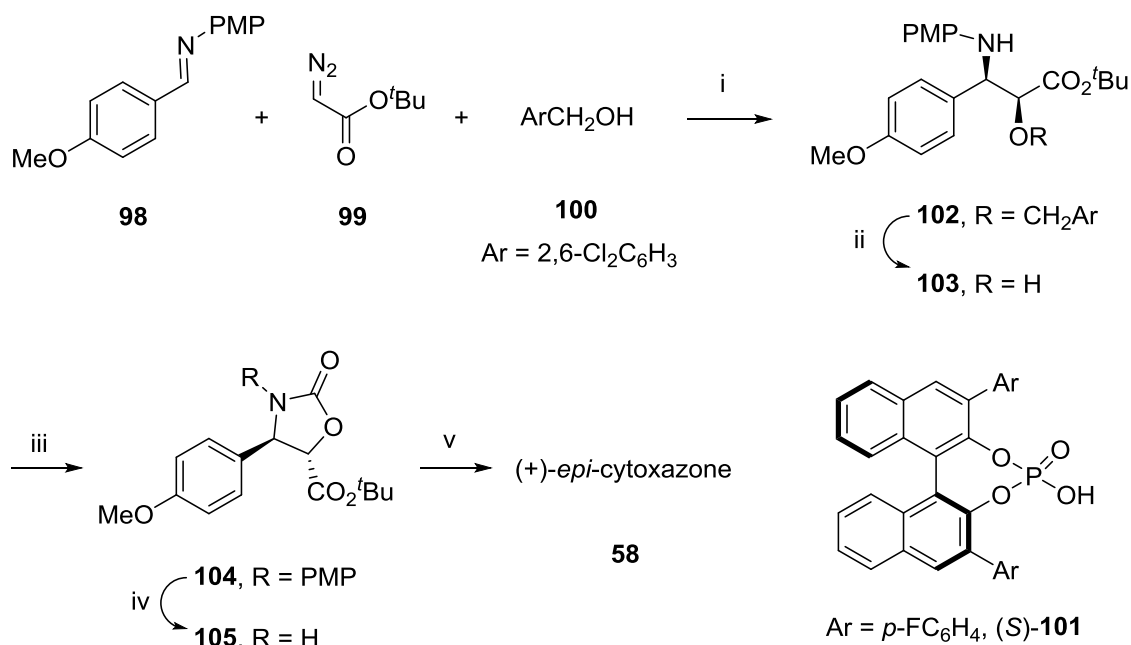
Kim *et al* have reported the synthesis of (+)-*epi*-cytosazone (58) by employing Mannich reaction as the key step. Thus, N-Boc-imine **50b** was treated with benzyloxy acetaldehyde **74** in the presence of a newly developed catalyst namely 2-pyrrole-derived imidazolidinone.TCA salt **95** to afford the corresponding β-aminoaldehyde **96** in 78% yield with a diastereomeric excess of 70% for the *syn*-isomer. Aldehyde **96** was then reduced with NaBH₄ to afford β-aminoalcohol **97** in 98% yield. Deprotection of **97** by hydrogenolysis of the benzyl group afforded the desired diol **94** in 91% yield. Finally, treatment of compound **94** with sodium hydride in THF gave (+)-*epi*-cytosazone (**58**) in 85% yield (**Scheme 23**).



Scheme 23: (i) imidazolidinone.TCA (**95**) (20 mol%), CHCl_3 , -30 °C, 78%; (ii) NaBH_4 , CH_3OH , 0 °C to 25 °C, 98%; (iii) 10% Pd/C, H_2 (1 atm), EtOH, 91%; (iv) NaH, THF, 85%.

Qian's approach (2010)⁵⁴

Qian *et al* have reported the synthesis of (+)-*epi*-cytoxazone (**58**) utilizing chiral Bronsted acid and Rh(II)-catalyzed three component reaction as the key step. Key intermediate *i.e.* amino alcohol **102** was obtained in 68% yield with 80% ee (*syn* : *anti* = 57:43) from $\text{Rh}_2(\text{OAc})_4$ catalyzed three-component reaction of imine **98**, ethyl diazoacetate (**99**) and 2,6-dichlorophenylmethanol (**100**) in the presence of chiral (*S*)-phosphoric acid **101** as cocatalyst. Deprotection of **102** by hydrogenolysis of the benzyl group provided the alcohol **103**, which was then protected with triphosgene to give oxazolidinone **104** in 87% yield. Subsequent PMP deprotection with CAN followed by reduction of ester with superhydride resulted in the formation of (+)-*epi*-cytoxazone (**58**) (Scheme 24).



Scheme 24: (i) Rh₂(OAc)₄ (2 mol %), phosphoric acid **101** (5 mol %), 4 Å MS, CH₂Cl₂, 2 h, 68%; (ii) 10% Pd/C (1 atm), CH₃OH, 35 °C, 2 h, 84%; (iii) triphosgene, ^tPr₂EtN, CH₂Cl₂, 0 °C, 2.5 h, 87%; (iv) CAN, CH₃CN:H₂O, 0 °C, 1 h, 62%; (v) LiEt₃BH, THF, 0 °C, 4 h, 62%.

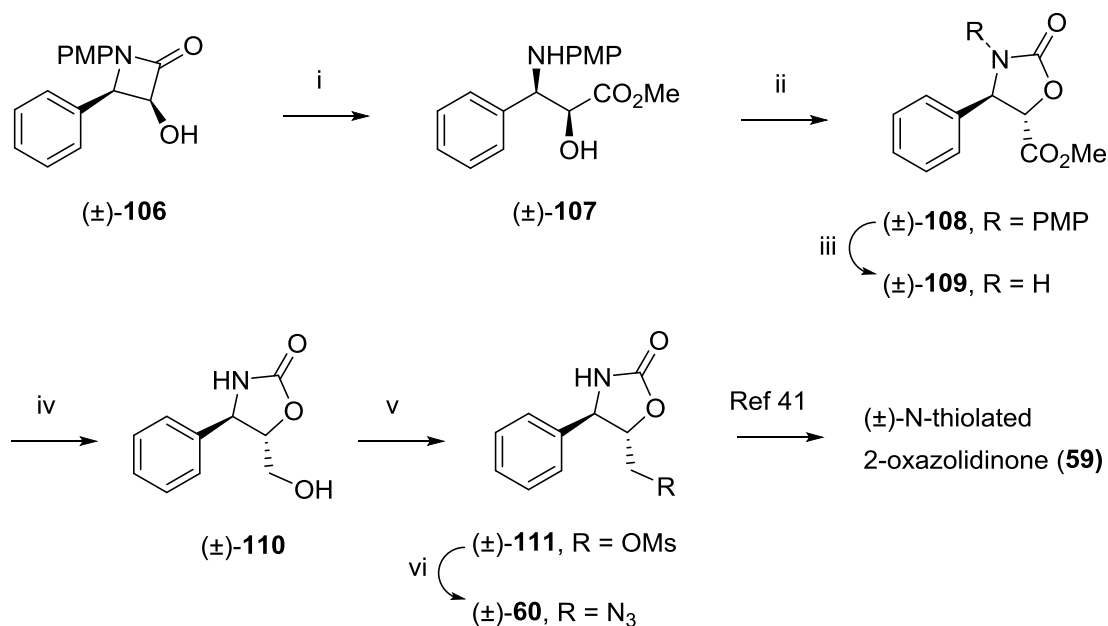
(b) Review of Literature for *N*-Thiolated-2-oxazolidinone (**59**)

Literature search revealed that there is only one report available for the racemic synthesis of antiMRSA *N*-thiolated-2-oxazolidinone (**59**) and its precursor 2-oxazolidinone (**60**), which is presented below.

Turos's approach (2007)⁵⁵

Turos *et al.* have reported the synthesis of (±)-*N*-thiolated-2-oxazolidinone (**59**), starting from lactam (±)-**106**. This was hydrolyzed with Me₃SiCl in refluxing methanol to afford the *syn*-aminol (±)-**107** in 95% yield. Treatment of (±)-**107** with triphosgene and Hunig's base in CH₂Cl₂ led to the isolation of oxazolidinone (±)-**108** (80% yield). The *N*-methoxyphenyl moiety on the oxazolidinone ring was then cleaved (CAN, CH₃CN and H₂O) to give the *N*-protio oxazolidinone (±)-**109** in 70% yield. The ester functionality of oxazolidinone (±)-**109** was selectively reduced

(NaBH₄, aq. THF) to furnish the alcohol (±)-**110** in 92% yield. The conversion of the hydroxyl group into an azide group proceeded through mesylate (±)-**111**; It gave the azide (±)-**60** in 72% yield after treatment with sodium azide in DMF (**Scheme 25**).



Scheme 25: (i) Me₃SiCl, CH₃OH, reflux, 95 %; (ii) triphosgene, ^tPr₂EtN, CH₂Cl₂, 0 °C to 23 °C, 80%; (iii) (NH₄)₂Ce(NO₃)₆, CH₃CN:H₂O, 0 °C to 23 °C, 70%; (iv) NaBH₄, aq. THF, CH₂Cl₂, 0 °C to 25 °C, 92%; (v) MsCl, Et₃N, CH₂Cl₂, 0 °C to 25 °C, 98 %; (vi) NaN₃, DMF, reflux, 12 h, 72%.

3.2.4 Present Work:

3.2.4.1 Objective

Literature search revealed that several methods such as chemo-enzymatic and metal-catalyzed enantioselective synthesis have been reported for (+)-*epi*-cytosaxone (**58**). However, these methods suffer from disadvantages such as low overall yields, the use of expensive chiral reagents, usage of protecting groups, especially the need for separation of diastereomers. The synthetic precursors of (+)-*epi*-cytosaxone (**58**) and *N*-thiolated 2-oxazolidinone (**59**) are found to be *syn*-amino diols, which have been the subject of thorough synthetic efforts in recent years. We became interested in providing, a more practical method for the synthesis of (+)-*epi*-cytosaxone (**58**) and

N-thiolated 2-oxazolidinone (**59**). In this section, we describe a concise protecting group-free enantioselective synthesis of (+)-*epi*-cytosazone (**58**) and 2-oxazolidinone **60** using α -aminoxylation of β -aminoaldehydes as the key reaction.

Retrosynthetic analysis of (+)-*epi*-cytosazone (**58**) revealed that *syn*-amino diol **112** could be visualized as the key intermediate, which in turn can be obtained using α -aminoxylation of β -aminoaldehyde **51b** (**Fig. 12**).

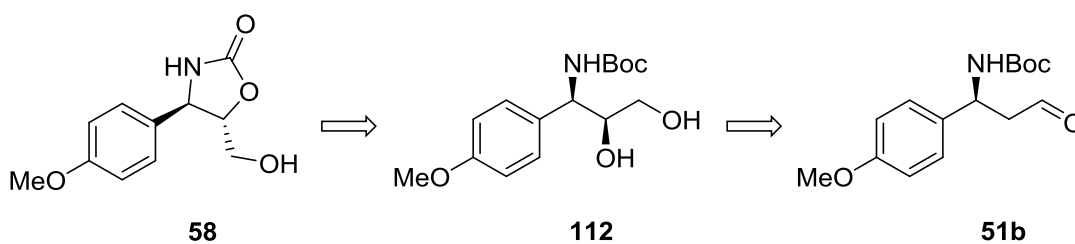


Fig. 12: Retrosynthetic analysis of (+)-*epi*-cytosazone (**58**)

Similarly, we envisaged that 2-oxazolidinone (**60**) could be obtained from *syn*-amino diol **113** through cyclization followed by azide displacement of hydroxyl group. The required *syn*-amino diol **113** could be prepared by means of α -aminoxylation of the corresponding β -aminoaldehyde **51a** (**Fig. 13**).

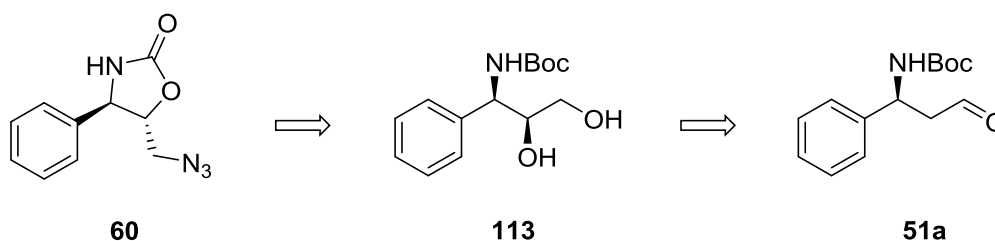
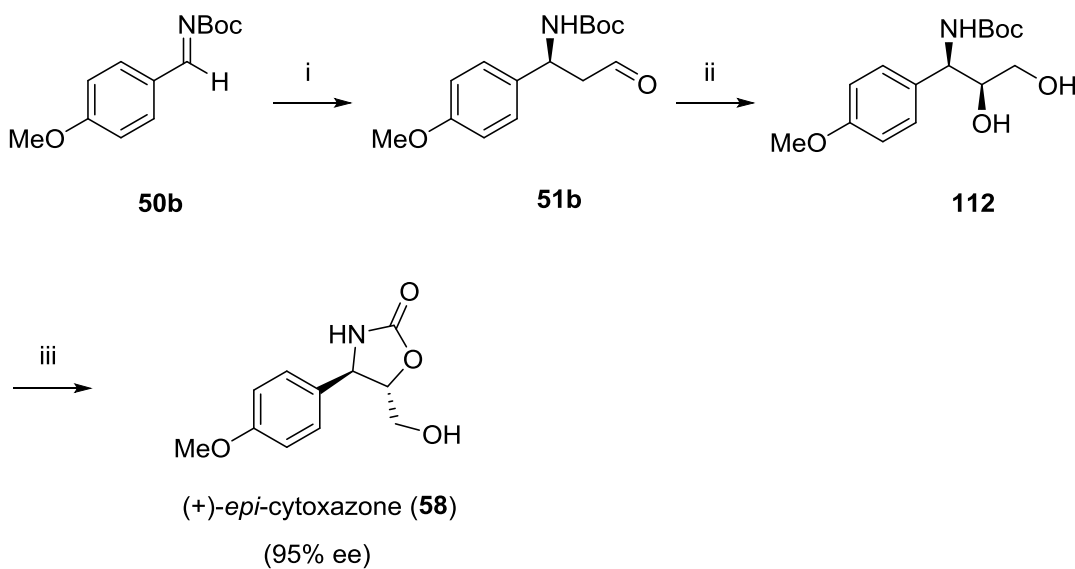


Fig. 13: Retrosynthetic analysis of 2-oxazolidinone (**60**)

3.2.5 Results and Discussion

(a) Concise Enantioselective Synthesis of (+)-*epi*-Cytoxazone (**58**)

The complete synthetic sequence for (+)-*epi*-cytoxazone (**58**), wherein α -amino oxylation of β -aminoaldehyde **51b** constitutes a key step, is presented in **Scheme 26**.



Scheme 26: (i) CH₃CHO, L-proline (20 mol%), CH₃CN, 0 °C, 3 h, 56%; (ii) (a) PhNO (0.8 equiv), D-proline (20 mol%), CH₃CN, -10 °C, 18 h; then NaBH₄, CH₃OH, 0 °C, 10 min; (b) Cu(OAc)₂·H₂O, CH₃OH, 25 °C, 16 h, 68% (over two steps); (iii) NaH, dry THF, 25 °C, 3 h, 90%.

Accordingly, the synthesis of (+)-*epi*-cytoxazone (**58**) was undertaken commencing from arylaldimine **50b**, which on subjecting to Mannich reaction with acetaldehyde [L-proline, CH₃CN, 0 °C, 3 h] gave β -aminoaldehyde **51b** in 56% yield. Compound **51b** was confirmed from its ¹H and ¹³C NMR spectroscopy. The ¹H NMR spectrum of **51b** showed a typical triplet at δ 9.72 (t, J = 1.6 Hz, 1H) and a multiplet at δ 2.80-2.94 (m, 2H) corresponding to aldehydic and homobenzylic protons respectively. Its ¹³C NMR spectrum showed two characteristic carbon signals at δ 193.4 and 200.0 due to carbamate and aldehydic carbonyl carbons respectively (**Fig. 14**).

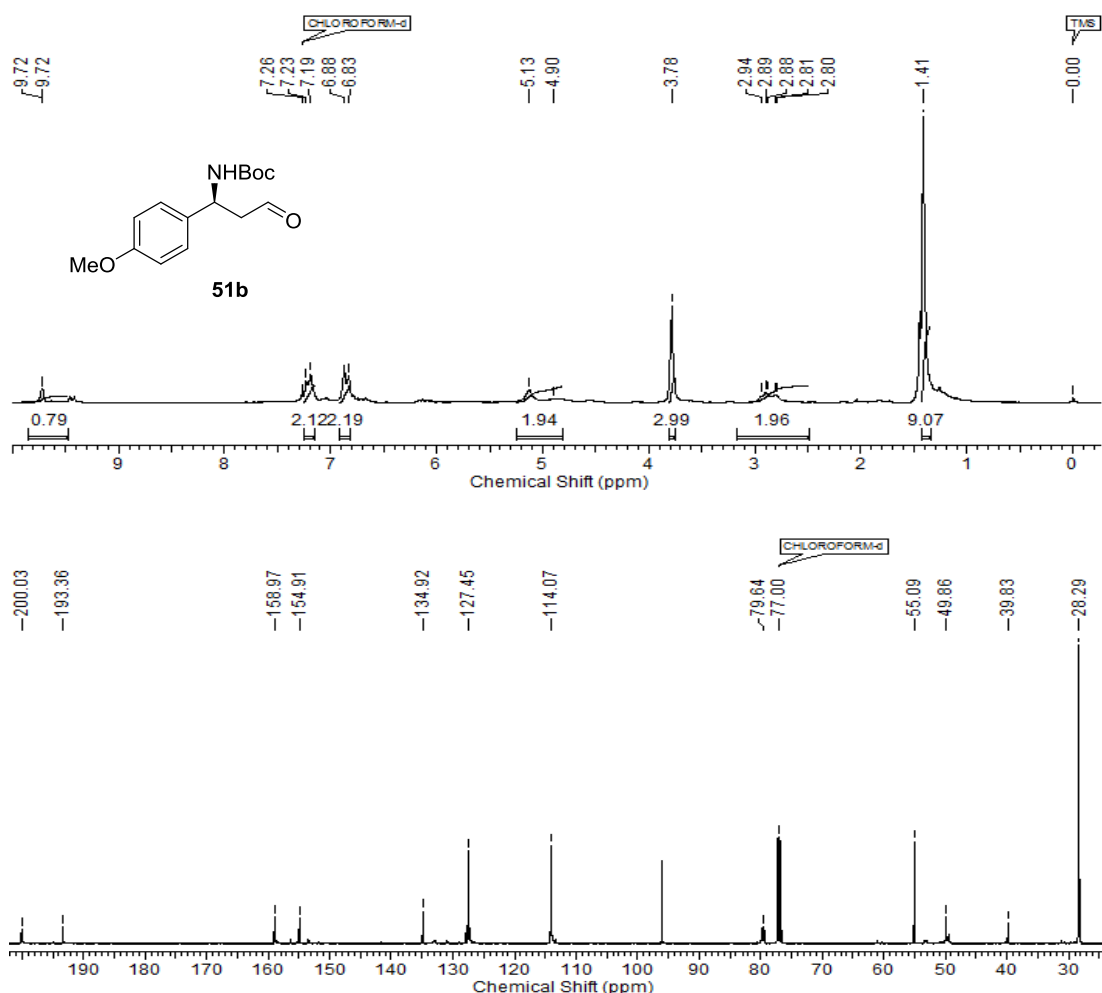


Fig. 14: ^1H and ^{13}C NMR spectra of β -aminoaldehyde **51b**

β -Aminoaldehyde **51b** was subjected to α -aminoxylation [D-proline, PhNO] to give α -aminoxyaldehyde, which was *in situ* reduced with NaBH_4 . It was then followed by subsequent reduction of the crude aminoxy product with $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ that provided a single diastereomer of *syn*-3-amino-1,2-diol **112** in 68% yield and 95% ee. The formation of *syn*-3-amino-1,2-diol **112** was confirmed from its ^1H NMR spectrum, which showed a characteristic multiplet at δ 3.42-3.56 (m, 2H) due to methylene (- CH_2 -OH) protons. Its ^{13}C NMR spectrum showed two characteristic carbon signals at δ 63.1 and 74.1 attributed to methylene and methine carbons attached to hydroxyl groups respectively. The mass spectrum also confirmed the formation of *syn*-3-amino-1,2-diol **112** (Fig. 15).

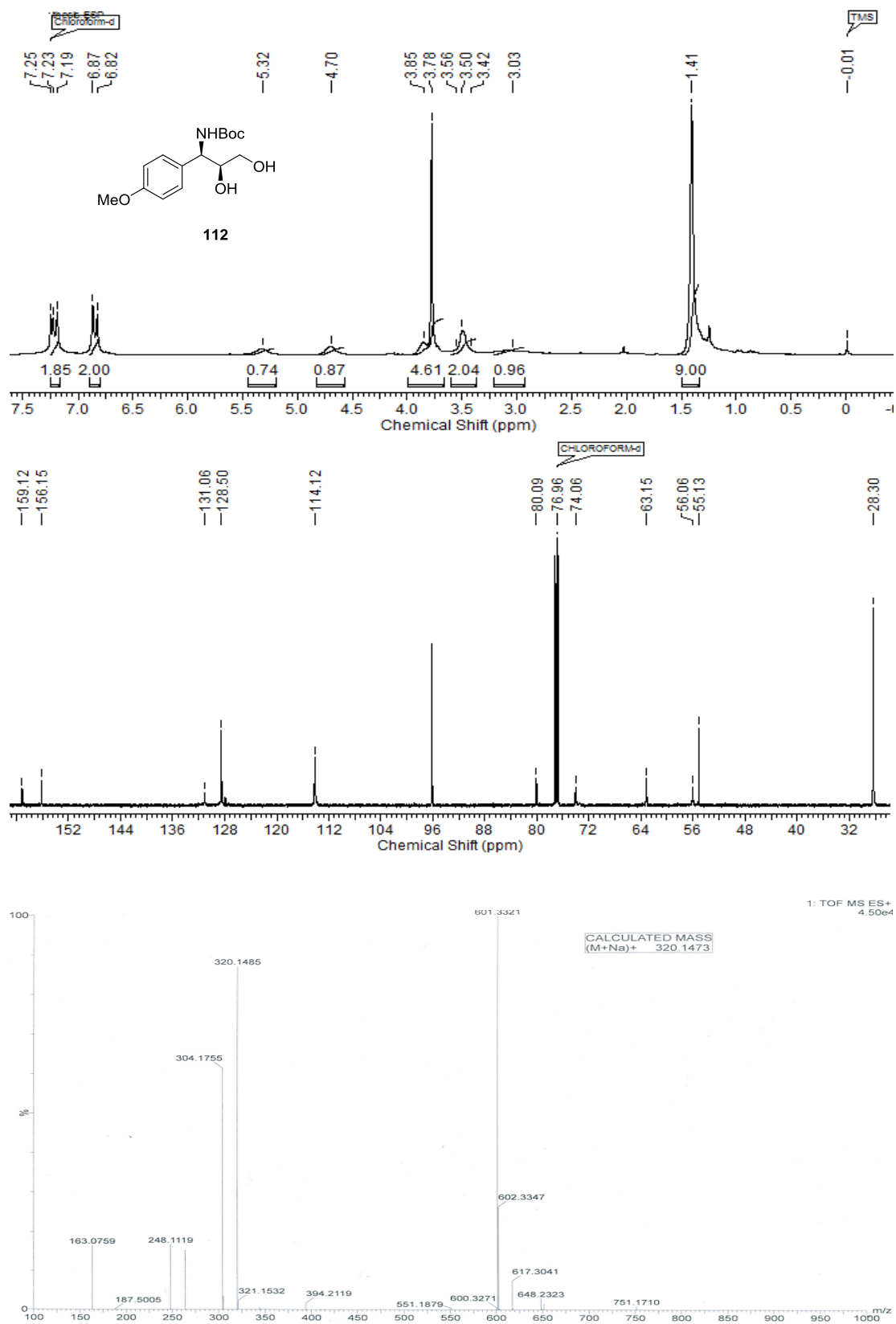


Fig. 15: ¹H, ¹³C NMR and mass spectra of *syn*-3-amino-1,2-diol **112**

Finally, the regioselective intramolecular cyclization of **112** using NaH in THF gave (+)-*epi*-cytoxazone (**58**) in 90 % yield and 95% ee.

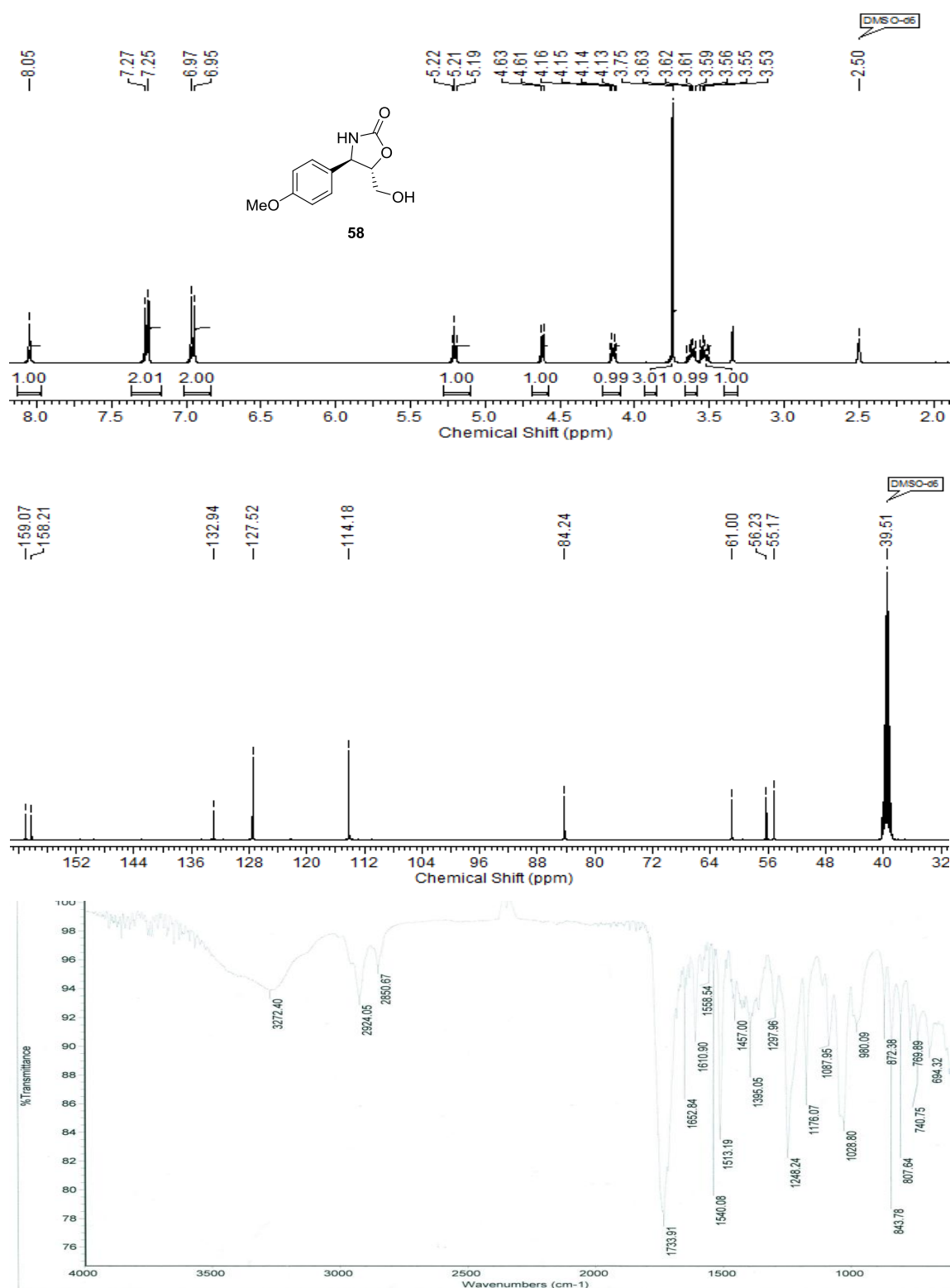
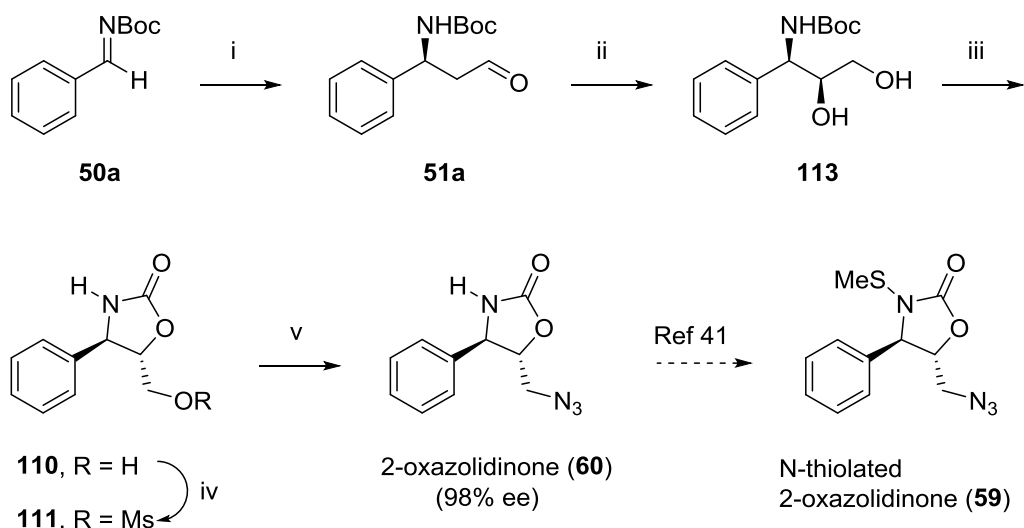


Fig. 16: ¹H, ¹³C NMR and IR spectra of (+)-*epi*-cytoxazone (**58**)

The ^1H , ^{13}C NMR and IR spectra of **58** confirmed the formation of (+)-*epi*-cytosaxone (**Fig. 16**). The ^1H NMR spectrum of (+)-*epi*-cytosaxone (**58**) showed a typical signal at δ 8.05 (s, 1H) for N-H proton of oxazolidinone ring. Its ^{13}C NMR spectrum showed a characteristic signal at δ 159.1 due to the carbonyl carbon in oxazolidinone ring. Its IR spectrum exhibited a characteristic oxazolidinone carbonyl absorption frequency at 1733 cm^{-1} . The spectral data of (+)-*epi*-cytosaxone (**58**) were in complete agreement with the reported values.⁵¹

(b) Facile Enantioselective Synthesis of 2-Oxazolidinone (**60**)

The synthetic scheme for 2-oxazolidinone (**60**), wherein α -aminoxylation of β -amino aldehyde **51a** constitutes a key step, is presented in **Scheme 27**.



Scheme 27: (i) CH_3CHO , L-proline (20 mol%), CH_3CN , $0\text{ }^\circ\text{C}$, 3 h, 55%; (ii) (a) PhNO (0.8 equiv), D-proline (20 mol%), CH_3CN , $-10\text{ }^\circ\text{C}$, 18 h; then NaBH_4 , CH_3OH , $0\text{ }^\circ\text{C}$, 10 min; (b) $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$, CH_3OH , $25\text{ }^\circ\text{C}$, 16 h, 64% (over two steps); (iii) NaH, dry THF, $25\text{ }^\circ\text{C}$, 3 h, 94%; (iv) MsCl, Et_3N , CH_2Cl_2 , $0\text{ }^\circ\text{C}$, 1 h; (v) NaN_3 , DMF, $60\text{ }^\circ\text{C}$, 12 h, 82% (over two steps).

The present synthesis of 2-oxazolidinone **60** commenced from benzaldimine **50a**, which on subjecting to Mannich reaction with acetaldehyde [L-proline, CH₃CN, 0 °C, 3 h] gave β-aminoaldehyde **51a** in 55% yield. β-Aminoaldehyde **51a** on subjecting to α-aminoxylation [D-proline, PhNO] provided α-aminoxyaldehyde, which was *in situ* reduced with NaBH₄. It was then followed by immediate reduction of the crude aminoxy product with Cu(OAc)₂·H₂O that provided a single diastereomer of *syn*-3-amino-1,2-diol **113** in 64% yield and 98% ee.

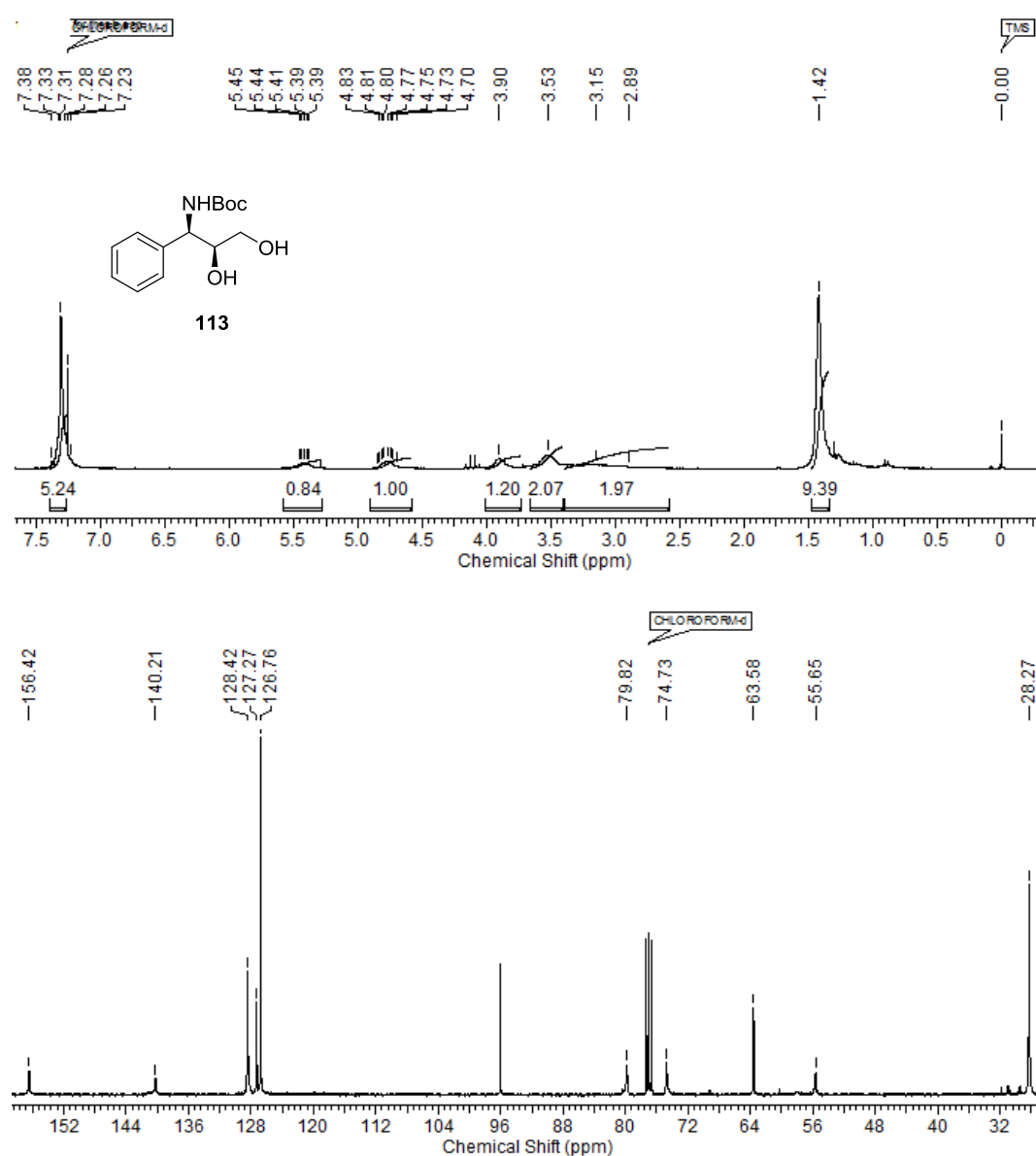


Fig. 17: ¹H and ¹³C NMR spectra of *syn*-3-amino-1,2-diol **113**

The formation of *syn*-3-amino-1,2-diol **113** was confirmed from its ^1H NMR spectrum, which showed a characteristic broad singlet at δ 3.53 (br s, 2H) for methylene ($-\text{CH}_2\text{-OH}$) protons. Its ^{13}C NMR spectrum also showed two characteristic carbon signals at δ 63.6 and 74.7 attributed to methylene and methine carbons attached to hydroxyl groups respectively (**Fig. 17**).

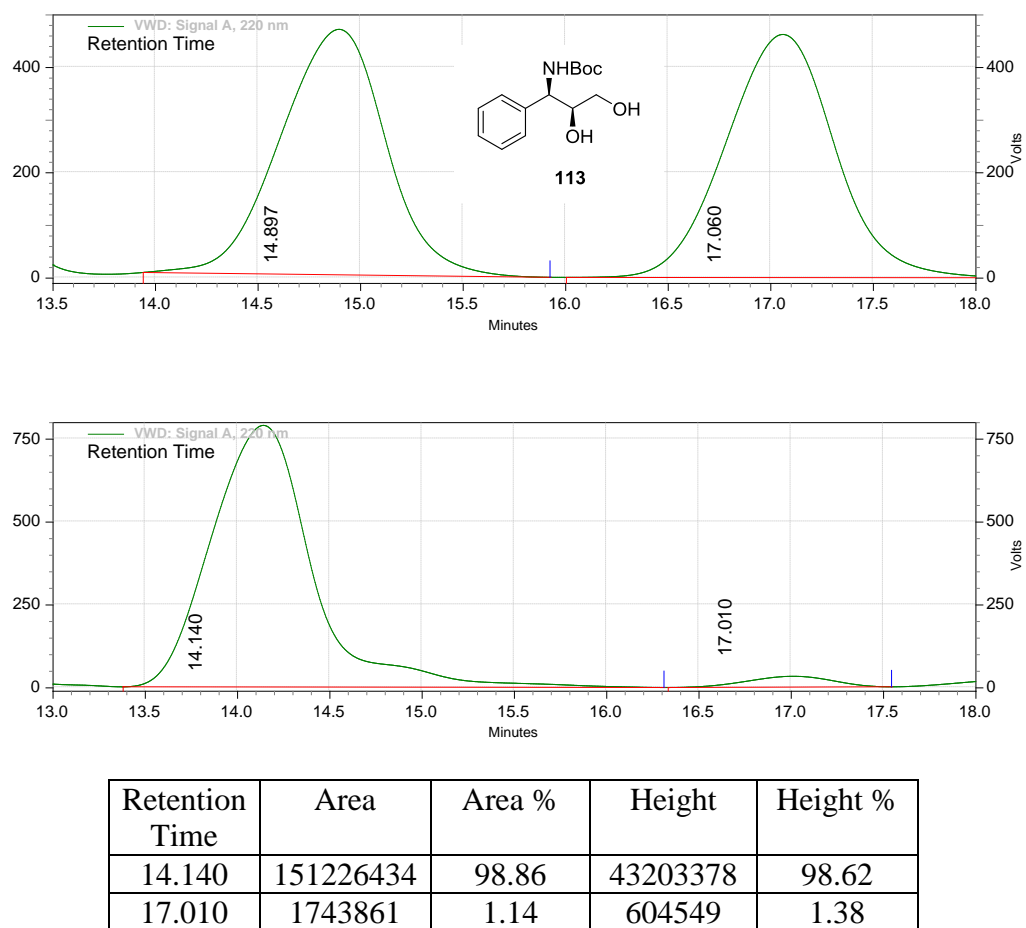


Fig. 18: HPLC chromatogram of *syn*-3-amino-1,2-diol **113**

The optical purity of *syn*-3-amino-1,2-diol **113** was determined to be 98% ee from chiral HPLC analysis (Chiracel AD-H, *n*-hexane/*i*PrOH, 85:15, 0.5 mL/min) retention time 14.14 min (98.86%) and 17.01 min (1.14%) (**Fig.18**).

The regioselective intramolecular cyclization of *syn*-3-amino-1,2-diol **113** with NaH in THF gave oxazolidinone **110** (mp: 106-108 °C; lit.⁵⁵ mp: 105-107 °C) in 94% yield. The formation of oxazolidinone **110** was confirmed from its ¹H and ¹³C NMR spectra. The ¹H NMR spectrum of oxazolidinone **110** showed a typical singlet at δ 8.14 (s, 1H) corresponding to the N-H proton of oxazolidinone ring. Its ¹³C NMR spectrum showed a characteristic signal at δ 158.1 due to the carbonyl carbon of oxazolidinone ring (Fig. 19).

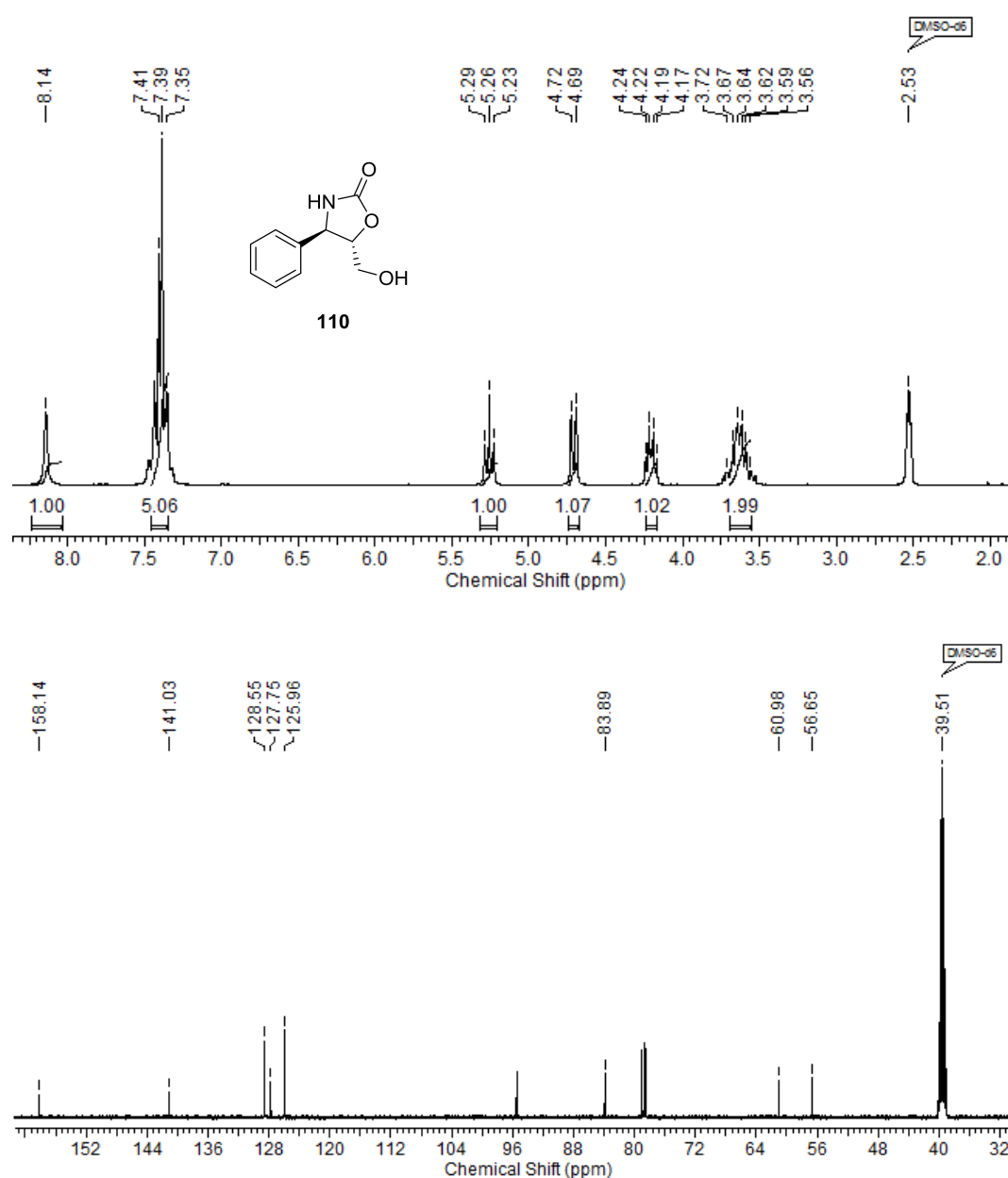


Fig. 19: ¹H and ¹³C NMR spectra of oxazolidinone **110**

Finally, mesylation of primary alcohol in oxazolidinone **110** gave the mesylate, which on subsequent displacement with NaN_3 in DMF at $60\text{ }^\circ\text{C}$ afforded (+)-2-oxazolidinone azide (**60**) in 82% yield.

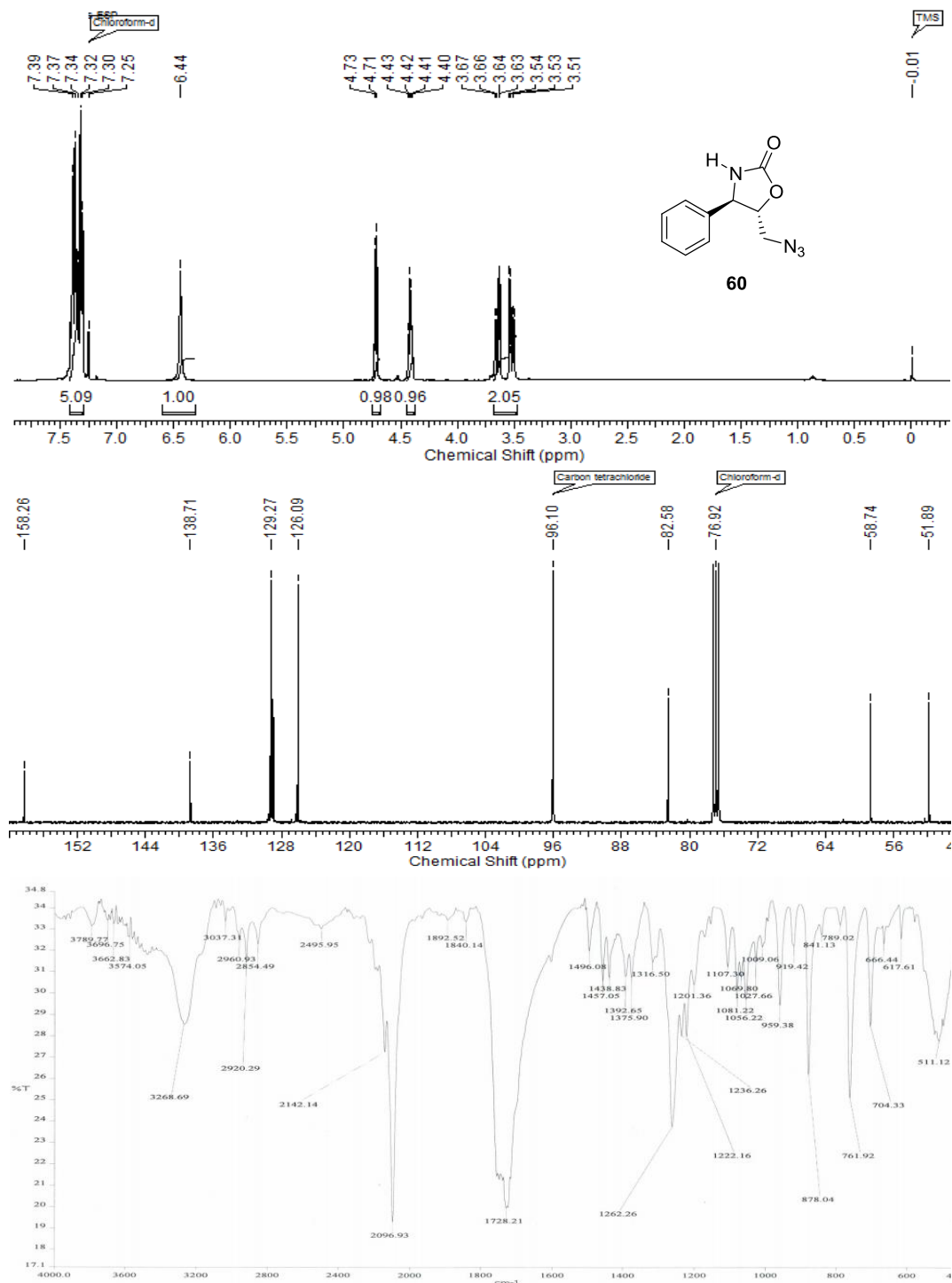


Fig. 20: ^1H , ^{13}C NMR and IR spectra of (+)-2-oxazolidinone **60**

The formation of (+)-2-oxazolidinone **60** was confirmed from its ^1H , ^{13}C NMR and IR spectra. The methylene proton attached to azide moiety resonated at δ 3.60 (dddd, $J = 4.5, 13.6$ Hz, 2H) in its ^1H NMR spectrum. It was further substantiated by the appearance of a carbon signal at δ 51.9 corresponding to carbon attached to azide group in its ^{13}C NMR spectrum. Its IR spectrum exhibited a characteristic strong absorption bands at 1728 cm^{-1} and 2096 cm^{-1} for oxazolidinone carbonyl and azide function respectively (**Fig. 20**). The conversion of (+)-2-oxazolidinone **60** to *N*-thiolated-2-oxazolidinone **59** has already been reported in the literature.⁴¹

3.2.6 Conclusion

A short and protecting group-free synthesis of (+)-*epi*-cytosazone (**58**) and (+)-2-oxazolidinone **60** has been achieved. D-Proline-catalyzed α -aminoxylation of β -aminoaldehydes was used as the key reaction, which proceeded to give high enantioselectivity. This methodology can be used for a viable synthesis of other diastereomers of (+)-*epi*-cytosazone (**58**) family by suitably employing L-proline as catalyst for α -aminoxylation of β -aminoaldehydes.

3.2.7 Experimental Section

Vide supra on section I of the same chapter for experimental procedure and spectral details of compounds **51a** and **51b**.

(2*S*, 3*R*)-3-(*tert*-Butoxycarbonylamino)-3-(4-methoxyphenyl)-1,2-propanediol (**112**)

To a stirred, precooled ($-10\text{ }^\circ\text{C}$) acetonitrile (25 mL) solution of β -aminoaldehyde **51b** (4.78 g, 17 mmol) and nitrosobenzene (1.45 g, 13.6 mmol) was added D-proline (0.039 g, 20 mol%). The reaction mixture was allowed to stir at the same temperature for 18 h followed by the addition of CH_3OH (10 mL) and NaBH_4 (0.97 g, 25 mmol)

to the reaction mixture. It was stirred for another 10 min. After addition of phosphate buffer, the resulting mixture was extracted with EtOAc (3 × 30 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude aminoxy alcohol, which was directly taken up for the next step without purification.

To a CH₃OH (25 mL) solution of the above crude aminoxy alcohol was added Cu(OAc)₂·H₂O (0.501 g, 2.6 mmol) at 25 °C and the reaction mixture was allowed to stir for 16 h at that temperature. After addition of phosphate buffer, the resulting mixture was extracted with CHCl₃ (3 × 30 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (30:70) as an eluent to give *syn*-amino diol **112** (2.74 g) as a colorless solid.

Yield: 68%; colorless solid; **mp:** 140-143 °C, (lit.⁵³ **mp:** 141-142 °C); $[\alpha]_D^{25}$ -36.5 (*c* 1.0, CHCl₃); lit.⁵³ $[\alpha]_D^{25}$ -36.1 (*c* 1.0, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 720, 832, 1070, 1250, 1512, 1688, 2934, 3384; **¹H NMR** (200 MHz, CDCl₃): δ 1.41 (s, 9H), 3.03 (br s, 1H), 3.42-3.56 (m, 2H), 3.78 (s, 3H), 3.85 (br s, 2H), 4.70 (br s, 1H), 5.32 (br s, 1H), 6.84 (d, *J* = 8.5 Hz, 2H), 7.21 (d, *J* = 8.5 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.3, 55.1, 56.1, 63.1, 74.1, 80.1, 114.1, 128.5, 131.1, 156.1, 159.1; **HRMS** (ESI) *m/z* calcd for C₁₅H₂₃NO₅ [M + Na]⁺: 320.1473, found: 320.1485; **Analysis:** C₁₅H₂₃NO₅ requires C, 60.59; H, 7.80; N, 4.71; found: C, 60.36; H, 7.63; N 4.62%.

(4*S*, 5*S*)-5-(Hydroxymethyl)-4-(4-methoxyphenyl)oxazolidin-2-one:
[(+)-*epi*-cytosazone] (58)

To a solution of *syn*-3-amino-1,2-diol **112** (0.3 g, 1.0 mmol) in dry THF (10 mL) was added NaH (0.05 g, 60% w/w, 2.0 mmol) at 25 °C, and the mixture was stirred under nitrogen atmosphere for 3 h. The reaction mixture was concentrated and the resulting mixture was extracted with EtOAc (3 × 10 mL), washed with saturated aq. NH₄Cl (5

mL) and brine solution (5 mL). The organic layers were separated, dried over anhyd. Na₂SO₄, and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (60:40) as an eluent to give **58** (0.20 g) as a colorless solid.

Yield: 90%; colorless solid, **mp:** 158-160 °C, (lit.⁵³ **mp:** 161-162 °C); $[\alpha]_D^{25} +31.90$ (*c* 1, MeOH); lit.⁵³ $[\alpha]_D^{25} +32.8$ (*c* 0.6, MeOH); **IR** (CHCl₃, cm⁻¹): ν_{\max} 769, 843, 1028, 1248, 1395, 1513, 1610, 1733, 2580, 2924, 3272; **¹H NMR** (400 MHz, DMSO-*d*₆): δ 3.50-3.53 (m, 1H), 3.59-3.65 (m, 1H), 3.75 (s, 3H), 4.13-4.16 (m, 1H), 4.62 (d, *J* = 6.3 Hz, 1H), 5.21 (t, *J* = 5.8 Hz, 1H), 6.96 (d, *J* = 8.6 Hz, 2H), 7.26 (d, *J* = 8.6 Hz, 2H), 8.05 (s, 1H); **¹³C NMR** (100 MHz, DMSO-*d*₆): δ 55.2, 56.2, 61.0, 84.2, 114.2, 127.5, 132.9, 158.2, 159.1; **Analysis:** C₁₁H₁₃NO₄ requires C, 59.19; H, 5.87; N, 6.27; found: C, 59.01; H, 5.69; N, 6.13%.

(2*S*, 3*R*)-3-(*tert*-Butoxycarbonylamino)-3-phenyl-1,2-propanediol (113)

To a stirred, precooled (-10 °C) acetonitrile (25 mL) solution of β -aminoaldehyde **51a** (4.23 g, 17 mmol) and nitrosobenzene (1.45 g, 13.6 mmol) was added D-proline (0.039 g, 20 mol%). The reaction mixture was allowed to stir at the same temperature for 18 h followed by the addition of CH₃OH (10 mL) and NaBH₄ (0.97 g, 25 mmol) to the reaction mixture. It was stirred for further 10 min. After addition of phosphate buffer, the resulting mixture was extracted with EtOAc (3 \times 30 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude aminoxy alcohol, which was directly taken up for the next step without purification.

To a CH₃OH (25 mL) solution of the above crude aminoxy alcohol was added Cu(OAc)₂.H₂O (0.501 g, 2.6 mmol) at 25 °C and the reaction mixture was allowed to stir for 16 h at that temperature. After addition of phosphate buffer, the resulting mixture was extracted with CHCl₃ (3 \times 30 mL) and the combined organic phases were

dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (35:65) as an eluent to give *syn*-amino diol **113** (2.32 g) as a colorless gum.

Yield: 64%; colorless gum; $[\alpha]_D^{25}$ -28.0 (*c* 0.7, CHCl₃); 98% ee from chiral HPLC analysis (Chiracel AD-H, *n*-hexane/*i*PrOH, 85:15, 0.5 mL/min) retention time 14.1 min (98.9%) and 17.0 min (1.1%); **IR** (CHCl₃, cm⁻¹): ν_{\max} 832, 1035, 1070, 1250, 1512, 1699, 2934, 3363; **¹H NMR** (200MHz, CDCl₃): δ 1.42 (s, 9H), 3.15 (br s, 2H), 3.53 (br s, 2H), 3.90 (br s, 1H), 4.70-4.85 (m, 1H), 5.39-5.45 (d, *J* = 5.0 Hz, 1H), 7.23-7.38 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.3, 55.6, 63.6, 74.7, 79.8, 126.8, 127.3, 128.4, 140.2, 156.4; **Analysis:** C₁₄H₂₁NO₄ requires C, 62.90; H, 7.92; N, 5.24; found: C, 62.69; H, 7.76; N, 5.10%.

(4S, 5S)-5-Hydroxymethyl-4-phenyloxazolidin-2-one (110)

To a stirred solution of *syn*-3-amino-1,2-diol **113** (0.267g, 1.0 mmol) in dry THF (10 mL) was added NaH (0.05 g, 60% w/w, 2.0 mmol) at 25 °C and the mixture was stirred under nitrogen atmosphere for 3 h. The reaction mixture was concentrated and the resulting mixture was extracted with EtOAc (3 x 10 mL), washed with saturated aq. NH₄Cl (5 mL) and brine solution (5 mL). The organic layers were separated, dried over anhyd. Na₂SO₄, and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (70:30) as an eluent to give **110** (0.181 g) as a colorless solid.

Yield: 94%; colorless solid, **mp:** 106-108 °C, (lit.⁵⁵ **mp:** 105-107 °C); $[\alpha]_D^{25}$ +26.7 (*c* 1, MeOH); **IR** (CHCl₃, cm⁻¹): ν_{\max} 772, 832, 1104, 1252, 1395, 1522, 1570, 1610, 1724, 2580, 2924, 3244; **¹H NMR** (200 MHz, DMSO-*d*₆): δ 3.56-3.72 (m, 2H), 4.17-4.24 (m, 1H), 4.70 (d, *J* = 6.2 Hz, 1H), 5.26 (d, *J* = 5.9 Hz, 1H), 7.35-7.41 (m, 5H), 8.14 (s, 1H); **¹³C NMR** (50 MHz, CDCl₃+DMSO-*d*₆) δ : 56.6, 60.9, 83.9, 125.9, 127.7,

128.5, 141.0, 158.1; **Analysis:** C₁₀H₁₁NO₃ requires C, 62.17; H, 5.74; N, 7.25; found: C, 62.03; H, 5.47; N, 7.13%.

(4S, 5S)-5-Azidomethyl-4-phenyloxazolidin-2-one (60)

To a stirred solution of alcohol **110** (0.791 g, 4.1 mmol) in CH₂Cl₂ (5 mL) was added Et₃N (0.82 g, 8.2 mmol) at 0 °C. After 5 min of stirring, methane sulfonyl chloride (0.47 g, 4.1 mmol) was added drop-wise over a period of 5 min. The reaction mixture was then stirred for another 1 h at 0 °C. After the completion of the reaction, as monitored by TLC, it was extracted with CH₂Cl₂ (3 x 20 mL) washed with water, brine and dried over anhyd. Na₂SO₄. The combined organic layers were concentrated under reduced pressure to give crude methane sulfonate ester **111** in almost quantitative yield. To a solution of the crude mesylate **111** in DMF (10 mL) was added NaN₃ (0.53 g, 8.2 mmol) and the reaction mixture was heated at 60 °C for 12 h. After the completion of the reaction, as monitored by TLC, it was extracted with EtOAc (3 x 20 mL) washed with water, brine and dried over anhyd. Na₂SO₄. The combined organic layers were concentrated under reduced pressure to give the crude 2-oxazolidinone **60**, which was purified by column chromatography using pet. ether:EtOAc (85:15) to give **60** (0.732 g) as colorless solid.

Yield: 82%; colorless solid, **mp:** 80-83 °C, (lit.⁵⁵ **mp:** 82-84 °C); [α]_D²⁵ +30.4 (*c* 1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 761, 878, 1222, 1262, 1457, 1728, 2096, 2142, 2920, 3268; **¹H NMR** (200 MHz, CDCl₃): δ 3.60 (dddd, *J* = 4.5, 13.6 Hz, 2H), 4.40-4.43 (m, 1H), 4.72 (d, *J* = 6.5 Hz, 1H), 6.44 (s, 1H), 7.30-7.39 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃) δ : 51.9, 58.7, 82.6, 126.1, 129.0, 129.3, 138.7, 158.3; **Analysis:** C₁₀H₁₀N₄O₂ requires C, 55.04; H, 4.62; N, 25.68; found: C, 55.09; H, 4.48; N, 25.48%.

3.2.8 References

1. Bloch, R. *Chem. Rev.* **1998**, *98*, 1407 and references cited therein.
2. (a) Rowinsky, E. K.; Cazenave, L. A.; Donehower, R. C. *J. natl. Cancer Inst.* **1990**, *82*, 1247; (b) Schiff, P. B.; Fant, J.; Horwitz, S. B. *Nature* **1979**, *277*, 665.
3. (a) For a recent review about deoxystreptamine, see: Busscher, G. F.; Rutjes, F. P. J. T.; van Delf, F. L. *Chem. Rev.* **2005**, *105*, 775; (b) Umezawa, W. *Adv. Carbohydr. Chem. Biochem.* **1974**, *30*, 111; (c) Rinehart, K. L.; Stroshane, R. M. *J. Antibiot.* **1976**, *29*, 319; (d) Daniels, P. J. L. *Kirk-Othmer Encycl. Chem. Technol.* **1978**, *2*, 819.
4. Bonini, C.; Chiummiento, L.; Bonis, M.; Funicello, M.; Lupattelli, P.; Suanno, G.; Berti, G.; Campaner, P. *Tetrahedron* **2005**, *61*, 6580.
5. Padron, J. M.; Martin, V. S.; Litina, H.; Noula, C.; Kokotou, V. C.; Peters, G. J.; Kokotos, G. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 821.
6. Konradi, A. W.; Pedersen, S. F. *J. Org. Chem.* **1990**, *55*, 4506.
7. (a) Jager, V.; Franz, T.; Hein, M.; Veith, U.; Peters, E.; Peters, K.; Schnering, H. G. V. *Angew. Chem., Int. Ed.* **1994**, *33*, 1298; (b) Jager, V.; Wehner, V. *Angew. Chem., Int. Ed.* **1989**, *28*, 469.
8. Bunnage, M. E.; Chernega, A. N.; Davies, S. G.; Goodwin, C. J. *J. Chem. Soc. Perkin Trans I*, **1994**, 2373.
9. Pasto, M.; Moyano, A.; Pericks, M. A.; Riera, A. *Tetrahedron: Asymmetry* **1995**, *6*, 2329.
10. Petasis, N. A.; Zavialov, I. A. *J. Am. Chem. Soc.* **1998**, *120*, 11798.
11. Merino, P.; Castillo, E.; Franco, S.; Merchtn, F. L.; Tejero, T. *Tetrahedron*, **1998**, *54*, 12301.
12. Chandrasekhar, S.; Mohapatra, S.; Yadav, J. S. *Tetrahedron*, **1999**, *55*, 4763.
13. Righi, G.; Chionne, A.; Bonini, C. *Eur. J. Org. Chem.* **2000**, 3127.
14. (a) Hutin, P.; Larcheveque, M. *Tetrahedron Lett.* **2000**, *41*, 2369; (b) Hutin, P.; Larcheveque, M. *Synthesis* **2000**, 220.
15. Kwon, S. J.; Ko, S. Y. *Bull. Korean Chem. Soc.* **2003**, *24*, 1053.
16. Bickley, J. F.; Roberts, S. M.; Runhui, Y.; Skidmore, J.; Smith, C. B. *Tetrahedron*, **2003**, *59*, 5731.
17. Concellon, J. M.; Suarez, J. R.; Granda, S. G.; Diaz, M. R. *Org. Lett.* **2005**, *7*, 247.
18. Yang, J. W.; Chandler, C.; Stadler, M.; Kampen, D.; List, B. *Nature* **2008**, *452*, 453.
19. (a) Hayashi, Y.; Yamaguchi, J.; Hibino, K.; Shoji, M. *Tetrahedron Lett.* **2003**, *44*, 8293; (b) Zhong, G. *Angew. Chem., Int. Ed.* **2003**, *42*, 4247; (c) Hayashi, Y.; Yamaguchi, J.; Sumiya, T.; Shoji, M. *Angew. Chem., Int. Ed.* **2003**, *43*, 1112; (d) Brown, S. P.; Brochu, M. P.; Sinz, C. J.; MacMillan, D. W. C. *J. Am. Chem. Soc.* **2003**, *125*, 10808; (e) Cordova, A.; Sunden, H.; Bogevig, A.; Johansson, M.; Himo, F. *Chem. Eur. J.* **2004**, *10*, 3673.
20. Seayad, J.; List, B. *Org. Biomol. Chem.* **2005**, *3*, 719.
21. List, B.; Lerner, R. A.; Barbas III, C. F. *J. Am. Chem. Soc.* **2000**, *122*, 2395.
22. (a) Sabitha, G.; Fatima, N.; Reddy, E. V.; Yadav, J. S. *Adv. Synth. Catal.* **2005**, *347*, 1353; (b) Ramachary, D. B.; Chowdari, N. S.; Barbas, C. F. III *Synlett* **2003**, 1910.
23. Hechavarria, F. M. T.; List, B.; *Angew. Chem., Int. Ed.* **2004**, *43*, 3958.

24. For α -functionalization reviews: (a) Franzen, J.; Marigo, M.; Fielenbach, D.; Wabnitz, T. C.; Kjarsgaard, A.; Jorgensen, K. A. *J. Am. Chem. Soc.* **2005**, *127*, 18296; (b) Guillena, G.; Ramon, D. J. *Tetrahedron: Asymmetry* **2006**, *17*, 1465.
25. For a review of proline-catalyzed asymmetric reactions see: (a) List, B. *Tetrahedron* **2002**, *58*, 5573; (b) Aleman, J.; Cabrera, S. *Chem. Soc. Rev.* **2013**, *42*, 774.
26. (a) Kleinnmann, E. F. In *Comprehensive Organic Synthesis*; Trost, B. M., Ed.; Pergamon Press: New York, 1991; Vol. 2, Chapter 4.1; (b) Arend, M.; Westermann, B.; Risch, N. *Angew. Chem., Int. Ed.* **1998**, *37*, 1044.
27. (a) Fujieda, H.; Kanai, M.; Kambara, T.; Iida, A.; Tomioka, K. *J. Am. Chem. Soc.* **1997**, *119*, 2060; (b) Ishitani, H.; Ueno, M.; Kobayashi, S. *J. Am. Chem. Soc.* **1997**, *119*, 7153; (c) Kobayashi, S.; Ishitani, H.; Ueno, M. *J. Am. Chem. Soc.* **1998**, *120*, 431. (d) Hagiwara, E.; Fujii, A.; Sodeoka, M. *J. Am. Chem. Soc.* **1998**, *120*, 2474; (e) Ferraris, D.; Young, B.; Dudding, T.; Lectka, T. *J. Am. Chem. Soc.* **1998**, *120*, 4548; (f) Yamasaki, S.; Iida, T.; Shibasaki, M. *Tetrahedron Lett.* **1999**, *40*, 307.
28. Mukherjee, S.; Yang, J. W.; Hoffmann, S.; List, B. *Chem. Rev.* **2007**, *107*, 5471.
29. (a) Enders, D.; Vrettou, M. *Synthesis* **2006**, 2155; (b) Yang, J. W.; Stadler, M.; List, B. *Angew. Chem. Int. Ed.* **2007**, *46*, 609.
30. (a) Porter, E. A.; Wang, X.; Lee, H. S.; Weisblum, B.; Gellman, S. H. *Nature* **2000**, *404*, 565; (b) Seebach, D. *Helv. Chim. Acta* **2000**, *83*, 2115.
31. (a) Davis, F. A.; Bang-Chi, C. *Chem. Rev.* **1992**, *92*, 919; (b) Morikawa, K.; Park, J.; Andersson, P. G.; Hashiyama, T.; Sharpless, K. B. *J. Am. Chem. Soc.* **1993**, *115*, 8463; (c) Adam, W.; Fell, R. T.; Stegmann, V. R.; Saha-Moller, C. R. *J. Am. Chem. Soc.* **1996**, *118*, 708; (d) Zhu, Y.; Tu, Y.; Yu, H.; Shi, Y. *Tetrahedron Lett.* **1998**, *39*, 7819.
32. Merino, P.; Tejero, T. *Angew. Chem. Int. Ed.* **2004**, *43*, 2995.
33. Davis, F. A.; Szweczyk, J. M. *Tetrahedron Lett.* **1998**, *39*, 5951.
34. Carswell, E. L.; Snapper, M. L.; Hoveyda, A. H. *Angew. Chem., Int. Ed.* **2006**, *45*, 7230.
35. Mishra, R. K.; Coates, C. M.; Revell, K. D.; Turos, E. *Org. Lett.* **2007**, *9*, 575.
36. Kakeya, H.; Morishita, M.; Kobinata, K.; Osono, M.; Ishizuka, M.; Osada, H. *J. Antibiot.* **1998**, *51*, 1126.
37. Kakeya, H.; Morishita, M.; Koshino, H.; Morita, T.; Kobayashi, K.; Osada, H. *J. Org. Chem.* **1999**, *64*, 1052.
38. Hamersak, Z.; Sepac, D.; Zihner, D.; Sunjic, V. *Synthesis* **2003**, 375.
39. List, B. *J. Am. Chem. Soc.* **2000**, *122*, 9336.
40. (a) Brickner, S. J. *Curr. Pharm. Des.* **1996**, *2*, 175; (b) Phillips, O. A. *Curr. Opin. Invest. Drugs* **2003**, *4*, 117; (c) Brickner, S. J.; Hutchinson, D. K.; Barbachyn, M. R.; Manninen, P. R.; Ulanowicz, D. A.; Garmon, S. A.; Grega, K. C.; Hendges, S. K.; Toops, D. S.; Ford, C. W.; Zurenko, G. E. *J. Med. Chem.* **1996**, *39*, 673.
41. (a) Mishra, R. K.; Revell, K. D.; Coates, C. M.; Turos, E.; Dickeyb, S.; Limb, D. V. *Bioorg. Med. Chem.* **2006**, *16*, 2081; (b) Steven, R. W.; Hisao, I.; Marvin, J. M.; *Tetrahedron Lett.* **1985**, *26*, 3891.

42. (a) Seder, R. A.; Paul, W. E. *Annu. Rev. Immunol.* **1994**, *12*, 635; (b) Finkelman, F. D.; Shearman, T.; Goldhill, J.; Sullivan, C. A.; Morris, S. C.; Madden, K. B.; Gause, W. C.; Urban, J. F. *Annu. Rev. Immunol.* **1997**, *15*, 505.
43. Stirling, R. G.; Chung, K. F. *Eur. Respir. J.* **2000**, *16*, 1158; (b) Renauld, J. C. *J. Clin. Pathol.* **2001**, *54*, 577.
44. (a) Gazzinelli, R. T.; Makino, M.; Chattopadhyay, S. K.; Snapper, C. M.; Sher, A.; Hugin, A. W.; Morise, H. C. III *J. Immunol.* **1992**, *148*, 182; (b) Romagnani, S. *Immunol. Today* **1990**, *11*, 316; (c) Secrist, H.; Chelen, C. J.; Wen, Y.; Marshall, J. D.; Umetsu, D. T. *J. Exp. Med.* **1993**, *178*, 2123.
45. Tuross, E.; Coates, C.; Shim, J.-Y.; Wang, Y.; Leslie, J. M.; Long, T. E.; Reddy, G. S. K.; Ortiz, A.; Culbreath, M.; Dickey, S.; Lim, D. V.; Alonso, E.; Gonzalez, J. *Bioorg. Med. Chem.* **2005**, *13*, 6289.
46. Sakamoto, Y.; Shiraishi, A.; Seonhee, J.; Nakata, T. *Tetrahedron Lett.* **1999**, *40*, 4203.
47. Hamerak, Z.; Ljubovic, E.; Mercep, M.; Mesic, M.; Sunjic V. *Synthesis* **2001**, 1989.
48. Carter, P. H.; LaPorte, J. R.; Scherle, P. A.; Decicco, C. P. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1237.
49. Milicevic, S.; Matovic, R.; Saicic, R. N. *Tetrahedron Lett.* **2004**, *45*, 955.
50. Paraskar, A. S.; Sudalai, A. *Tetrahedron* **2006**, *62*, 5756.
51. Narina, S. V.; Siva Kumar, T.; George, S.; Sudalai, A. *Tetrahedron Lett.* **2007**, *48*, 65.
52. Reddy, R. S.; Chouthaiwale, P. V.; Suryavanshi, G.; Chavan .V. B.; Sudalai. A. *Chem. Commun.* **2010**, *46*, 5012.
53. Sung-Gon Kim, S-G.; Park, T. H. *Tetrahedron: Asymmetry*, **2008**, *19*, 1626.
54. Qian, Y.; Xu, X.; Jiang, L.; Prajapati, D.; Hu, W. *J. Org. Chem.* **2010**, *75*, 7583.
55. Mishra, R. K.; Coates, C. M.; Revell, K. D.; Tuross, E. *Org. Lett.*, **2007**, *9*, 575.

CHAPTER 4

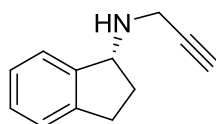
**Asymmetric Synthesis of Rasagiline and NHC-
catalyzed Esterification of Aromatic
Aldehydes**

Section I:

Enantioselective Synthesis of Rasagiline an anti-Parkinson Drug

4.1.1 Introduction and Pharmacology

Neurodegenerative diseases are the third cause of death among aged population all over the world. Parkinson's disease (PD) named after the English doctor James Parkinson is the most common neurodegenerative disorder and manifests as rigidity, resting tremor and posture instability.¹ PD is a degenerative disorder of the central nervous system, resulting from the death of cells that use dopamine to transmit their signals, resulting in decreased synaptic signal strength and concomitant symptomology.² It is generally characterized by insufficient formation and activity of dopamine produced within substantia nigra, a region of the midbrain. Modern treatments are effective at managing the early stages of the disease, mainly through the use of dopamine agonists.³ As the disease progresses and dopaminergic neurons continue to be lost, these drugs eventually become ineffective at treating the symptoms and at the same time produce a complication called dyskinesia, marked by involuntary writhing movements.⁴



Rasagiline (1)

Fig. 1: Structure of anti-Parkinson drug, rasagiline (1)

The existing therapies are still well-below the expectations for the advanced stages of the disease. Therefore, there is a need for novel therapeutic drug to ameliorate the

pathophysiological process of the disease. It is well-established that human cells contain two forms of monoamine oxidase (MAO) type A and type B. In the brain, MAO type B is far more prevalent and is responsible for the breakdown of dopamine after its release into the synapse.⁵ Since PD is generally regarded as a dopamine deficiency-disorder, there is a need for drugs, which can correct dopamine deficiency. In this regard, rasagiline (**1**) (**Fig. 1**) (Azilect®, Teva Pharmaceutical Industries Ltd., Israel), an irreversible MAO B inhibitor, was approved first in Israel (January 2005), followed by European agency (February 2005) and by US federal drug administration (May 2006) for the treatment of idiopathic PD as monotherapy or as adjunct therapy with L-dopa in advanced cases.⁶ Rasagiline (**1**) presents a very significant and selective activity concerning monoamine oxidase B inhibition.⁷ It is selective for MAO B over MAO A by a factor of fourteen.⁸ By inhibiting the breakdown of dopamine in the synapse, rasagiline (**1**) permits the signaling neurons to re-absorb more of dopamine.⁹ Structure activity studies show that its neuroprotective efficacy is strongly related to the propargylamino group.¹⁰ Very recently studies have revealed that rasagiline salts decrease human melanoma tumor growth *in vivo*, proving its potential as multi-target drug candidature for melanoma and PD.^{10e}

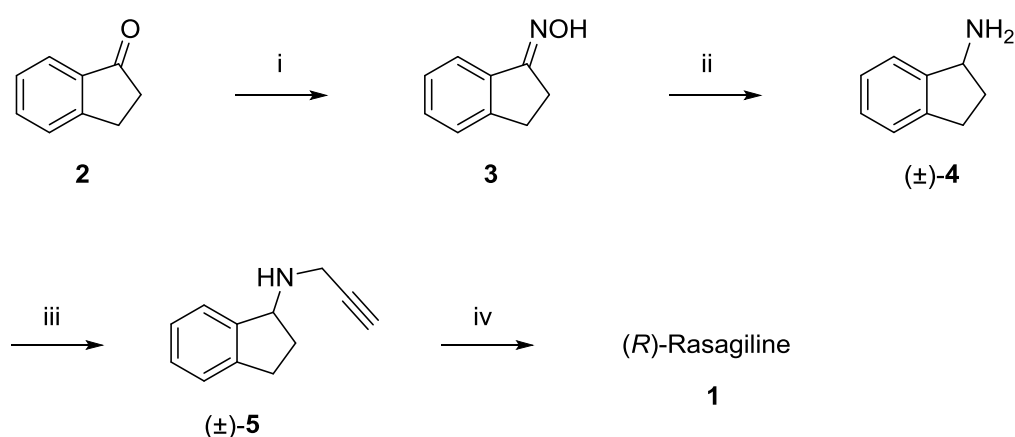
4.1.2 Review of Literature

Literature search revealed that there are only few reports available on the synthesis of rasagiline (**1**)¹¹⁻¹⁶ and its analogs.¹⁷ A short description on the reported synthesis of rasagiline (**1**) is presented below.

Moussa's approach (1991)^{11,12}

Moussa *et al.* have reported the synthesis of rasagiline (**1**) by using high-performance liquid chromatography for the separation of racemic mixture as the key technique.

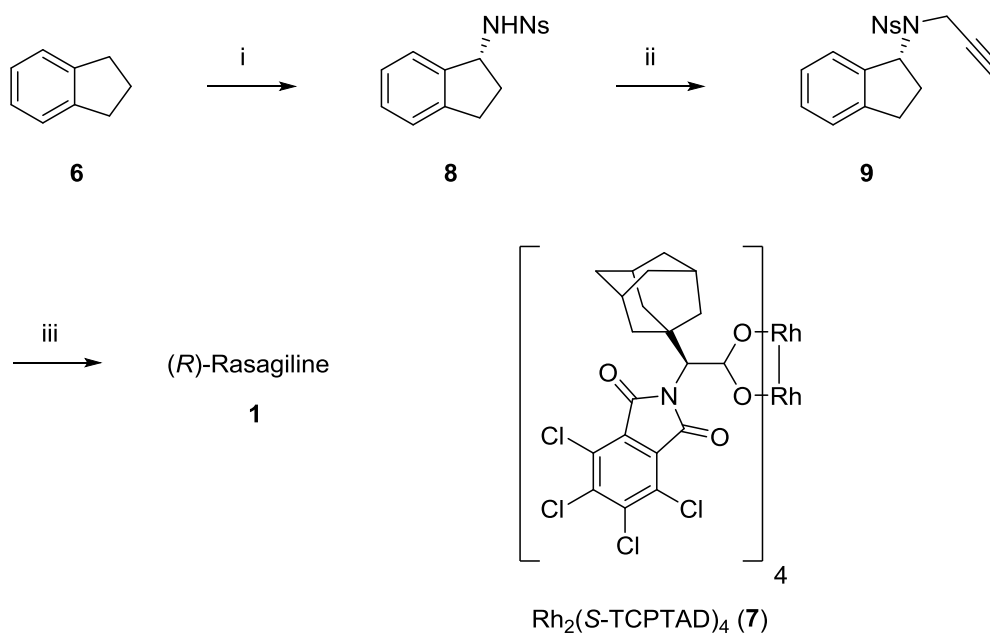
Thus, indanone **2** was treated with hydroxyl amine under basic condition to provide 1-indanone oxime **3**. Hydrogenation of oxime **3** using Raney Ni afforded racemic mixture of primary amine (\pm)-**4** in 87% yield. Propargylation of amine (\pm)-**4** gave racemic mixture of rasagiline (\pm)-**5**, from which (+)-rasagiline (**1**) was isolated by resolving the racemic mixture on a Chiracel OJ (cellulose tris[*p*-methylbenzoate]) preparative HPLC column (**Scheme 1**).



Scheme 1: (i) $\text{NH}_2\text{OH}\cdot\text{HCl}$, H_2O , 50% aq. NaOH , $\text{C}_2\text{H}_5\text{OH}$, reflux, 15 min, 98%; (ii) Raney Ni, H_2 (80 psi), 16% aq. NH_3 , CH_3OH , 25 °C, 25 h, 87%; (iii) K_2CO_3 , propargyl chloride, CH_3CN , 60 °C, 16 h, 56%; (iv) Chiral HPLC separation.

Davies' approach (2006)¹³

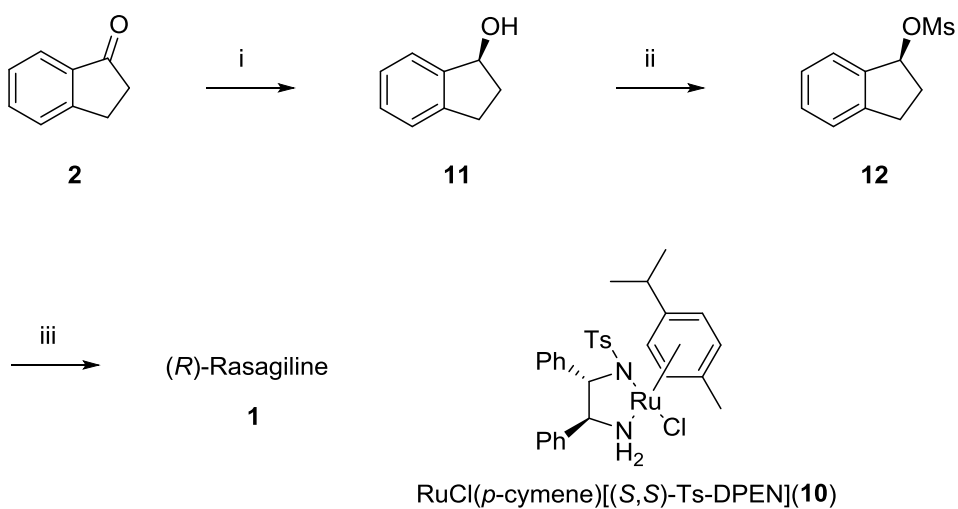
Davies *et al.* have achieved the synthesis of (+)-rasagiline (**1**) using catalytic enantioselective C-H amination as the key step. Thus, sulfonamide **8** was obtained *via* enantioselective amination of indane **6** in trifluorotoluene with newly developed catalyst *i.e.* $\text{Rh}_2(\text{S-TCPTAD})_4$ (**7**) (2 mol%), $\text{PhI}(\text{OAc})_2$, MgO and NsNH_2 as amine source in 95% yield and 94% ee. Alkylation of sulfonamide **8** with propargyl bromide forming nosyl protected rasagiline **9**, followed by the removal of nosyl group under Fukuyama's protocol, afforded rasagiline (**1**) (**Scheme 2**).



Scheme 2: (i) $\text{Rh}_2(\text{S-TCPTAD})_4$ (**7**), $\text{PhI}(\text{OAc})_2$, trifluorotoluene, NsNH_2 , MgO , 23 °C, 3 h, 95%; (ii) propargyl bromide, K_2CO_3 , CH_3CN , 60 °C, 16 h, 75%; (iii) $\text{HSCH}_2\text{CH}_2\text{OH}$, DBU, DMF, 25 °C, 1 h, 64%.

Boulton's approach (2009)¹⁴

Boulton *et al.* have developed a useful synthetic method for the synthesis of (+)-rasagiline (**1**) using chiral ketone reduction as the key step.

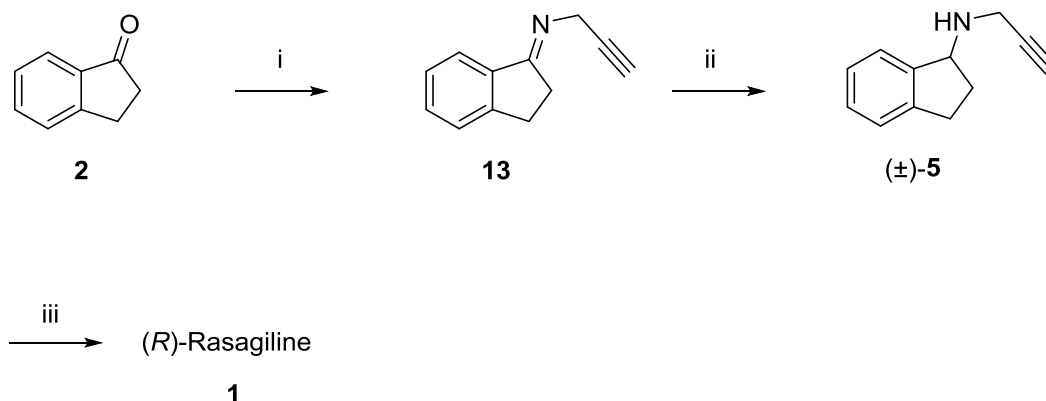


Scheme 3: (i) $\text{RuCl}(p\text{-cymene})[(\text{S},\text{S})\text{-Ts-DPEN}]$ (**10**), HCOOH , Et_3N , CH_2Cl_2 , 30 °C, 21 h, 96%; (ii) $(\text{CH}_3\text{SO}_2)\text{O}$, Et_3N , CH_2Cl_2 , -20 °C, 45 min; (iii) propargyl amine, CH_2Cl_2 , 25 °C, 12 h, 68% (over two steps).

Thus, chiral reduction of l-indanone **2** in the presence of an optically active Ru-catalyst *i.e.* RuCl(*p*-cymene)[(*S,S*)-Ts-DPEN](**10**) gave the corresponding (*S*)-indanol **11** in 96% yield with 98% ee. Mesylation of the hydroxyl moiety of indanol **11** provided mesylate **12**, which on subsequent benzylic SN₂ substitution with propargyl amine afforded rasagiline (**1**) (Scheme 3).

Praveen's approach (2010)¹⁵

Praveen *et al.* have reported the synthesis of rasagiline (**1**) by employing classical resolution, for the separation of racemic mixture as the key step. Indanone **2** was thus condensed with propargyl amine to afford indanylimine **13**. Reduction of imine **13** with NaBH₄ afforded racemic mixture of rasagiline (±)-**5**, from which (+)-rasagiline (**1**) was isolated by resolving the racemic mixture in the presence of tartaric acid as the chiral resolving agent (Scheme 4).



Scheme 4: (i) propargyl amine, CH₃OH, 30 °C, 20 h; (ii) NaBH₄, CH₃OH, 0 °C to 25 °C, 6 h, 47% (over two steps); (iii) (a) tartaric acid, ^tPrOH, 60 °C, 8 h, 77%; (b) NaOH, H₂O, 35 °C, 4 h, 41%.

Tatendra's approach (2011)¹⁶

Tatendra *et al.* have developed a useful synthetic method for the synthesis of rasagiline (**1**) commencing from chiral starting material. Reaction of (*S*)-indanol **11** with *p*-TsCl (**14**) in presence of triethylbenzylammonium chloride (TEBA) as PTC

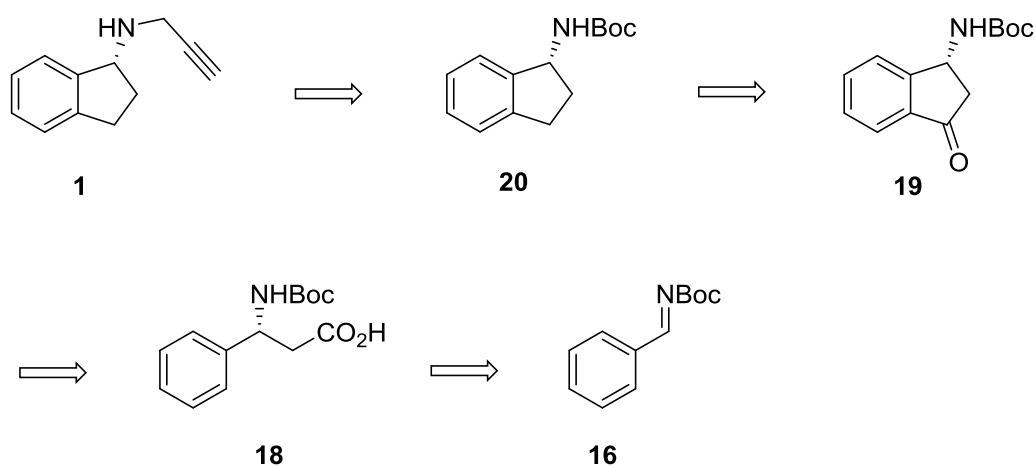
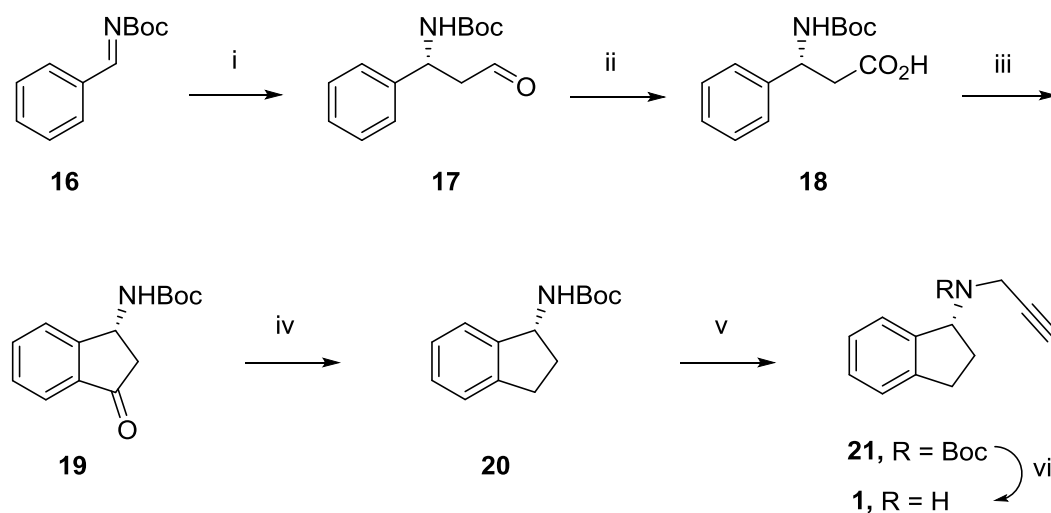


Fig. 2: Retrosynthetic analysis of rasagiline (1)

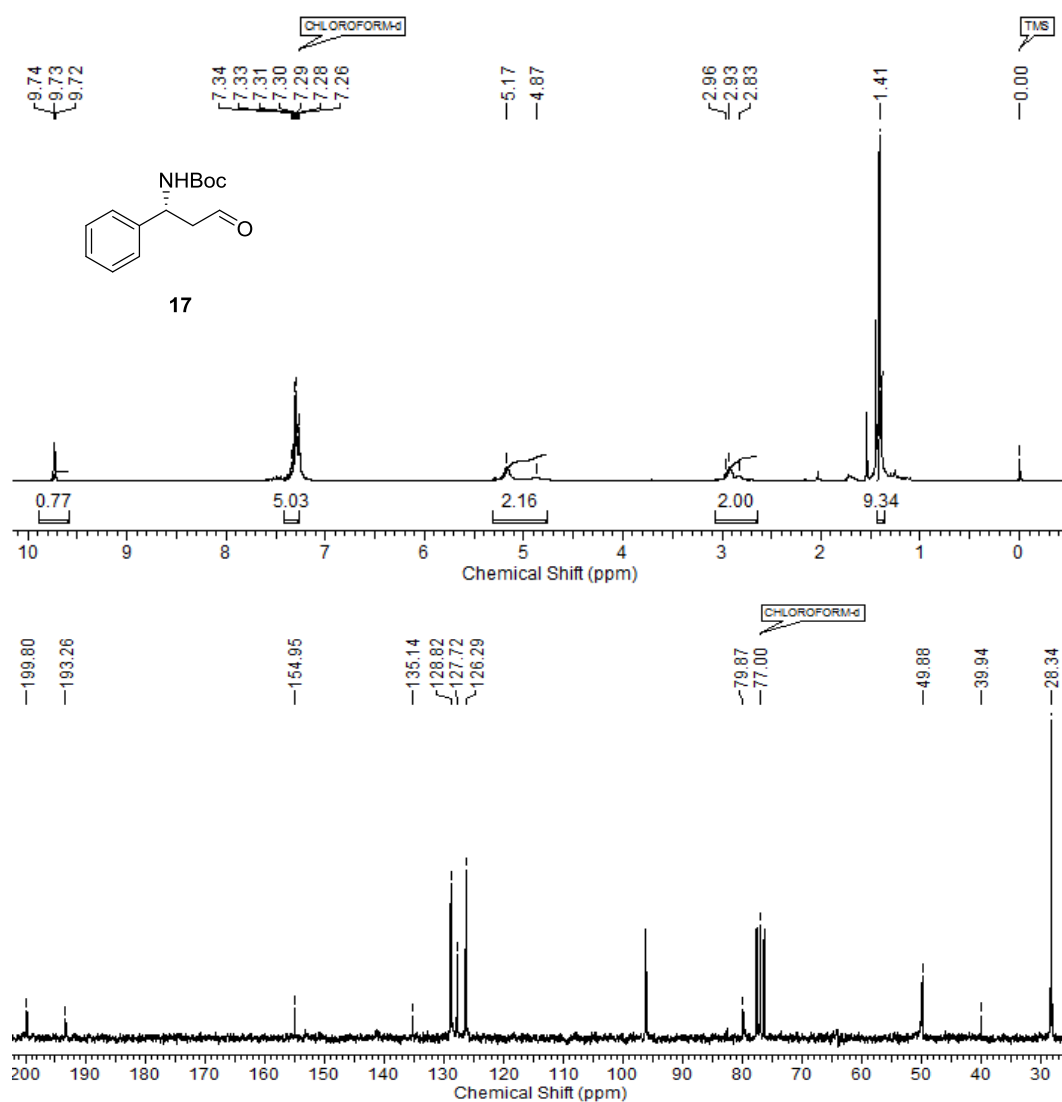
4.1.4 Results and Discussion

The complete synthetic sequence for rasagiline (1) is shown in **Scheme 6**, wherein proline-catalyzed Mannich reaction of acetaldehyde constitutes a key step for the introduction of chirality in the molecule.



Scheme 6: (i) CH_3CHO , D-proline (20 mol%), CH_3CN , 0 °C, 3 h, 62%; (ii) NaClO_2 , NaH_2PO_4 , *tert*-BuOH:H₂O (5:1), 25 °C, 30 min, 93%; (iii) ClSO_3H , CH_2Cl_2 , 25 °C, 2 h, then $(\text{Boc})_2\text{O}$, Et_3N , cat. DMAP, CH_2Cl_2 , 25 °C, 3 h, 83%; (iv) $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$, KOH, diethylene glycol, reflux, 4 h, 88%; (v) NaH, propargyl bromide, DMF, 25 °C, 4 h, 81%; (vi) 37% aq. HCl, dioxane, 25 °C, 30 min, 97%.

The present synthesis of rasagiline (**1**) started from Boc-protected benzaldimine **16**, which on Mannich reaction with acetaldehyde (D-proline, CH₃CN, 0 °C)¹⁸ afforded β -aminoaldehyde **17** in 62% yield and 98% ee. The formation of β -aminoaldehyde **17** was confirmed from its ¹H NMR spectrum, which showed typical proton signals at δ 9.73 (t, J = 1.7 Hz, 1H) and δ 2.83-2.96 (m, 2H) corresponding to aldehydic and homo benzylic methylene protons respectively. This was further ascertained by the presence of two characteristic carbon signals at δ 199.8 and 193.3 in its ¹³C NMR spectrum corresponding to carbamate and aldehydic carbonyl carbons respectively (**Fig. 3**).



The Pinnick oxidation of β -aminoaldehyde **17** with NaClO_2 and NaH_2PO_4 gave the corresponding β -aminocarboxylic acid **18** in 93% yield.¹⁹ The formation of carboxylic acid **18** was confirmed from its ^1H NMR spectrum, which showed two characteristic proton signals at δ 2.67 (br s, 1H) and 2.86 (br s, 1H) corresponding to homo benzylic methylene protons, while its ^{13}C NMR spectrum showed a typical carbon signal at δ 170.1 due to the presence of carboxylic acid carbonyl carbon (**Fig. 4**). Its IR spectrum displayed a characteristic strong absorption band at 1707 cm^{-1} indicating the presence of carboxylic acid group.

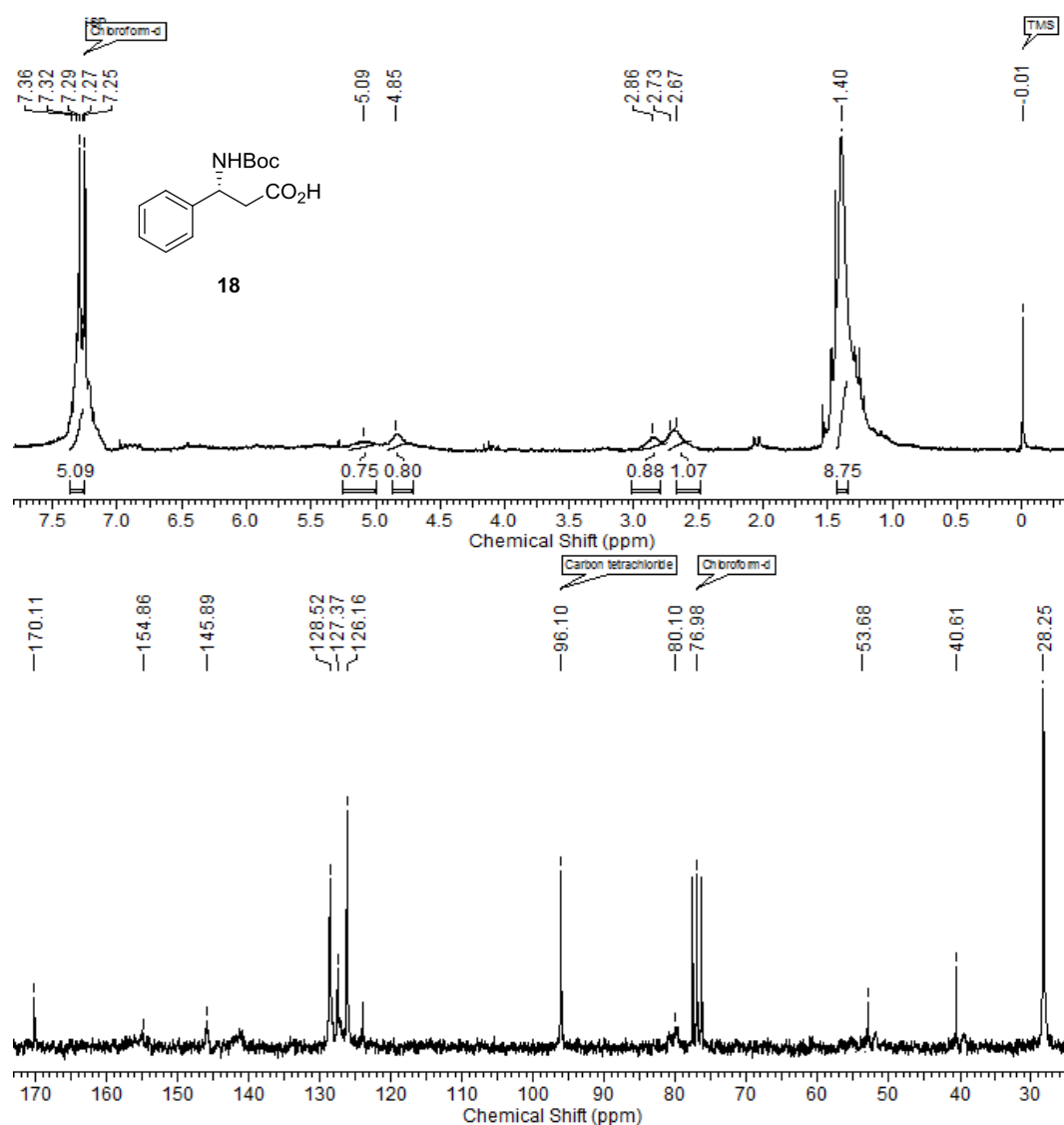


Fig. 4: ^1H and ^{13}C NMR spectra of carboxylic acid **18**

The indanone **19** was obtained by the Friedel-Crafts' intramolecular acylation²⁰ of β -amino acid **18** on reaction with ClSO_3H , followed by the protection of primary amine $[(\text{Boc})_2\text{O}, \text{Et}_3\text{N}, \text{cat. DMAP}]$. Indanone **19** was confirmed from its ^1H NMR spectrum, which showed two characteristic doublet of doublets at δ 2.45 (dd, $J = 3.2, 19.2$ Hz, 1H) and 3.16 (dd, $J = 7.6, 19.2$ Hz, 1H), corresponding to the diastereotopic methylene protons, while its ^{13}C NMR spectrum showed a characteristic carbon signal at δ 202.9 due to ketone carbonyl carbon (**Fig. 5**).

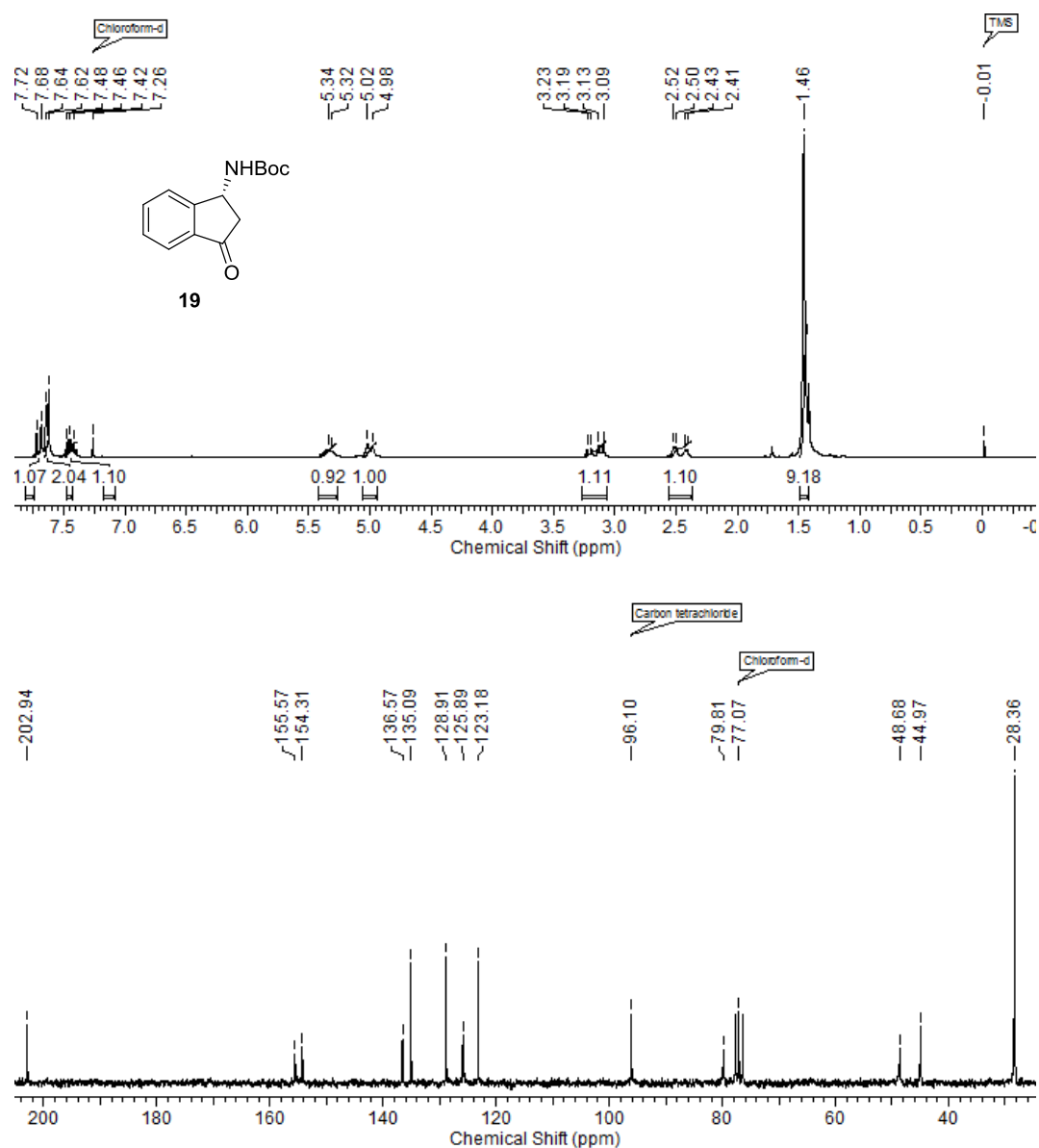


Fig. 5: ^1H and ^{13}C NMR spectra of indanone **19**

The formation of indanone **19** was also confirmed from its IR spectrum, which displayed a strong absorption band at 1712 cm^{-1} indicating the presence of ketone functional group. The optical purity of **19** was determined to be 98% ee from chiral HPLC analysis (Chiralcel AD-H, *n*-hexane/ *i*PrOH, 90:10, 0.5 mL/min) retention time 14.36 (99.08%) and 19.88 (0.92%) (**Fig.6**).

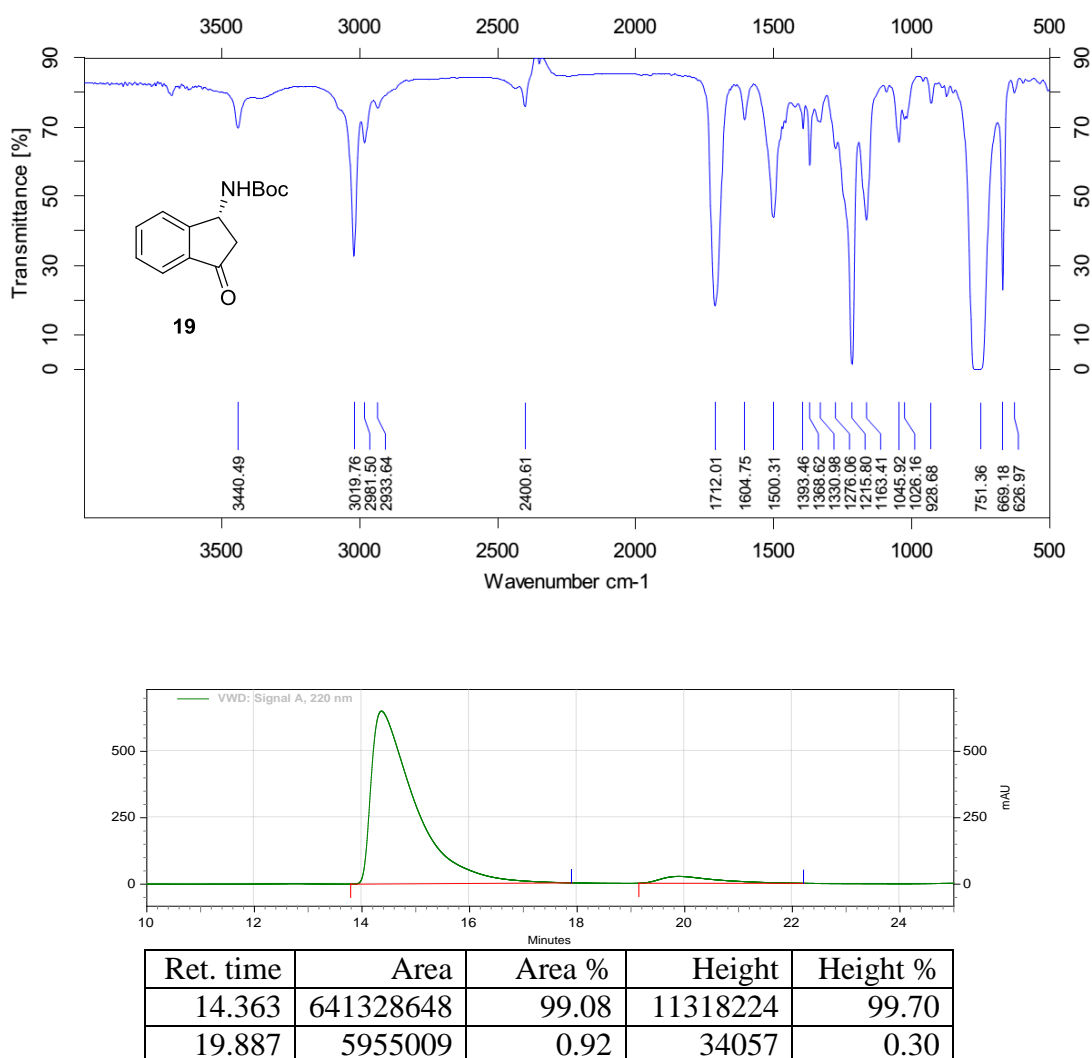


Fig. 6: IR spectrum and HPLC chromatogram of indanone **19**

Wolff-Kishner reduction of indanone **19** with $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$ and KOH afforded carbamate **20** in 88% yield.²¹ The formation of carbamate **20** was established from its ^1H NMR spectrum, which showed two characteristic proton signals at δ 2.74-3.03 (m, 2H) and 5.17 (d, $J = 7.7$ Hz, 1H) due to benzylic protons, while its ^{13}C NMR spectrum displayed the disappearance of carbon signal at δ 202.9 corresponding to the ketone carbonyl carbon, confirming the reduction of ketone group (**Fig.7**).

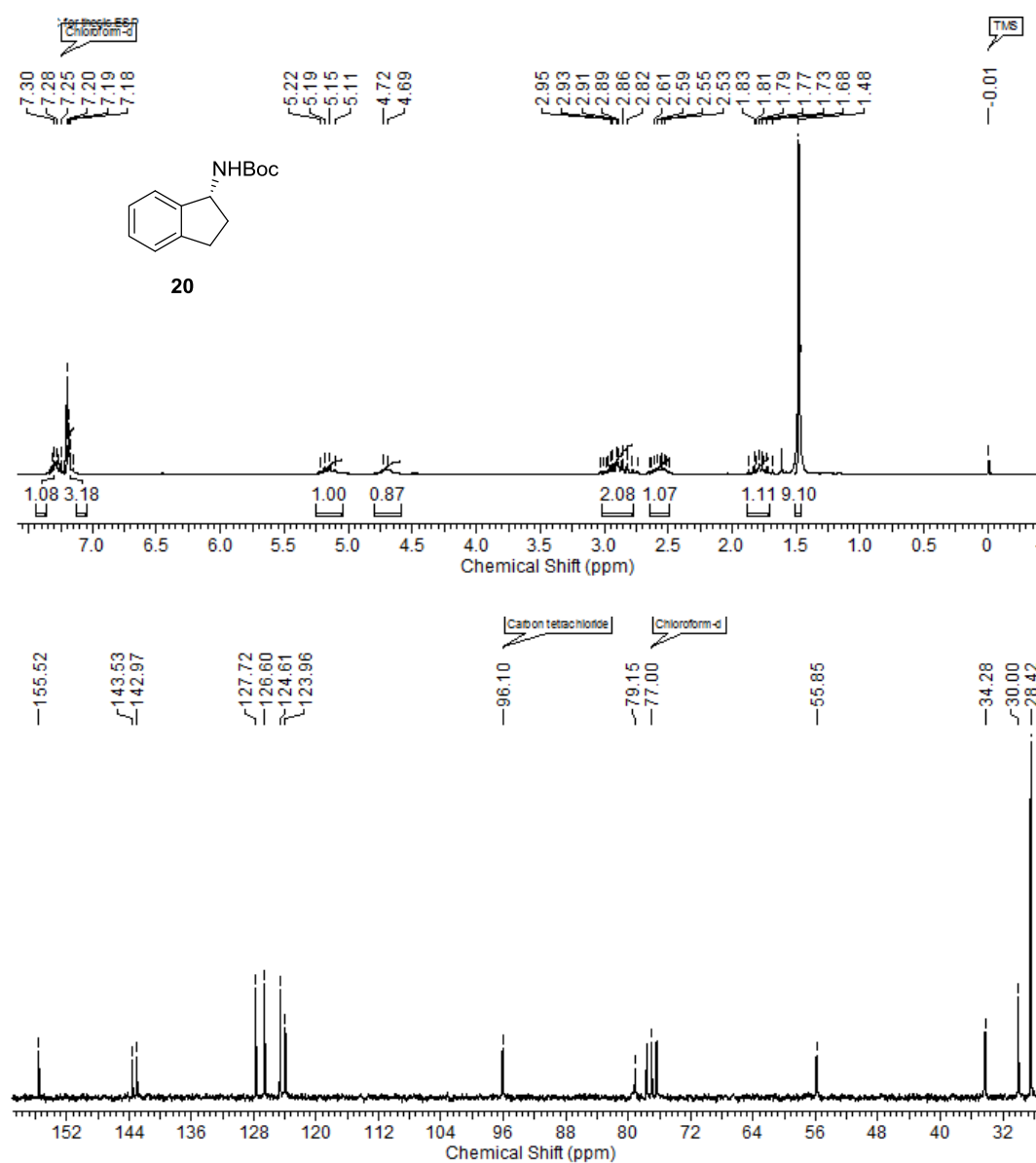


Fig. 7: ^1H and ^{13}C NMR spectra of carbamate **20**

Alkylation of carbamate **20** was achieved with propargyl bromide under basic conditions to afford the protected rasagiline **21** in 81% yield.²² The formation of protected rasagiline **21** was confirmed from its ¹H NMR spectrum, which showed three typical proton signals at δ 2.96-3.10 (m, 1H), 3.32-3.63 (m, 1H) and 3.82-4.16 (m, 1H) corresponding to terminal alkyne and diastereotopic methylene protons attached to alkyne respectively. Its ¹³C NMR spectrum showed two characteristic carbon signals at δ 61.1 and 69.6 corresponding to alkyne carbons (**Fig. 8**).

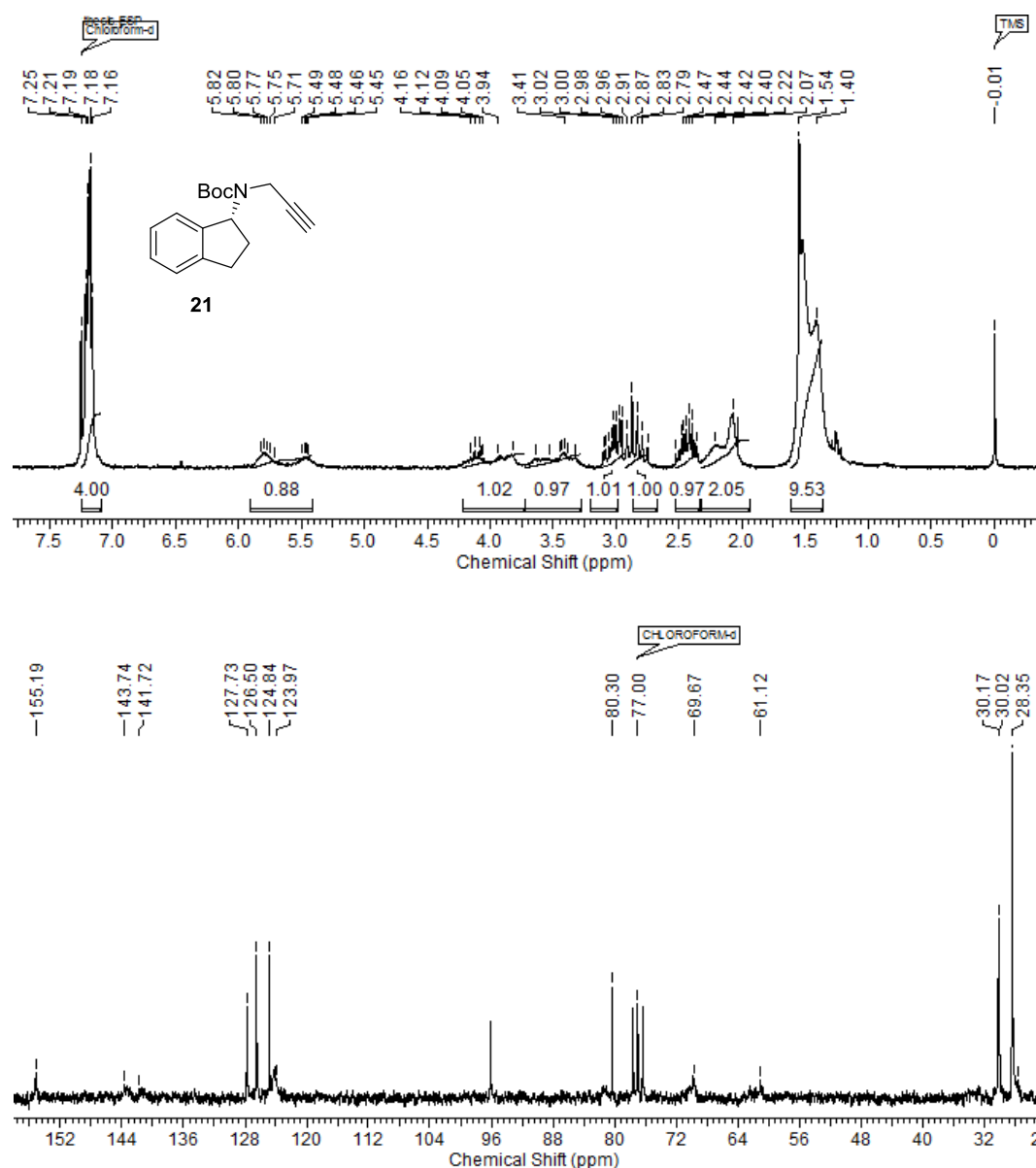


Fig. 8: ¹H and ¹³C NMR spectra of protected rasagiline **21**

Finally, deprotection of the Boc protecting group with 37% aq. HCl in dioxane furnished rasagiline (**1**) in 97% yield and 98% ee, $[\alpha]_{25}^D +19.4$ (c 0.5, CHCl_3) [lit.¹³ $[\alpha]_{25}^D +18.8$ (c 1.7, CHCl_3)]. The ^1H NMR spectrum of rasagiline (**1**) showed two typical proton signals at δ 2.23 (t, $J = 2.4$ Hz, 1H) and 3.52 (t, $J = 2.4$ Hz, 2H) corresponding to benzylic methine and diastereotopic methylene protons attached to alkyne respectively.

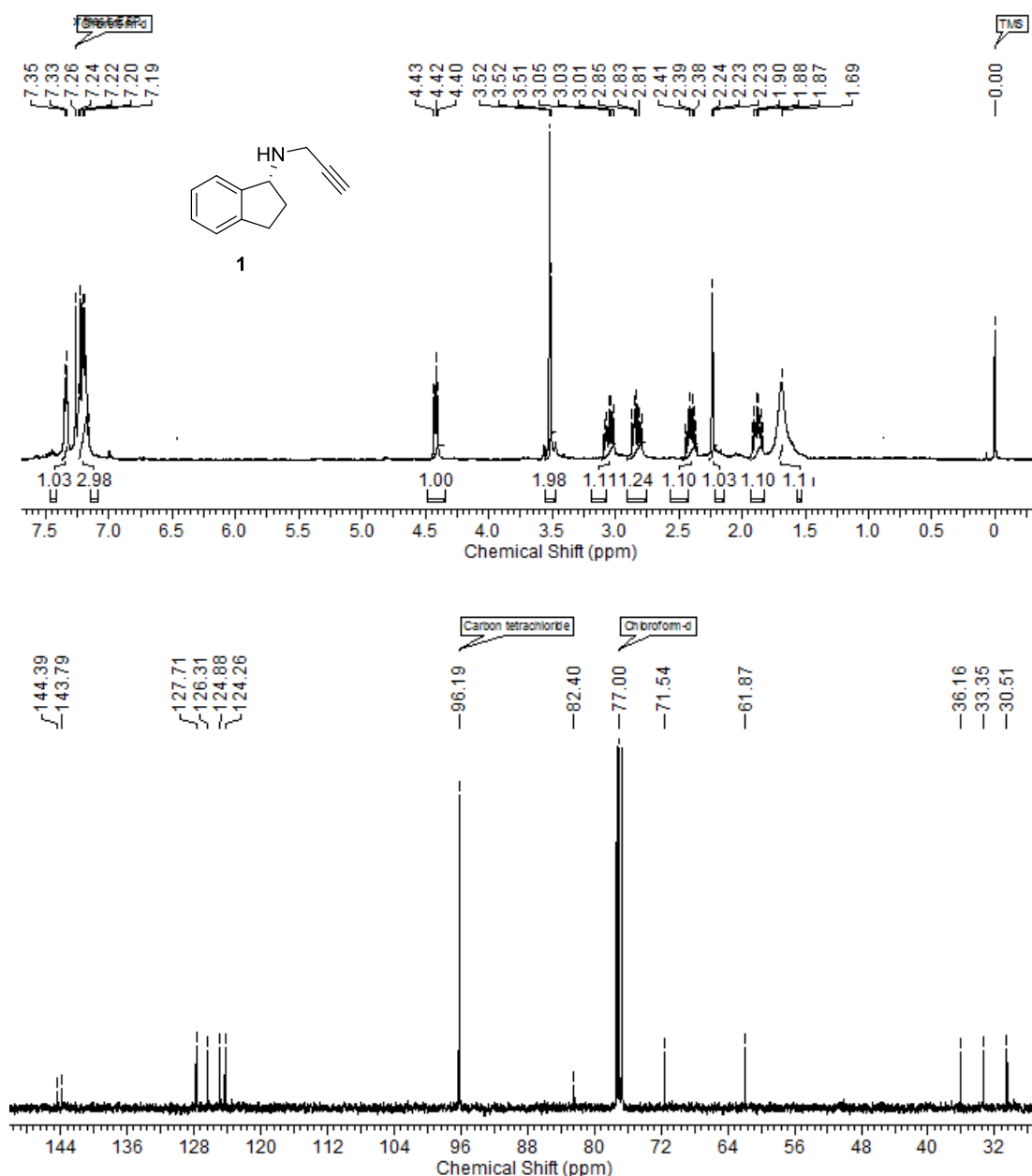


Fig. 9: ^1H and ^{13}C NMR spectra of rasagiline (**1**)

Its ^{13}C NMR spectrum showed characteristic carbon signals at δ 61.8, 71.5 and 82.4 corresponding to benzylic methine and alkyne carbons respectively (**Fig. 9**). The spectral data of rasagiline (**1**) were in complete agreement with the reported values.²²

4.1.5 Conclusion

In conclusion, we have achieved the asymmetric synthesis of rasagiline (**1**) (33% overall yield, 98% ee) using proline-catalyzed Mannich reaction, Friedel-Crafts acylation and Wolff-Kishner reduction as the key steps. The utilization of organocatalyst for introduction of chirality into the molecule and the high enantiomeric excess obtained in this method render our approach a good alternative to the known methods.

4.1.6 Experimental Section

(R)-tert-Butyl (3-oxo-1-phenylpropyl)carbamate (17)

To a stirred solution of benzaldehyde-derived N-Boc-imine **16** (2.870 g, 14 mmol) and redistilled acetaldehyde (3.9 mL, 70 mmol) in CH_3CN (150 mL) at 0 °C was added D-proline (0.320 g, 20 mol%) and the mixture was stirred further at 0 °C for 3 h. After the completion of reaction (monitored by TLC), it was quenched with water and extracted with Et_2O (3 x 100 mL). The combined organic layers were washed with brine, dried over anhyd. Na_2SO_4 , filtered and concentrated under reduced pressure to give the crude aldehyde, which was purified by column chromatography over silica gel using pet. ether:EtOAc (85:15) as eluent, to afford β -amino aldehyde **17** (2.16 g).

Yield: 62%; yellow solid; **mp:** 90-93 °C, (lit.²³ **mp:** 92-93.5 °C); $[\alpha]_{25}^{\text{D}} +29.6$ (*c* 1.7, CHCl_3); lit.²³ $[\alpha]_{25}^{\text{D}} +29.0$ (*c* 1.4, CHCl_3); **IR** (CHCl_3 , cm^{-1}): ν_{max} 720, 1024, 1052,

1258, 1371, 1394, 1479, 1513, 1692, 2979, 3346; **¹H NMR** (200 MHz, CDCl₃): δ 1.41 (s, 9H), 2.83-2.96 (m, 2H), 4.87 (br s, 1H), 5.17 (br s, 1H), 7.26-7.34 (m, 5H), 9.73 (t, *J* = 1.7 Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.3, 39.9, 49.9, 79.9, 126.3, 127.7, 128.8, 135.2, 155.0, 193.3, 199.8; **Analysis:** C₁₄H₁₉NO₃ requires C, 67.45; H, 7.68; N, 5.62; found: C, 67.26; H, 7.53; N, 5.49%.

(*R*)-3-((*tert*-Butoxycarbonyl)amino)-3-phenylpropanoic acid (18)

NaH₂PO₄ (3.37 g, 24.4 mmol) and NaClO₂ (1.502 g, 24.4 mmol) were added to a solution of β-amino aldehyde **17** (2.02 g, 8.14 mmol) in *tert*-BuOH:H₂O (45:9 mL) at 25 °C and the mixture allowed to stir for 30 min. After TLC showed the complete disappearance of starting material, it was diluted with EtOAc (100 mL) and dried over anhyd. Na₂SO₄ and concentrated to give the crude acid, which was purified by column chromatography over silica gel using pet. ether:EtOAc (60:40) as eluent, to afford carboxylic acid **18** (2.01 g).

Yield: 93%; colorless solid; **mp:** 126-129 °C; [α]₂₅^D +40.1 (*c* 1.2, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{max} 758, 1029, 1046, 1215, 1420, 1507, 1707, 2400, 2980, 3019; **¹H NMR** (200 MHz, CDCl₃): δ 1.40 (s, 9H), 2.67 (br s, 1H), 2.86 (br s, 1H), 4.85 (br s, 1H), 5.09 (br s, 1H), 7.27-7.36 (m, 5H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.2, 40.6, 53.7, 80.1, 126.2, 127.4, 128.5, 145.9, 154.8, 170.1; **Analysis:** C₁₄H₁₉NO₄ requires C, 63.38; H, 7.22; N, 5.28; found: C, 63.16; H, 7.11; N, 5.21%.

(*R*)-*tert*-Butyl (3-oxo-2,3-dihydro-1*H*-indenyl)carbamate (19)

To a solution of carboxylic acid **18** (2 g, 7.5 mmol) in dry CH₂Cl₂ (40 mL) was added ClSO₃H (2.5 mL, 37.5 mmol). The reaction mixture was stirred at 25 °C for 2 h, and quenched with saturated solution of NaHCO₃, extracted with Et₂O (3 × 50 mL). The combined organic phases were dried over anhyd. Na₂SO₄ and concentrated under

reduced pressure to afford the crude product, which was directly used for the next step without purification.

To a stirred solution of crude amine in CH_2Cl_2 (40 mL) and Et_3N (3.2 mL, 22.5 mmol) was added catalytic amount of DMAP (0.091 g, 0.75 mmol). After stirring for 5 min at 0 °C, $(\text{Boc})_2\text{O}$ (3.27 g, 15.0 mmol) was added drop-wise and the reaction mixture was allowed to stir for another 3 h. After the completion of the reaction, it was extracted with Et_2O (3×50 mL), washed with water, brine and dried over anhyd. Na_2SO_4 and concentrated to give the crude ketone, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (85:15) to furnish indanone **19** (1.54 g).

Yield: 83%; colorless solid; **mp:** 104-106 °C; $[\alpha]_{25}^{\text{D}}$ -14.6 (c 0.8, CHCl_3); 98% ee from chiral HPLC analysis (Chiralcel AD-H, *n*-hexane/ *i*PrOH, 90:10, 0.5 mL/min) retention time 14.36 (99.08%) and 19.88 (0.92%); **IR** (CHCl_3 , cm^{-1}): ν_{max} 751, 1045, 1163, 1276, 1393, 1500, 1604, 1712, 2400, 2933, 2981, 2981, 3019, 3440; **¹H NMR** (200 MHz, CDCl_3): δ 1.46 (s, 9H), 2.45 (dd, $J = 3.2, 19.2$ Hz, 1H), 3.16 (dd, $J = 7.6, 19.2$ Hz, 1H), 5.00 (d, $J = 8.4$ Hz, 1H), 5.34 (br s, 1H), 7.41-7.48 (m, 1H), 7.63 (d, $J = 3.8$ Hz, 2H), 7.70 (d, $J = 7.6$ Hz, 1H); **¹³C NMR** (50 MHz, CDCl_3): δ 28.4, 44.9, 48.7, 79.8, 123.2, 125.9, 128.9, 135.1, 136.6, 154.3, 155.6, 202.9; **Analysis:** $\text{C}_{14}\text{H}_{17}\text{NO}_3$ requires C, 68.00; H, 6.93; N, 5.66; found: C, 67.94; H, 6.73; N, 5.46%.

(*R*)-tert-Butyl (2,3-dihydro-1H-indenyl)carbamate (20)

To a stirred solution of indanone **19** (1.632 g, 6.6 mmol) and $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$ (1.668 g, 33 mmol) in diethylene glycol (40 mL) was added KOH pellets (1.847 g, 33 mmol). The resulting solution was refluxed for 4 h. After TLC showed the complete disappearance of starting material, the reaction mixture was diluted with H_2O , extracted with EtOAc (3×50 mL) and dried over anhyd. Na_2SO_4 and concentrated to

give the crude carbamate, which was purified by column chromatography over silica gel using pet. ether:EtOAc (85:15) as eluent, to afford carbamate **20** (1.355 g).

Yield: 88%; colorless gum; $[\alpha]_{25}^D +24.3$ (*c* 1, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 768, 849, 928, 1024, 1050, 1168, 1215, 1421, 1505, 1706, 2400, 2979, 3019; **¹H NMR** (200 MHz, CDCl₃): δ 1.48 (s, 9H), 1.68-1.87 (m, 1H), 2.49-2.65 (m, 1H), 2.74-3.03 (m, 2H), 4.70 (d, *J* = 7.3 Hz, 1H), 5.17 (d, *J* = 7.7 Hz, 1H), 7.15-7.20 (m, 3H), 7.27-7.32 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 28.4, 30.0, 34.3, 55.1, 79.1, 123.9, 124.6, 126.6, 127.7, 142.9, 143.5, 155.5; **Analysis:** C₁₄H₁₉NO₂ requires C, 72.07; H, 8.21; N, 6.00; found: C, 72.01; H, 8.14; N, 5.87%.

(*R*)-tert-Butyl (2,3-dihydro-1H-indenyl)(prop-2-yn-1-yl)carbamate (21)

To a stirred solution of carbamate **21** (0.500 g, 2.14 mmol) and NaH (0.098 g, 60% w/w, 2.35 mmol) in dry DMF (5 mL) was added propargyl bromide (80% solution in toluene, 286 μ L, 2.57 mmol) at 25 °C and the mixture was stirred under nitrogen atmosphere for 4 h. The reaction was quenched with H₂O and the resulting mixture was extracted with EtOAc (3 x 10 mL), washed with saturated aq. NH₄Cl (5 mL) and brine solution (5 mL). The organic layers were separated, dried over anhyd. Na₂SO₄, and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (95:05) as an eluent to give protected rasagiline **21** (0.470 g).

Yield: 81%; yellow oil; $[\alpha]_{25}^D -14.4$ (*c* 0.7, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{\max} 669, 761, 929, 1159, 1215, 1330, 1405, 1439, 1456, 1477, 1521, 1687, 2126, 2400, 2935, 2980, 3019, 3308; **¹H NMR** (200 MHz, CDCl₃): δ 1.50 (br s, 9H), 2.07 (br s, 1H), 2.22 (br s, 1H), 2.36-2.52 (m, 1H), 2.75-2.91 (m, 1H), 2.96-3.10 (m, 1H); 3.32-3.63 (m, 1H), 3.82-4.16 (m, 1H), 5.45-5.82 (m, 1H), 7.16-7.21 (m, 4H); **¹³C NMR** (50 MHz, CDCl₃): δ 27.6, 28.3, 30.0, 30.1, 61.1, 69.6, 80.3, 123.9, 124.8, 126.5, 127.7, 141.7,

143.7, 155.2; **Analysis:** C₁₇H₂₁NO₂ requires C, 75.25; H, 7.80; N, 5.26; found: C, 75.03; H, 7.56; N, 5.14%.

(R)-Indanyl-prop-2-ynyl-amine [rasagiline] (1)

To a solution of protected rasagiline **21** (0.300 g, 1.1 mmol) in dioxane (5 mL) was added 37% aq. HCl in H₂O (0.5 mL) and the mixture stirred for 30 min at 25 °C. The reaction mixture was quenched with sat. NaHCO₃. Dioxane was evaporated and the residue was extracted with EtOAc (3 × 10 mL) and the combined organic phases were dried over anhyd. Na₂SO₄ and concentrated to give the crude product, which was then purified by column chromatography over silica gel using pet. ether:EtOAc (70:30) to give rasagiline **1** (0.182 g).

Yield: 97%; yellow solid; **mp:** 146-149 °C, (lit.¹³ **mp:** 148 °C); [α]₂₅^D +19.4 (*c* 0.5, CHCl₃); lit.¹³ [α]₂₅^D +18.8 (*c* 1.7, CHCl₃); **IR** (CHCl₃, cm⁻¹): ν_{max} 649, 1088, 1161, 1215, 1349, 1456, 2400, 2848, 2929, 3281; **¹H NMR** (400 MHz, CDCl₃): δ 1.69 (br s, 1H), 1.84-1.92 (m, 1H), 2.23 (t, *J* = 2.4 Hz, 1H), 2.36-2.45 (m, 1H), 2.79-2.87 (m, 1H), 3.01-3.09 (m, 1H); 3.52 (t, *J* = 2.4 Hz, 2H), 4.42 (t, *J* = 6.2 Hz, 1H), 7.16-7.24 (m, 3H), 7.34 (d, *J* = 5.9 Hz, 1H); **¹³C NMR** (100 MHz, CDCl₃): δ 30.5, 33.3, 36.1, 61.8, 71.5, 82.4, 124.3, 124.9, 126.3, 127.7, 143.8, 144.4; **Analysis:** C₁₂H₁₃N requires C, 84.17; H, 7.65; N, 8.18; found: C, 84.13; H, 7.57; N, 8.12%.

Section II:

***N*-Heterocyclic Carbene-Catalyzed Esterification of Aromatic Aldehydes with Alcohols under Aerobic Condition**

4.2.1 Introduction

The direct transformation of aldehydes to the corresponding esters²⁴ with alcohols under mild conditions is often required in organic synthesis, especially in the synthesis of natural products.²⁵ For example: Lee *et al.* have utilized intramolecular oxidative esterification of ω -hydroxy aldehyde **22** as the key step in their synthesis of cytotoxic (+)-dactylolide (**24**) (Fig. 10).^{25e}

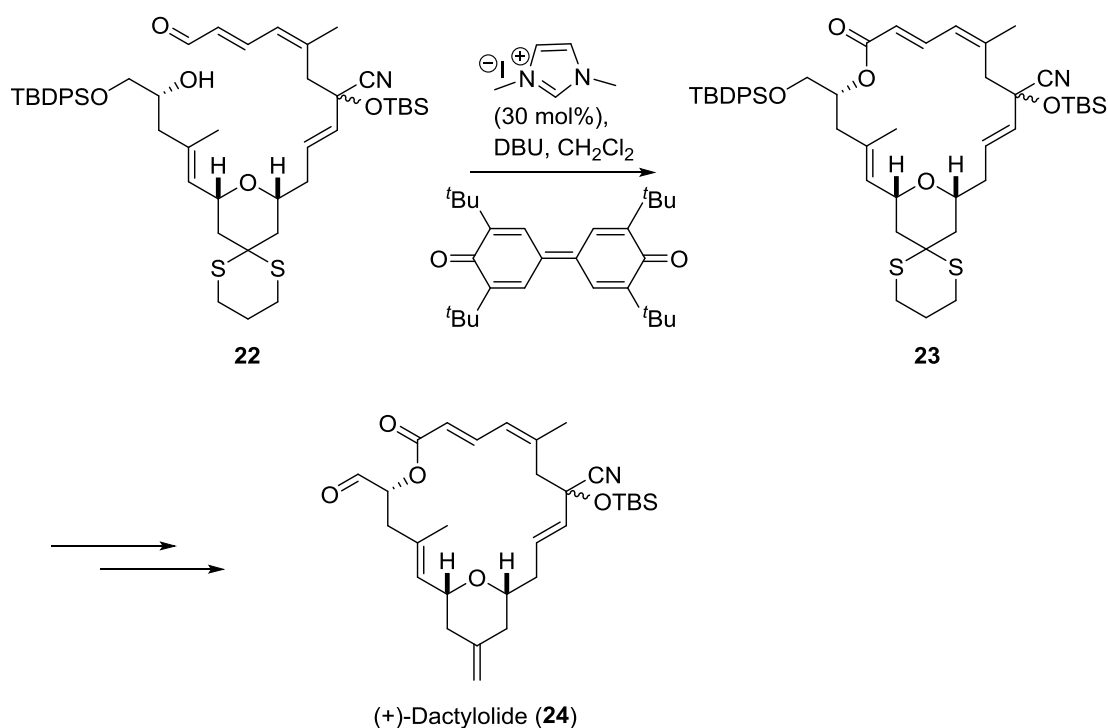


Fig. 10: Oxidative esterification of aldehyde in the synthesis of (+)-dactylolide (**24**)

Further esterification processes are widespread in the industrial synthesis of a variety of end-products such as fragrances, monomers, plasticizers, etc, many of which are

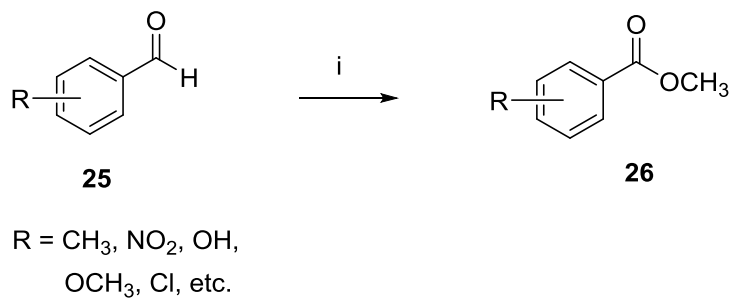
classified as high production volume (HPV) chemicals. In addition, applications to lower volume, high-value pharmaceutical and fine chemicals targets are prominent, and often require more stringent coupling protocols to achieve the desired chemo- and stereoselectivity. Aromatic esters are also important and useful structural elements finding tremendous applications in wide range of fields encompassing solvents, lubricants, plasticizing agents, perfumes, pharmaceuticals, agrochemicals, etc.²⁶ The conventional methods for the synthesis of carboxylic esters from aldehydes involve oxidation of aldehydes to carboxylic acids followed by esterification with alcohols catalyzed by acids. In contrast, the direct method of conversion of aldehydes to carboxylic esters holds considerable promise in organic synthesis as it minimizes the number of steps.

4.2.2 Review of Literature

Literature search revealed that there are several methods available for the direct transformation of aldehydes into the corresponding esters. Direct transformation of aldehydes into esters has been achieved using a variety of reagents such as V_2O_5/H_2O_2 ,²⁷ oxone[®],²⁸ pyridinium hydrobromide perbromide,²⁹ acetone cyanohydrins,³⁰ $(NaIO_4)/LiBr$,³¹ I_2 ,³² TBHP,³³ and electrochemical methods.³⁴ Recently, *N*-Heterocyclic carbenes (NHCs)-catalyzed oxidative esterification of aldehydes with alcohols,³⁵ alkyl halides³⁶ and boronic acids³⁷ has also been reported. Some of the recent developments on this transformation are discussed below.

Gopinath's approach (2000)²⁷

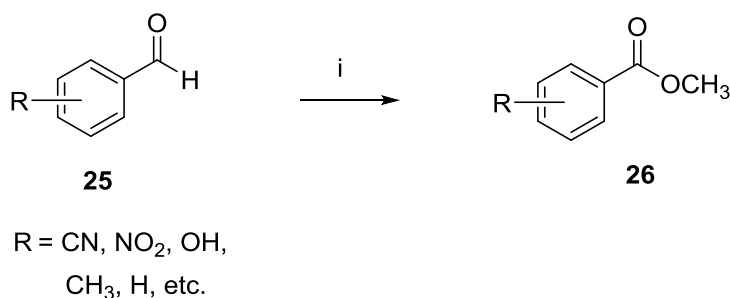
In Gopinath's approach, aldehydes **25**, in the presence of methanol, undergo oxidative transformation to the corresponding esters **26** upon treatment with catalytic amounts of V_2O_5 in combination with 30% aq. H_2O_2 as oxidant (**Scheme 7**).



Scheme 7: (i) V₂O₅ (cat.), 30% aq. H₂O₂, CH₃OH, 80 °C, 0.5-6 h, 83-100%.

Traivs' approach (2003)²⁸

Travis *et al.* have developed a highly efficient, mild, and simple protocol for the oxidation of aldehydes **25** to the corresponding carboxylic acids utilizing oxone as the sole oxidant. Direct conversion of aldehydes **25** in alcoholic solvents to their corresponding ester products **26** has also been reported. These reactions may prove to be valuable alternatives to traditional metal-mediated oxidations, however, it uses more than stoichiometric amounts of oxone (**Scheme 8**).

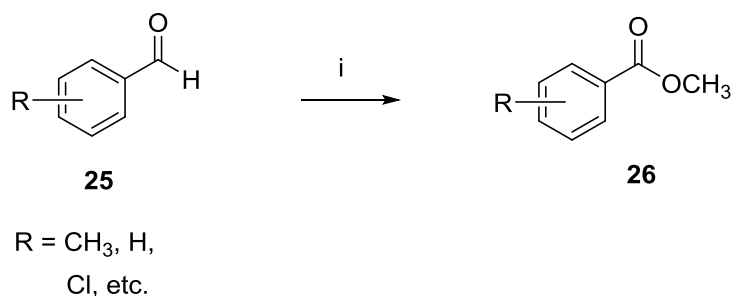


Scheme 8: (i) Oxone, CH₃OH, 18 h, 25 °C, 9-98%.

Onami's approach (2004)²⁹

In this approach, the direct esterification of aldehydes with alcohols was carried out with pyridinium hydrobromide perbromide (PHPB) in water at 25 °C. A variety of aldehydes **25** were converted to their corresponding esters **26**. Further, a variety of

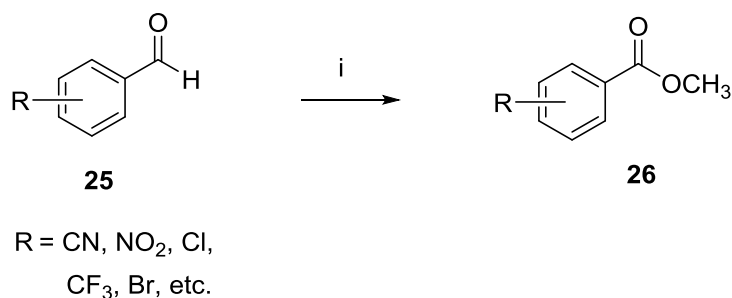
aliphatic alcohols were also converted to the corresponding Tishchenko-like dimeric esters in good yields under the same reaction conditions (**Scheme 9**).



Scheme 9: PHPB, CH₃OH, H₂O, 25 °C, 40-87 h, 70-94%.

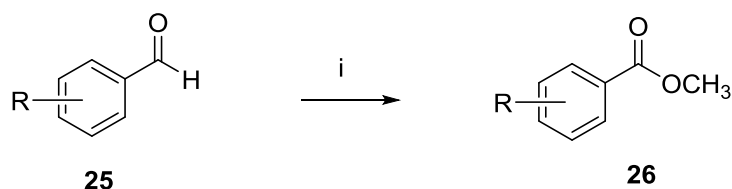
Sudalai's approach (2005), (2007)^{30, 31}

Sudalai *et al.* have described a simple procedure for the conversion of electron-deficient aldehydes **25** into the corresponding esters **26** on reaction with methanol in excellent yields mediated by acetone cyanohydrin and base (**Scheme 10**).



Scheme 10: (i) acetone cyanohydrin (5 mmol), Et₃N, CH₃OH, 25 °C, 2 h, 60-92%.

In yet another approach, these authors have converted aromatic aldehydes **25** directly to the corresponding aromatic esters **26** in high yields on treatment with CH₃OH using sodium metaperiodate (NaIO₄)/LiBr as oxidant under acidic medium (**Scheme 11**).

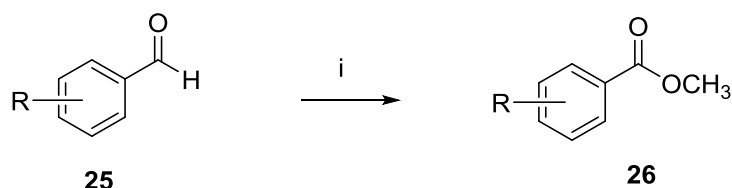


R = CN, NO₂, OH,
CH₃, H, etc.

Scheme 11: (i) LiBr, NaIO₄, conc. H₂SO₄, CH₃OH, 25 °C, 18 h, 78-98%.

Budhewar's approach (2006)³²

Budhewar *et al.* have developed a simple and mild procedure for the facile, direct oxidative methyl esterification of aldehydes **25** using molecular I₂ in combination with PhI(OAc)₂ in methanol (**Scheme 12**).

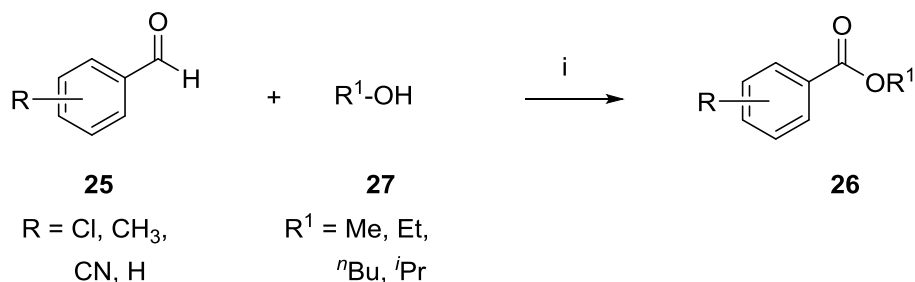


R = CH₃, NO₂, OH,
OCH₃, Cl, etc.

Scheme 12: (i) I₂, PhI(OAc)₂, CH₃OH, 25 °C, 10-14h, 67-89%.

Li's approach (2007)³³

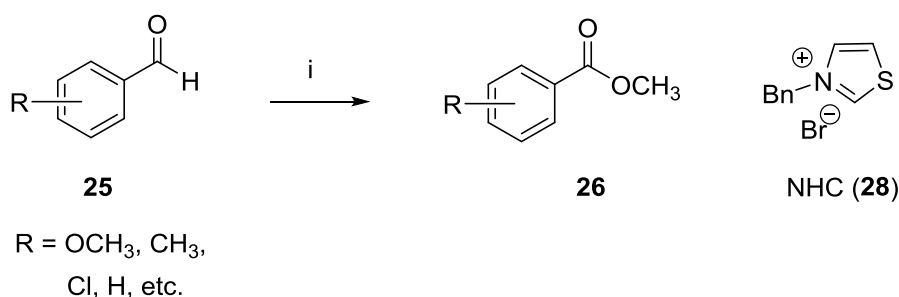
Li *et al.* have developed an oxidative esterification reaction between aldehydes **25** and alcohols **27** catalyzed by a combination of Cu(ClO₄)₂·6H₂O and InBr₃ using TBHP as an oxidant (**Scheme 13**).



Scheme 13: (i) Cu(ClO₄)₂·6H₂O, InBr₃, TBHP, 100 °C, 16 h, 42-91%.

Connon's approach (2008)^{35e}

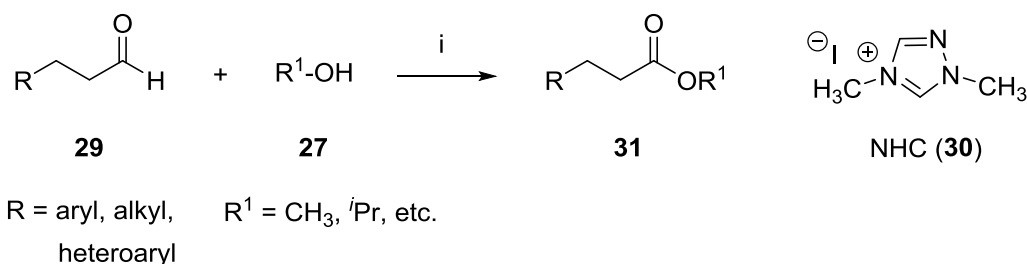
Connon *et al.* have developed a novel method of direct esterification of aldehydes with methanol using thiazolium NHC (**28**) as a catalyst and azobenzene as an oxidant in THF. Aromatic aldehydes **25** were thus converted to their corresponding esters **26** in moderate to good yields (**Scheme 14**).



Scheme 14: (i) NHC (**28**) (5 mol%), PhN=NPh, CH₃OH, THF, Et₃N, 25 to 60 °C, 24-48 h, 16-97%.

Scheidt's approach (2008)^{35f}

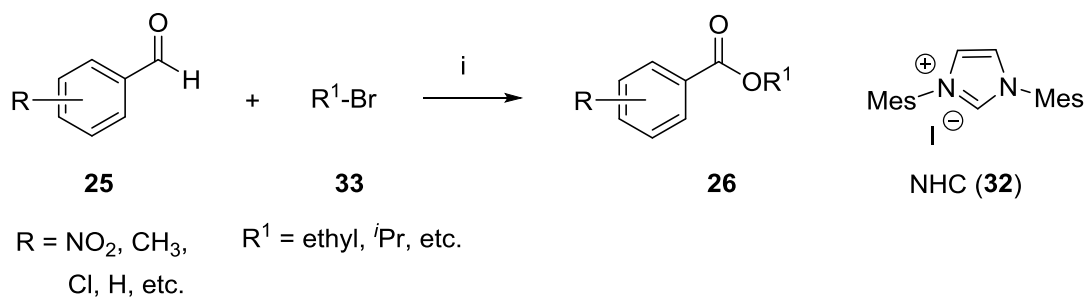
Scheidt *et al.* have described NHC-catalyzed oxidation of unactivated aldehydes to the corresponding carboxylic esters. Thus the reaction of unactivated aldehydes **29** with alcohols **27** in the presence of triazolium NHC catalyst (**30**) and oxidant MnO₂ provided esters **31** in high yields (**Scheme 15**).



Scheme 15: (i) NHC (**30**) (10 mol%), DBU, MnO₂, CH₂Cl₂, 25 °C, 0.5-3 h, 56-98%.

Xin's approach (2011)^{36a}

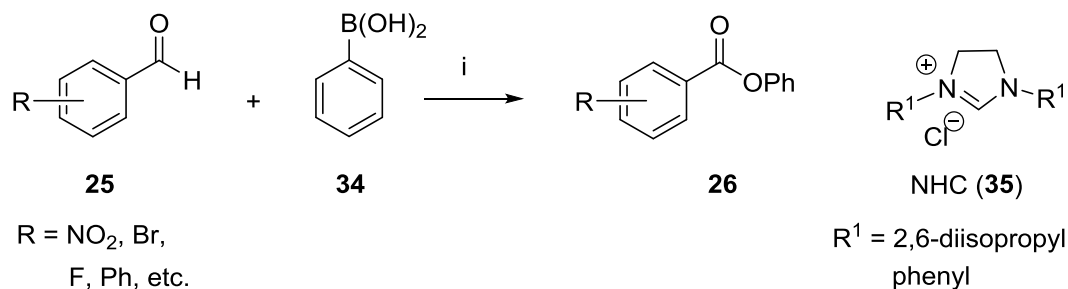
Xin *et al.* have developed an oxidative esterification reaction between aldehydes **25** and alkyl halides **33** catalyzed by imidazolium NHC (**32**) and molecular oxygen as an oxidant in THF at 50 °C that gave carboxylic esters in good yields (**Scheme 16**).



Scheme 16: (i) NHC (**32**) (10 mol%), DBU, O₂, THF, 50 °C, 24-72 h, 25-90%.

Arde's approach (2011)^{37a}

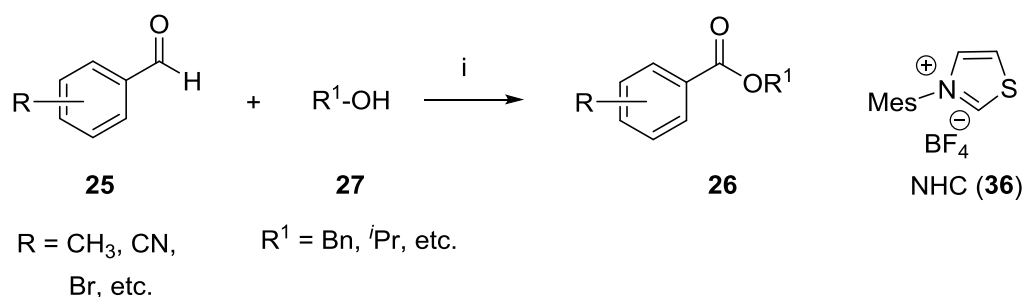
Arde *et al.* have developed an useful method of imidazolidine NHC (**35**) catalyzed aerobic oxidation of aromatic aldehydes **25** with boronic acids **34**. Aromatic aldehydes **25** were thus converted into their corresponding esters **26** in good yields (**Scheme 17**).



Scheme 17: (i) NHC (**35**) (10 mol%), Cs₂CO₃, O₂, toluene, 25 °C, 3-48 h, 25-99%.

Boydston's approach (2012)³⁴

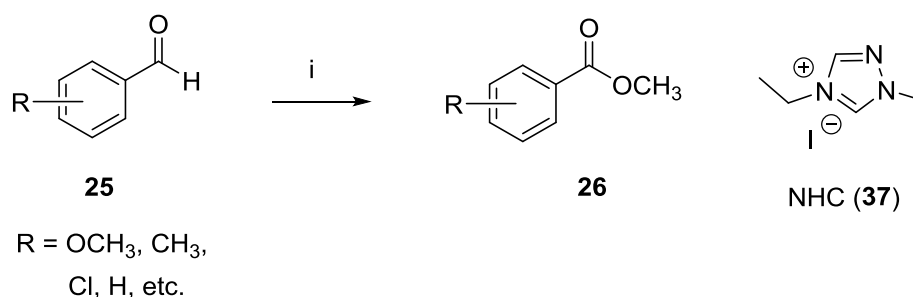
This methodology involves thiazolium NHC (**36**) catalyzed anodic oxidation of aldehydes **25** to the corresponding carboxylic esters **26** on reaction with alcohols **27** (**Scheme 18**). The electrochemical approach assumes the formation of electroactive intermediates that react with alcohol to provide esters.



Scheme 18: (i) NHC (**36**) (10 mol%), DBU, TBAB, CH₃CN, graphite anode, Pt cathode, +0.1V vs. Ag/AgNO₃, 2-56 h, 45-98%.

Delany's approach (2013)^{35j}

This methodology employs an additive-free mild protocol for triazolium NHC (**37**)-catalyzed direct esterification of aldehydes **25** with CH₃OH using O₂ as oxidant. No other stoichiometric oxidants or catalysts for O₂ activation were required (**Scheme 19**).



Scheme 19: (i) NHC (**37**) (15 mol%), DBU, THF:CH₃OH (1:1), O₂, 25 °C, 12-92 h, 15-94%.

4.2.3 Present Work

4.2.3.1 Objective

Although several reports for transformation of aldehydes to the corresponding esters in the presence of alcohols has been reported, there are certain drawbacks associated with them such as (i) heavy metals (Pd, Mn, Fe, etc) as oxidants; (ii) harsh reaction conditions; (iii) excess use of bases and alcohols/halides; (iv) use of more than stoichiometric amounts of oxidants and (v) effective for a limited range of substrates. In this regard, an organocatalytic additive-free mild protocol for oxidative esterification of aldehydes is highly desirable. This section describes NHC-catalyzed direct esterification of aromatic aldehydes with a variety of alcohols under aerobic condition using catalytic amount of DBU. On a similar line Delany *et al.* have reported,^{35j} entitled “NHC-catalysed aerobic aldehyde esterifications with alcohols: no additives or cocatalysts required” (see **Scheme 19**) using triazolium NHC (**37**) as a new catalyst several months after our publication. Since the method involves NHC as catalyst, a brief account of NHC catalyst is described below.

4.2.3.2 *N*-Heterocyclic Carbene Catalysis

Proline, a naturally available amino acid, has been studied extensively as organo catalyst for reactions occurring through enamine as well as iminium ion as intermediates.³⁸ In the last two decades, *N*-heterocyclic carbenes (NHC) have emerged as an important and powerful class of organocatalysts with tremendous applications in a variety of synthetic transformations and are receiving much attention as proline, because of their unique electronic properties. **Fig. 11** shows the presence of a carbene moiety stabilized by two adjacent π -donating atoms in NHC. The unsaturation in the backbone makes this an aromatic system, so that the carbene p-orbital is available to act as a π -acid. The ability of NHCs to act as both electron

donors and acceptors permits them to serve as organocatalysts in a variety of coupling reactions.³⁹

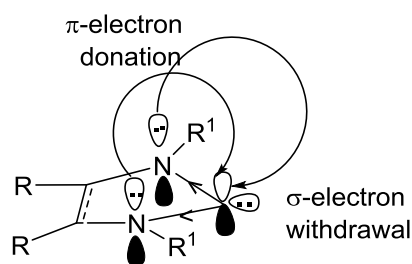


Fig. 11: Stabilization of *N*-heterocyclic carbenes

NHC can provide catalytic access to acyl anion equivalents, an umpolung strategy in which organic molecules react in an inverse manner compared to their innate polarity-driven reactivity.⁴⁰

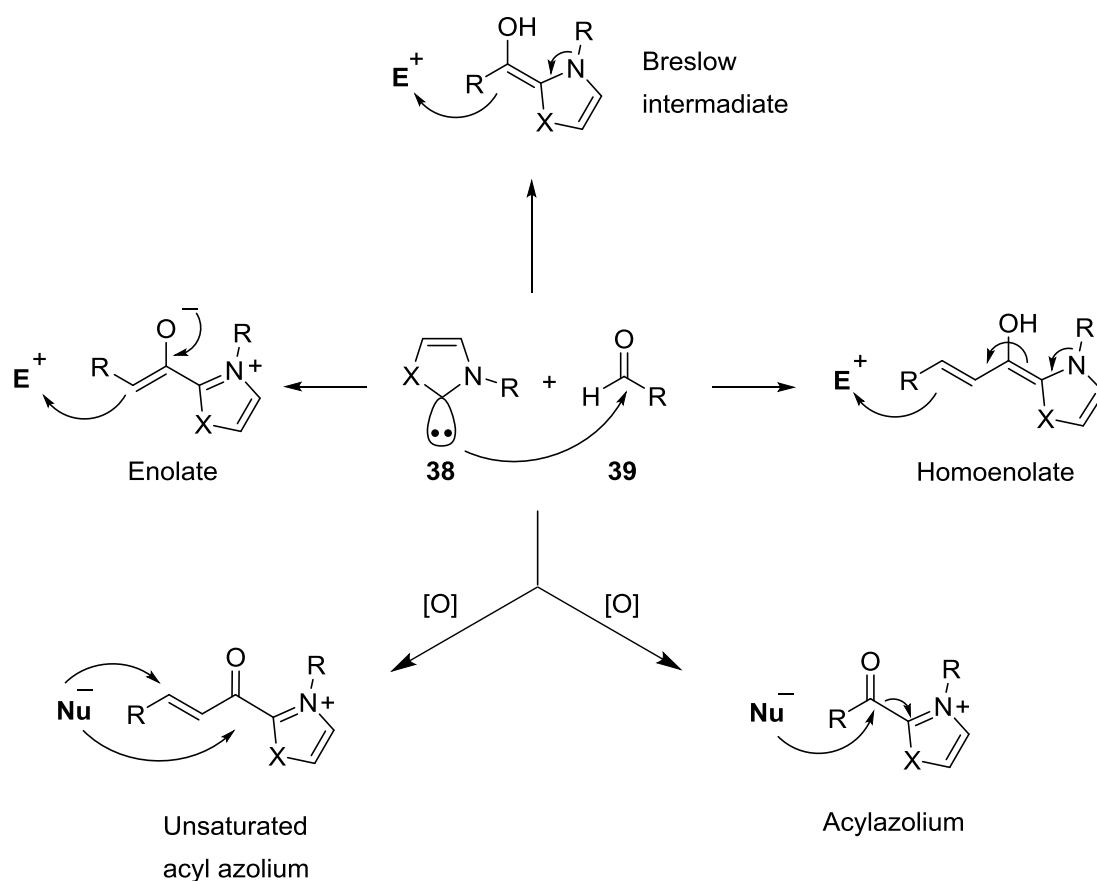


Fig. 12: Intermediates in NHC-organocatalysis

The majority of organocatalyzed transformations that are proceeding through umpolung are mediated by NHC. They have a number of modes of activation for a given substrate, like (i) acyl anion,⁴¹ (ii) hydroacylation,⁴² (iii) homoenolate,⁴³ and (iv) rebound catalysis,⁴⁴ in which it forms four important intermediates namely (i) Breslow intermediate, (ii) enolate, (iii) homoenolate and (iv) acylazolium and α,β -unsaturated acylazolium (**Fig. 12**). The most prominent intermediate in NHC catalysis is Breslow intermediate. It is assumed that the azolium precatalyst is deprotonated in the reaction mixture to afford a nucleophilic carbene **38**. Addition to an aldehyde **39** followed by proton transfers affords an enamine type intermediate, referred to as the Breslow intermediate,⁴⁵ in which the originally electrophilic carbon atom of the aldehyde has gained nucleophilic character. It can then attack a second molecule of aldehyde to furnish the benzoin product⁴⁶ or a Michael acceptor to afford a Stetter product.⁴⁷

Breslow intermediates also occur in biosynthesis; for example, 2-oxoacid dehydrogenases⁴⁸ such as pyruvate dehydrogenase, use thiamine pyrophosphate (TPP) as a carbene cofactor (**Fig. 13**). Decarboxylation is achieved by the reaction of carbene **40** with pyruvate to give **41**, that after CO₂ elimination provides Breslow intermediate **42**. Under aerobic conditions, enaminal **42** opens the dithiolane ring of a lipoyl group in a nucleophilic reaction to give an acetyl lipoamide thioester, which on transacylation with coenzyme A (CoASH) using enzymes eventually provides acetyl coenzyme A (CoASAc). Importantly, most published NHC-catalyzed processes mimic the reactivity of the naturally-occurring enaminal **42** of 2-oxoacid dehydrogenases and react *via* umpoled aldehydes. For anaerobic condition nature has developed an alternative reaction pathway for formation of CoASAc. Pyruvate ferredoxin oxidoreductase (PFOR), catalyses the decarboxylation of pyruvate to form

CoASAc *via* Breslow intermediate **42**, which reacts as a single electron-transfer reductant and the two electrons obtained during the turnover are transferred to ferredoxine. Electron transfer from the electron-rich enaminol **42** to a $[\text{Fe}_4\text{S}_4]^{2+}$ cluster leads to radical cation **43**. Renewed electron transfer in the presence of CoASH eventually provides CoASAc.

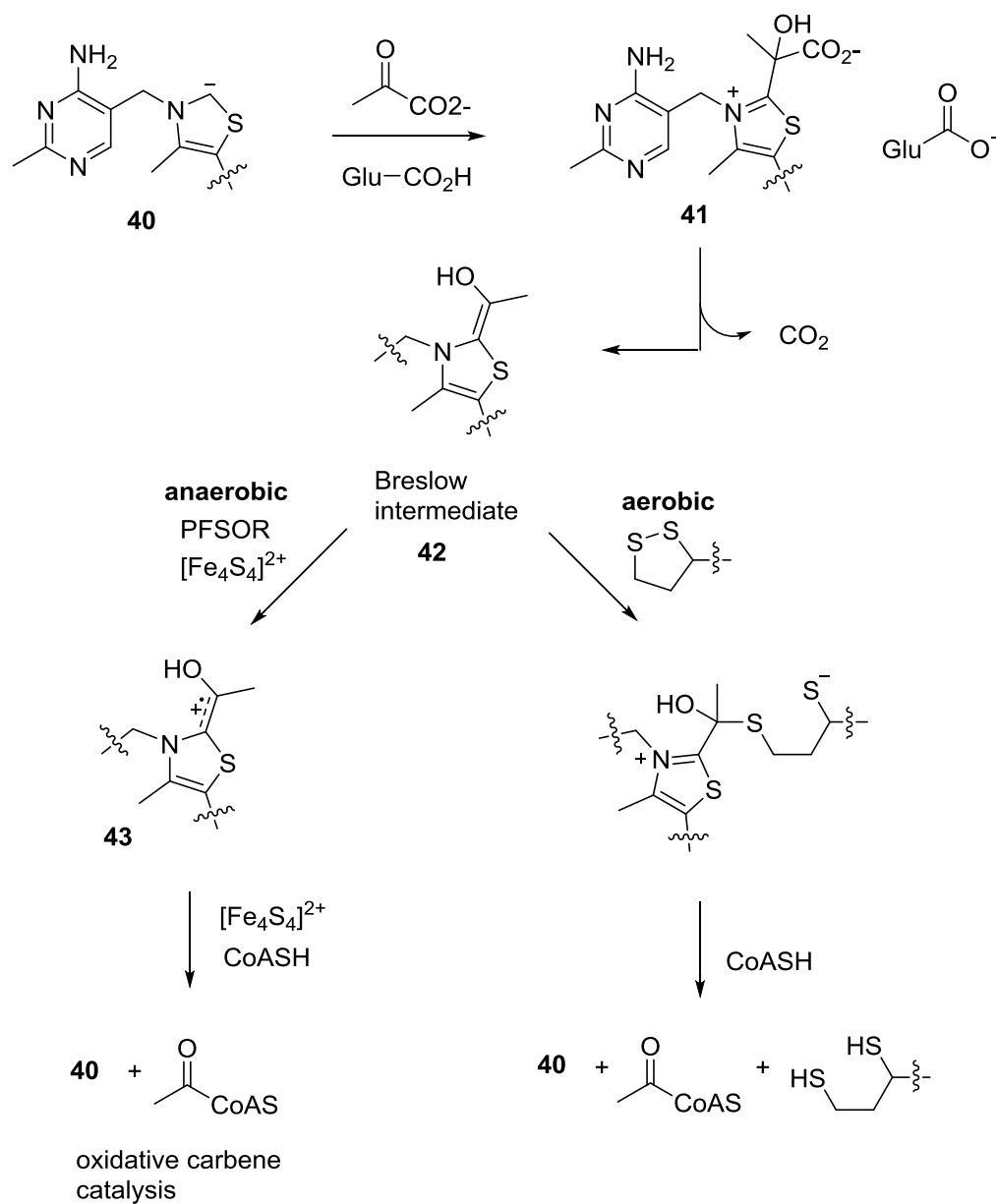


Fig. 13: TPP-mediated enzymatic transformation of pyruvate to CoASAc

4.2.4 Results and Discussion

The fact that Breslow intermediates can behave as single electron transfer reagents⁴⁸ inspired us to develop catalytic processes for oxidative esterification of aldehydes, that proceeds *via* oxidation of enaminol intermediates. The major challenge in the oxidation of aldehydes using NHC is the identification of mild oxidant, with the following characteristics (i) it is compatible with the carbene catalyst; (ii) avoids the formation of carboxylic acid by-products and (iii) shows high functional group compatibility. We envisaged that molecular O₂ should be an ideal oxidant for this purpose based on its nature of reactivity, low cost, and environment-friendly characteristics. **Fig. 14** shows some of the NHC precatalysts⁴⁹ **44-49** that were examined for the oxidative esterification of aldehydes using molecular O₂ as oxidant.

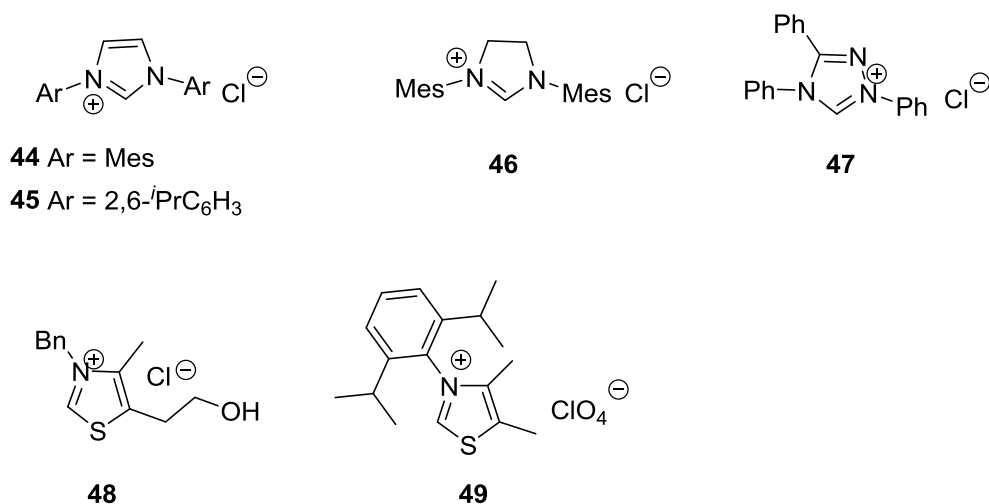
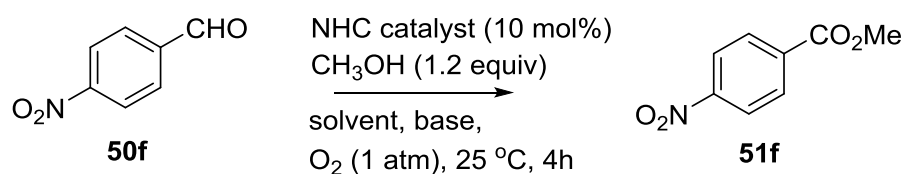


Fig. 14: *N*-Heterocyclic carbene precatalysts screened for the oxidative esterification of aromatic aldehydes

In a preliminary study, when 4-nitrobenzaldehyde **50f** was subjected to oxidative esterification with MeOH (1 equiv) in the presence of NHC **44** (10 mol%) and DBU (20 mol%) as base in THF under N₂ atmosphere at 25 °C, no trace of ester **51f** was

formed. When the same experiment was conducted in open air, **51f** was indeed isolated in low yields (45%). However, a dramatic increase in yield (70%) was realized when the experiment was carried out under O₂ atmosphere (O₂ balloon, 1 atm). Out of several NHC catalysts screened, imidazolium salt **44** showed higher catalytic activity,⁵⁰ providing the desired product **51f** in 70% yield.

Table 1: Oxidative esterification of 4-nitrobenzaldehyde: optimization studies^a



entry	NHC catalysts (10 mol%)	base (20 mol%)	solvent	yield of 51f (%) ^b
1	No catalyst	DBU	THF	-
2	1a	DBU	THF	76 (45) ^c
		DBU	CH ₂ Cl ₂	60
		DBU	CH ₃ CN	58
		DBU	DMSO	27
		DBU	toluene	60
3	1a	K ₂ CO ₃	THF	56
		Et ₃ N	THF	40
		ⁿ BuLi	THF	16
		K ^t OBu	THF	12
4	1b	DBU	THF	36
5	1c	DBU	THF	54
6	1d	DBU	THF	14
7	1e	DBU	THF	27
8	1f	DBU	THF	26

^aReaction conditions: 4-nitrobenzaldehyde (5 mmol), methanol (6 mmol), NHC catalyst (10 mol%), base (20 mol%), 25 °C, O₂ (1 atm), 4h. ^bisolated yield after column chromatographic purification. ^cunder open air.

When NHC catalyst loading was increased from 10 mol% up to 20 mol%, there was neither improvement in the yield of ester formed nor reduction in the time required for completion of the reaction. Additionally, increasing the temperature from 25 °C to 50 °C had deleterious effect in the yield, which may be due to the formation of unwanted acid by-product. Surprisingly, increasing the amount of alcohol from 1 equiv to 1.2 equiv resulted in improved yield (76%) of the ester formed. However, a further systematic increase in the amount of alcohol (from 1.2 up to 3 equiv) had no effect on yield improvement. Among several solvents and bases screened for the reaction, THF and DBU were found to be the effective solvent and base (**Table 1**).

With the optimized conditions (aldehyde (1 equiv), MeOH (1.2 equiv), NHC **44** (10 mol%), DBU (20 mol%), O₂, and THF at 25 °C) in hand, we then turned our attention to a variety of aromatic aldehydes **50a-p** as substrates having both electron-donating and -withdrawing groups in order to gauge the scope and generality of the reaction. The results of such studies are presented in **Table 2**. As can be seen, several aromatic aldehydes underwent oxidative esterification with methanol smoothly under mild conditions in moderate to good yields. Remarkably, substrates with electron-withdrawing groups showed higher reactivity as compared to electron-releasing substituents. Heteroaromatic aldehydes such as 3-pyridine carboxaldehyde **50o** also gave the corresponding ester in 76% yield, which are otherwise difficult to obtain under acid-catalysed esterifications due to salt formation of aldehyde **50o**. It is noteworthy that, 4-(methylthio)benzaldehyde **50e**, was not over-oxidized under our reaction condition. In the case of cinnamaldehyde, an inseparable mixture of saturated and unsaturated methyl esters were obtained,⁵¹ while aliphatic aldehydes failed to undergo this reaction, which may be a limitation.

Table 2: Oxidative esterification of aryl aldehydes: substrate scope ^a

ArCHO 50a-p		NHC 44 (10 mol%) MeOH (1.2 equiv) → DBU (20 mol%) O ₂ (1 atm), THF, 25 °C	ArCO ₂ Me 51a-p	
entry	substrates, Ar (50a-p)	time (h)	yield of 51a-p (%) ^b	
a	<i>m</i> -tolualdehyde	10	78	
b	4-OMe-benzaldehyde	30	72	
c	3,4-(OMe) ₂ -benzaldehyde	36	70	
d	3,4,5-(OMe) ₃ -benzaldehyde	36	65	
e	4-SMe-benzaldehyde	28	68	
f	4-NO ₂ -benzaldehyde	4	76	
g	3- NO ₂ -benzaldehyde	10	82	
h	4-Br-benzaldehyde	14	78	
i	3- Br-benzaldehyde	18	72	
j	4-Cl-benzaldehyde	18	79	
k	3-Cl-benzaldehyde	14	70	
l	4-F-benzaldehyde	10	76	
m	4-CF ₃ -benzaldehyde	7	69	
n	4-CN-benzaldehyde	6	72	
o	3-pyridinecarboxaldehyde	20	76	
p	furfural	24	63	

^aReaction conditions: aldehyde (5 mmol), methanol (6 mmol), NHC **44** (10 mol%), DBU (20 mol%), 25 °C, O₂ (1 atm). ^bisolated yield after column chromatographic purification.

A wide range of alcohols were then examined for oxidative esterification with 4-nitrobenzaldehyde as the substrate; the results are summarized in **Table 3**. Both primary and secondary alcohols including allylic, propargylic and benzylic alcohols underwent this reaction to give the corresponding esters in excellent yields.

Table 3: Oxidative esterification of 4-nitrobenzaldehyde: alcohol scope ^a

entry	alcohol components	time (h)	yield of ester % ^b
1	ethanol	4	80
2	2-propanol	7	76
3	benzyl alcohol	4	80
4	allyl alcohol	4	76
5	propargyl alcohol	5	82
6	(<i>S</i>)-tetrahydrofuran-3-ol ^c	16	66 (99% ee)

^aReaction conditions: 4-nitrobenzaldehyde (5 mmol), alcohol (6 mmol), NHC **44** (10 mol%), DBU (20 mol%), 25 °C, O₂ (1 atm). ^bisolated yield after column chromatographic purification; ^c the (*R*)-isomer gave the corresponding ester in 64% yields and 99% ee.

The formation of all ester products were confirmed unambiguously from their corresponding ¹H, ¹³C NMR and IR spectral data.

Example 1: The ¹H NMR spectrum of methyl 4-cyanobenzoate (**51n**) showed a typical singlet at δ 3.96 (s, 3H) corresponding to methoxyl protons (-OCH₃), while other signals at δ 7.75 (d, *J* = 8.6 Hz, 2H) and 8.14 (d, *J* = 8.6 Hz, 2H) correspond to aromatic protons. Its ¹³C NMR spectrum showed two characteristic carbon signals at δ 52.6 and 165.1 attributed to methoxyl carbon (-OCH₃) and ester carbonyl carbon

(C=O) respectively. Its IR spectrum displayed two characteristic strong absorption frequencies at 1727 and 2229 cm^{-1} indicating the presence of ester carbonyl and cyano functional groups respectively (**Fig. 15**).

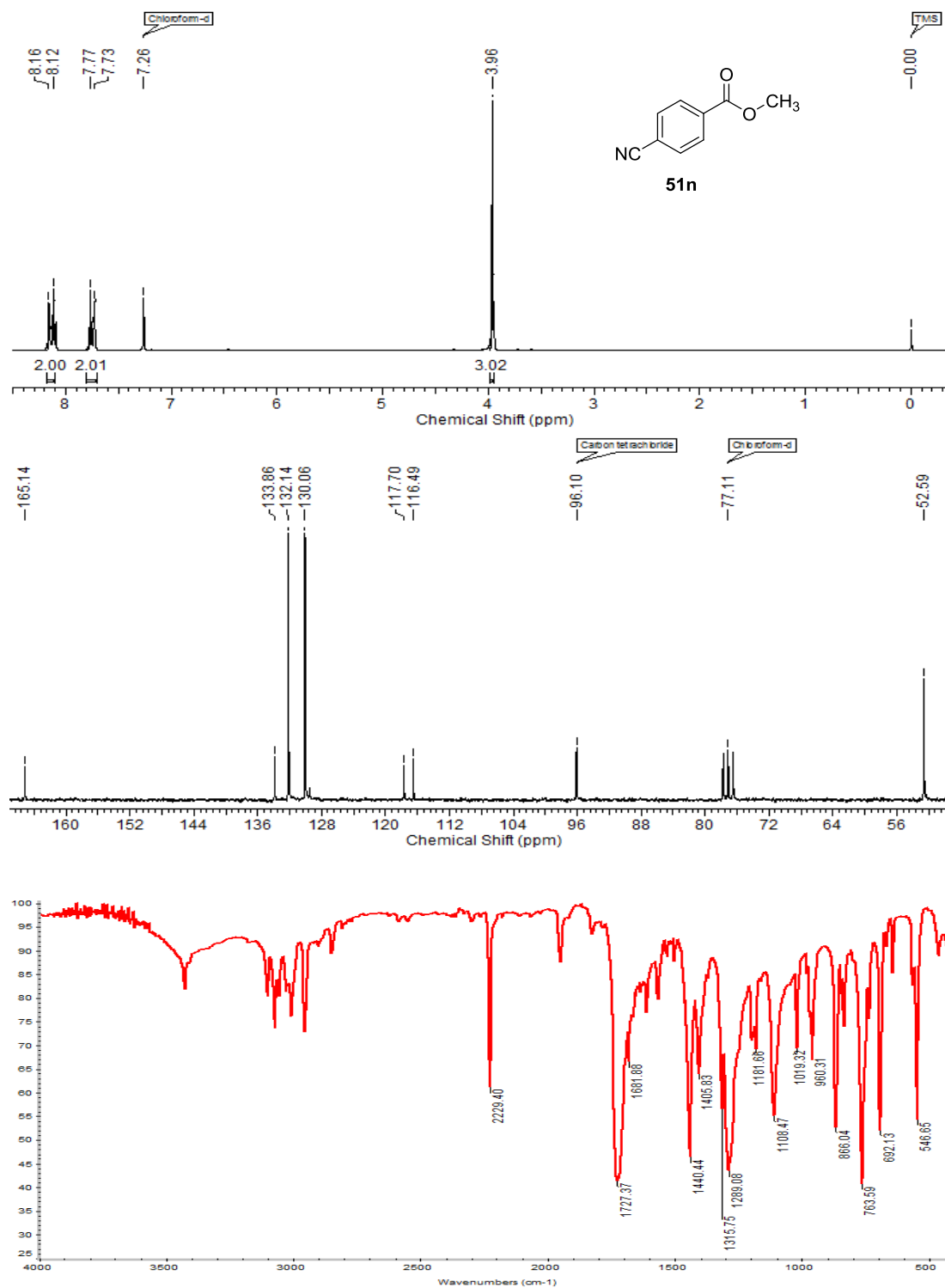


Fig. 15: ¹H, ¹³C NMR and IR spectra of methyl 4-cyanobenzoate (**51n**)

Example 2: The ^1H NMR spectrum of allyl 4-nitrobenzoate (**51t**) showed two typical proton signals at δ 4.86 (d, $J = 5.8$ Hz, 2H) and 5.31-5.46 (m, 2H) corresponding to allylic methylene and terminal olefinic protons respectively, while its ^{13}C NMR spectrum showed characteristic carbon signals at δ 119.0, 131.6 and 164.1 corresponding to olefinic and ester carbonyl carbons respectively (**Fig. 16**).

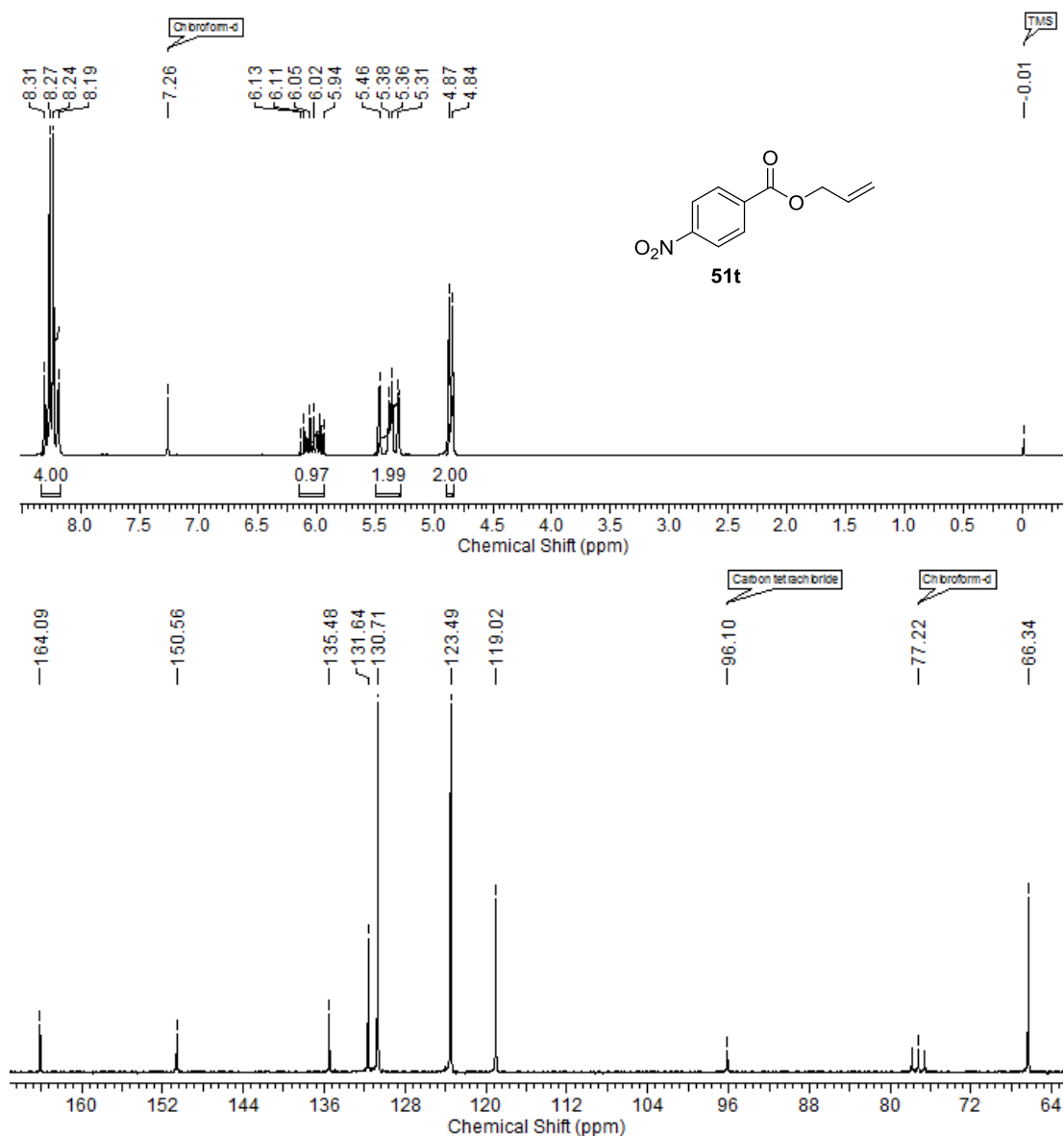
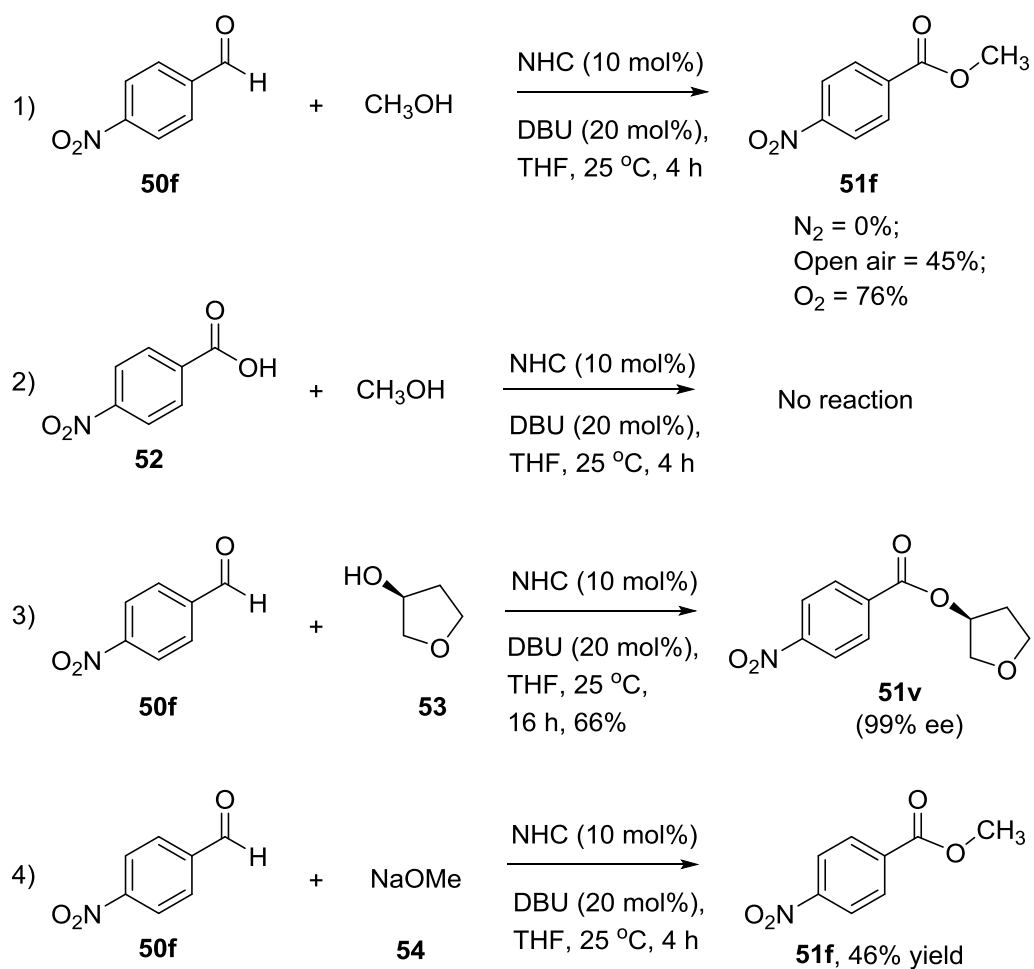


Fig. 16: ^1H and ^{13}C NMR spectra of allyl 4-nitrobenzoate (**51t**)

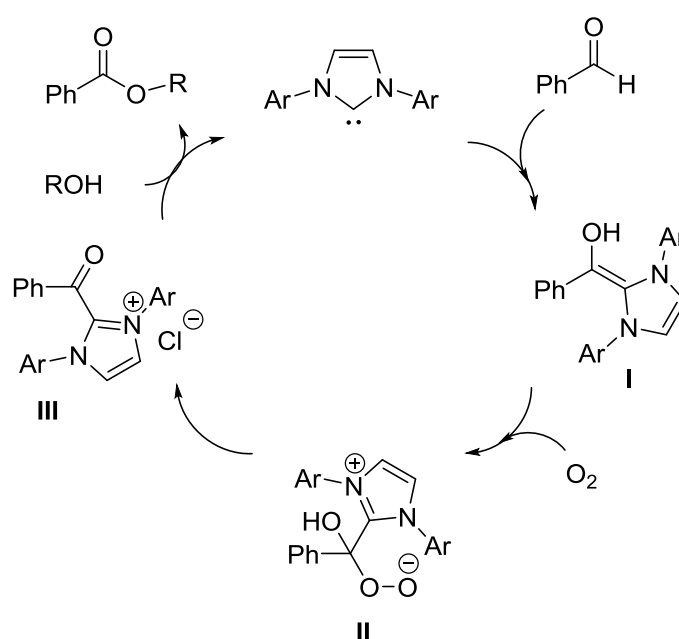
In order to gain insight into the mechanistic details of the reaction, the following experiments (see **Scheme 20**) were conducted: (i) for aldehyde **50f**, no esterification

took place under N₂ atmosphere, while at the same time low yield (45%) was obtained under open air conditions, suggesting the necessity of O₂ for realizing higher yields; (ii) the esterification reaction between *p*-nitrobenzoic acid (**52**) and methanol did not proceed under the reaction conditions, suggesting the absence of carboxylic acid as the intermediate; (iii) when (*S*)-tetrahydrofuran-3-ol **53** was used as alcohol partner, retention of configuration was indeed obtained in ester **51v**, thereby confirming the incorporation of alcohol oxygen into ester moiety; (iv) when 1 equivalent of sodium methoxide (**54**) was used as alcohol component, the corresponding methyl ester **51f** was obtained in 46% yield, suggesting the possibility of alkoxide anion formation as the intermediate (**Scheme 20**).



Scheme 20: A series of control experiments for mechanistic details

Based on the above results and literature precedents,^{37, 50} we have proposed a catalytic cycle in which peroxy anion **II**,³⁷ formed from reaction between Breslow intermediate **I** and O₂, has been depicted as the key intermediate in the esterification process. This on decomposition results in the formation of acyl intermediate **III**.^{37b} Subsequently, alkoxide ion⁵⁰ formed from alcohol reacts with **III** to give the corresponding ester with the liberation of NHC (**Scheme 21**).



Scheme 21: Probable mechanistic pathway for the oxidative esterification of aromatic aldehydes

4.2.5 Conclusion

A simple organocatalytic procedure for the direct oxidative esterification of aromatic aldehydes with alcohols employing NHC as catalyst and oxygen as oxidant at ambient conditions has been developed. The reaction is simple to carry out and the products are obtained in high yields and purity from stoichiometric amount of alcohol and catalytic amount of organic base.

4.2.6 Experimental Section

General experimental procedure for esterification of aromatic aldehydes

To a flame-dried round bottom flask equipped with a magnetic stir bar was added imidazolium salt **44** (0.170 g, 10 mol%), DBU (0.15 mL, 20 mol%) and THF (10 mL) in that order. The contents were evacuated and covered with molecular O₂ in balloon. The resultant reaction mixture was kept stirring at 25 °C for 45 min. To this mixture was added aromatic aldehydes **50a-p** (5 mmol) and alcohol (6 mmol) successively. It was allowed to stir at 25 °C. After completion of the reaction (monitored by TLC), THF was evaporated, H₂O (50 mL) added and the mixture extracted with EtOAc (3 x 50 ml). The combined organic layers were dried over anhyd. Na₂SO₄ concentrated to give crude ester, which was purified by silica gel-packed column chromatography to obtain pure esters, **51a-w**.

Methyl 3-methylbenzoate (**51a**)

Yield: 78%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 684, 815, 897, 976, 1043, 1166, 1239, 1381, 1607, 1682, 1722, 1873, 2496; **¹H NMR** (200 MHz, CDCl₃): δ 2.41 (s, 3H), 3.91 (s, 3H), 7.30-7.34 (m, 2H), 7.81-7.85 (m, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 21.2, 51.8, 126.7, 128.1, 130.0, 133.5, 137.9, 166.9; **Analysis:** C₉H₁₀O₂ requires C, 71.98; H, 6.71; found: C, 71.83; H, 6.59%.

Methyl 4-methoxybenzoate (**51b**)

Yield: 72%; colorless solid; **mp:** 49-51 °C, (lit.³² **mp:** 49 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 770, 848, 1029, 1103, 1168, 1256, 1280, 1317, 1434, 1458, 1606, 1716, 2953; **¹H NMR** (200 MHz, CDCl₃): δ 3.86 (s, 3H), 3.88 (s, 3H), 6.90 (d, *J* = 8.9 Hz, 2H), 7.98 (d, *J* = 8.9 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 51.7, 55.2, 113.5, 122.6, 131.5, 163.2, 166.5; **Analysis:** C₉H₁₀O₃ requires C, 65.05; H, 6.07; found: C, 64.91; H, 5.93%.

Methyl 3,4-dimethoxybenzoate (51c)

Yield: 70%; colorless solid; **mp:** 59-61 °C, (lit.³² **mp:** 60 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 764, 1133, 1271, 1294, 1434, 1514, 1600, 1714; **¹H NMR** (200 MHz, CDCl₃): δ 3.89 (s, 3H), 3.93 (s, 6H), 6.87 (d, $J = 8.4$ Hz, 1H), 7.53 (d, $J = 1.9$ Hz, 1H), 7.66 (dd, $J = 1.9, 8.4$ Hz, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 51.8, 55.8, 110.2, 111.9, 122.6, 123.5, 148.6, 152.9, 166.6; **Analysis:** C₁₀H₁₂O₄ requires C, 61.22; H, 6.16; found: C, 61.09; H, 6.12%.

Methyl 3,4,5-trimethoxybenzoate (51d)

Yield: 65%; colorless solid; **mp:** 82-85 °C, (lit.³² **mp:** 82 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 761, 1132, 1229, 1342, 1413, 1465, 1591, 1719; **¹H NMR** (200 MHz, CDCl₃): δ 3.90 (s, 3H), 3.91 (s, 9H), 7.28 (s, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.1, 56.1, 60.7, 106.7, 125.0, 142.1, 152.9, 166.4; **Analysis:** C₁₁H₁₄O₅ requires C, 58.40; H, 6.24; found: C, 58.31; H, 6.12%.

Methyl 4-(methylthio)benzoate (51e)

Yield: 68%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 759, 1116, 1302, 1281, 1402, 1708; **¹H NMR** (200 MHz, CDCl₃): δ 2.51 (s, 3H), 3.89 (s, 3H), 7.23 (d, $J = 8.6$ Hz, 2H), 7.91 (d, $J = 8.6$ Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 14.7, 51.8, 124.8, 126.3, 129.8, 145.3, 166.5; **Analysis:** C₉H₁₀O₂S requires C, 59.32; H, 5.53; found: C, 59.18; H, 5.41%.

Methyl 4-nitrobenzoate (51f)

Yield: 76%; colorless solid; **mp:** 94-96 °C, (lit.³² **mp:** 96 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 722, 818, 1076, 1104, 1136, 1262, 1298, 1338, 1536, 1618, 1719; **¹H NMR** (200 MHz, CDCl₃): δ 3.98 (s, 3H), 8.21 (d, $J = 8.6$ Hz, 2H), 8.30 (d, $J = 8.6$ Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.7, 123.5, 130.7, 135.4, 150.5, 165.0; **Analysis:** C₈H₇NO₄ requires C, 53.04; H, 3.89; N, 7.73; found: C, 52.92; H, 3.76; N, 7.62%.

Methyl 3-nitrobenzoate (51g)

Yield: 82%; colorless solid; **mp:** 78-80 °C, (lit.³² **mp:** 78 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 719, 823, 1072, 1100, 1133, 1266, 1292, 1350, 1528, 1615, 1722; **¹H NMR** (200 MHz, CDCl₃): δ 3.99 (s, 3H), 7.61-7.69 (m, 1H), 8.34-8.44 (m, 2H), 8.81-8.87 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.6, 124.4, 127.2, 129.5, 131.8, 135.1, 148.2, 164.6; **Analysis:** C₈H₇NO₄ requires C, 53.04; H, 3.89; N, 7.73; found: C, 52.96; H, 3.81; N, 7.67%.

Methyl 4-bromobenzoate (51h)

Yield: 78%; colorless solid; **mp:** 77-80 °C, (lit.³² **mp:** 79 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 758, 847, 1157, 1276, 1397, 1590, 1716; **¹H NMR** (200 MHz, CDCl₃): δ 3.92 (s, 3H), 7.57 (d, *J* = 10 Hz, 2H), 7.89 (d, *J* = 10 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.1, 127.9, 129.0, 131.0, 131.6, 166.0; **Analysis:** C₈H₇BrO₂ requires C, 44.68; H, 3.28; found: C, 44.59; H, 3.12%.

Methyl 3-bromobenzoate (51i)

Yield: 72%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 718, 746, 1067, 1121, 1260, 1293, 1436, 1571, 1727; **¹H NMR** (200 MHz, CDCl₃): δ 3.92 (s, 3H), 7.26-7.35 (m, 1H), 7.63-7.71 (m, 1H), 7.93-7.99 (m, 1H), 8.12-8.18 (m, 1H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.2, 122.4, 128.0, 129.8, 132.0, 132.6, 135.7, 165.4; **Analysis:** C₈H₇BrO₂ requires C, 44.68; H, 3.28; found: C, 44.53; H, 3.11%.

Methyl 4-chlorobenzoate (51j)

Yield: 79%; colorless gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 760, 1015, 1091, 1115, 1434, 1488, 1596, 1725; **¹H NMR** (200 MHz, CDCl₃): δ 3.92 (s, 3H), 7.41 (d, *J* = 8.6 Hz, 2H), 7.97 (d, *J* = 8.6 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 52.1, 128.6, 131.0, 139.3, 165.9; **Analysis:** C₈H₇ClO₂ requires C, 56.32; H, 4.14; found: C, 56.21; H, 4.03%.

Methyl 3-chlorobenzoate (51k)

Yield: 70%; colorless gum; **IR** (CHCl_3 , cm^{-1}): ν_{max} 748, 1126, 1259, 1283, 1295, 1437, 1728; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.92 (s, 3H), 7.32-7.40 (m, 1H), 7.48-7.54 (m, 1H), 7.88-7.94 (m, 1H), 7.97-8.02 (m, 1H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.2, 127.6, 129.5, 131.8, 132.8, 134.5, 165.5; **Analysis:** $\text{C}_8\text{H}_7\text{ClO}_2$ requires C, 56.32; H, 4.14; found: C, 56.23; H, 4.06%.

Methyl 4-fluorobenzoate (51l)

Yield: 76%; colorless gum; **IR** (CHCl_3 , cm^{-1}): ν_{max} 607, 767, 854, 1092, 1113, 1154, 1280, 1436, 1508, 1601, 1727; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.90 (s, 3H), 7.04-7.13 (m, 2H), 7.99-8.09 (m, 2H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.1, 115.4, 128.6, 131.0, 165.1, 166.1; **Analysis:** $\text{C}_8\text{H}_7\text{FO}_2$ requires C, 62.34; H, 4.58; found: C, 62.23; H, 4.39%.

Methyl 4-(trifluoromethyl)benzoate (51m)

Yield: 69%; pale yellow liquid; **IR** (CHCl_3 , cm^{-1}): ν_{max} 775, 865, 1019, 1068, 1327, 1413, 1440, 1730, 2957; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.92 (s, 3H), 7.70 (d, $J = 8.2$ Hz, 2H), 8.14 (d, $J = 8.2$ Hz, 2H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.3, 120.9, 125.2-125.4 (m), 126.3, 128.1, 129.9, 132.5-135.7 (m), 165.5; $\text{C}_9\text{H}_7\text{F}_3\text{O}_2$ requires C, 52.95; H, 3.46; found: C, 52.76; H, 3.33%.

Methyl 4-cyanobenzoate (51n)

Yield: 72%; colorless gum; **IR** (CHCl_3 , cm^{-1}): ν_{max} 763, 866, 960, 1019, 1108, 1181, 1289, 1315, 1440, 1727, 2229; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.96 (s, 3H), 7.75 (d, $J = 8.6$ Hz, 2H), 8.14 (d, $J = 8.6$ Hz, 2H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.6, 116.5, 117.7, 130.1, 132.1, 133.9, 165.1; **Analysis:** $\text{C}_9\text{H}_7\text{NO}_2$ requires C, 67.07; H, 4.38; N, 8.69; found: C, 66.91; H, 4.26; N, 8.52%.

Methyl nicotinate (51o)

Yield: 76%; colorless liquid; **IR** (CHCl_3 , cm^{-1}): ν_{max} 824, 959, 1072, 1292, 1350, 1436, 1528, 1615, 1722, 3045; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.93 (s, 3H), 7.33-7.40 (m, 1H), 8.23-8.29 (m, 1H), 8.71-8.76 (m, 1H), 9.16-9.19 (m, 1H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.3, 123.2, 126.0, 136.9, 150.8, 153.3, 165.5; **Analysis:** $\text{C}_7\text{H}_7\text{NO}_2$ requires C, 61.31; H, 5.14; N, 10.21; found: C, 61.26; H, 5.06; N, 10.12%.

Methyl furan-2-carboxylate (51p)

Yield: 63%; colorless liquid; **IR** (CHCl_3 , cm^{-1}): ν_{max} 797, 1121, 1177, 1197, 1306, 1479, 1731, 3127, 3144; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 3.90 (s, 3H), 6.50 (d, $J = 3.8$ HZ, 1H), 7.17 (d, $J = 3.8$ HZ, 1H), 7.57 (s, 1H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 52.0, 111.9, 118.0, 144.8, 146.5, 159.2; **Analysis:** $\text{C}_6\text{H}_6\text{O}_3$ requires C, 57.14; H, 4.80; found: C, 57.02; H, 4.69%.

Ethyl 4-nitrobenzoate (51q)

Yield: 80%; colorless solid; **mp:** 97-99 °C, (lit.^{37a} **mp:** 97-98 °C); **IR** (CHCl_3 , cm^{-1}): ν_{max} 757, 872, 1103, 1277, 1320, 1352, 1528, 1724; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 1.44 (t, $J = 7.4$ HZ, 3H), 4.43 (q, $J = 7.4$ HZ, 2H), 8.21 (d, $J = 8.9$ HZ, 2H), 8.30 (d, $J = 8.9$ HZ, 2H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 14.2, 61.9, 123.5, 130.6, 135.8, 150.5, 164.4; **Analysis:** $\text{C}_9\text{H}_9\text{NO}_4$ requires C, 55.39; H, 4.65; N, 7.18; found: C, 55.21; H, 4.52; N, 7.01%.

Isopropyl 4-nitrobenzoate (51r)

Yield: 76%; colorless solid; **mp:** 105-108 °C, (lit.^{37a} **mp:** 105-106 °C); **IR** (CHCl_3 , cm^{-1}): 717, 874, 1103, 1287, 1322, 1349, 1375, 1525, 1607, 1713; **$^1\text{H NMR}$** (200 MHz, CDCl_3): δ 1.39 (d, $J = 6.4$ HZ, 6H), 5.24-5.31 (m, 1H), 8.19 (d, $J = 8.4$ HZ, 2H), 8.27 (d, $J = 8.4$ HZ, 2H); **$^{13}\text{C NMR}$** (50 MHz, CDCl_3): δ 21.8, 69.7, 123.4, 130.6,

136.2, 150.4, 164.1; **Analysis:** C₁₀H₁₁NO₄ requires C, 57.41; H, 5.30; N, 6.70; found: C, 57.30; H, 5.19; N, 6.54%.

Benzyl 4-nitrobenzoate (51s)

Yield: 80%; colorless solid; **mp:** 82-84 °C, (lit.^{37a} **mp:** 81-82 °C); **IR** (CHCl₃, cm⁻¹): ν_{\max} 743, 1103, 1286, 1348, 1523, 1713; **¹H NMR** (200 MHz, CDCl₃): δ 5.40 (s, 2H), 7.37-7.46 (m, 5H), 8.21-8.31 (m, 4H); **¹³C NMR** (50 MHz, CDCl₃): δ 67.5, 123.5, 128.4, 128.6, 128.7, 130.7, 135.2, 135.4, 150.5, 164.3; **Analysis:** C₁₄H₁₁NO₄ requires C, 65.37; H, 4.31; N, 5.44; found: C, 65.23; H, 4.21; N, 5.31%.

Allyl 4-nitrobenzoate (51t)

Yield: 76%; colorless oil; **IR** (CHCl₃, cm⁻¹): ν_{\max} 720, 855, 933, 995, 1014, 1048, 1102, 1271, 1319, 1348, 1526, 1608, 1727; **¹H NMR** (200 MHz, CDCl₃): δ 4.86 (d, J = 5.8 Hz, 2H), 5.31-5.46 (m, 2H), 5.94-6.13 (m, 1H), 8.21 (d, J = 8.9 Hz, 2H), 8.29 (d, J = 8.9 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 66.3, 119.0, 123.5, 130.7, 131.6, 135.5, 150.6, 164.1; **Analysis:** C₁₀H₉NO₄ requires C, 57.97; H, 4.38; N, 6.76; found C, 57.78; H, 4.19; N, 6.59%.

Prop-2-ynyl 4-nitrobenzoate (51u)

Yield: 82%; yellow gum; **IR** (CHCl₃, cm⁻¹): ν_{\max} 715, 875, 1124, 1286, 1322, 1350, 1527, 1608, 1728, 3290; **¹H NMR** (200 MHz, CDCl₃): δ 2.55 (t, J = 2.5 Hz, 1H), 4.94 (d, J = 2.5 Hz, 2H), 8.24 (d, J = 8.4 Hz, 2H), 8.32 (d, J = 8.4 Hz, 2H); **¹³C NMR** (50 MHz, CDCl₃): δ 53.2, 75.8, 123.6, 130.9, 134.7, 150.8, 163.8; **Analysis:** C₁₀H₇NO₄ requires C, 58.54; H, 3.44; N, 6.83; found: C, 58.42; H, 3.29; N, 6.74%.

(S)-Tetrahydrofuran-3-yl 4-nitrobenzoate (51v)

Yield: 66%, colorless gum; $[\alpha]_{25}^D$ -31.24 (c 1.2, CH₂Cl₂); lit.^{37a} $[\alpha]_{25}^D$ +31.26 (c 1.0, CH₂Cl₂) for the corresponding (*R*)-enantiomer; **IR** (CHCl₃, cm⁻¹): ν_{\max} 720, 878, 1086, 1106, 1120, 1529, 1604, 1718, 2877, 2933, 3076; **¹H NMR** (200 MHz, CDCl₃):

δ 2.14-2.21 (m, 1H), 2.29-2.43 (m, 1H), 3.91-4.07 (m, 4H), 5.57-5.60 (m, 1H), 8.21 (d, $J = 8.8$ Hz, 2H), 8.30 (d, $J = 8.8$ Hz, 2H); ^{13}C NMR (50 MHz, CDCl_3): δ 32.8, 66.9, 72.9, 76.4, 123.5, 130.7, 135.2, 150.7, 164.1; **Analysis:** $\text{C}_{11}\text{H}_{11}\text{NO}_5$ requires C, 55.70; H, 4.67; N, 5.90; found: C, 55.62; H, 4.51; N, 5.79%.

(R)-Tetrahydrofuran-3-yl 4-nitrobenzoate (51w)

Yield: 64%; colorless gum; $[\alpha]_{25}^{\text{D}} +31.24$ (c 1.2, CH_2Cl_2); lit^{37aa} $[\alpha]_{25}^{\text{D}} +31.26$ (c 1.0, CH_2Cl_2); **IR** (CHCl_3 , cm^{-1}): ν_{max} 721, 879, 1084, 1105, 1122, 1528, 1604, 1718, 2877, 2933, 3076; **^1H NMR** (200 MHz, CDCl_3): δ 2.14-2.20 (m, 1H), 2.30-2.35 (m, 1H), 3.91-3.93 (m, 1H), 3.98-4.03 (m, 3H), 5.55-5.58 (m, 1H), 8.20 (d, $J = 8.8$ Hz, 2H), 8.29 (d, $J = 8.8$ Hz, 2H); ^{13}C NMR (50 MHz, CDCl_3): δ 32.8, 66.9, 72.9, 76.4, 123.5, 130.7, 135.2, 150.7, 164.1; **Analysis:** $\text{C}_{11}\text{H}_{11}\text{NO}_5$ requires C, 55.70; H, 4.67; N, 5.90; found: C, 55.60; H, 4.49; N, 5.76%.

4.2.7 References

1. Samii, A.; Nutt, J. G.; Ransom, B. R. *Lancet* **2004**, *363*, 1783.
2. Jankovic, J. *J. Neurol. Neurosurg. Psychiatr.* **2008**, *79*, 368.
3. Bronstein, J. M.; Tagliati, M.; Alterman, R. L.; Lozano, A. M.; Volkmann, J.; Stefani, A.; Horak, F.; Okun, M. S.; Foote, K. D.; Krack, P.; Pahwa, R.; Henderson, J. M.; Hariz, M. I.; Bakay, R. A.; Rezaei, A.; Marks, W. J.; Moro, E.; Vitek, J. L.; Weaver, F. M.; Gross, R. E.; DeLong, M. R. *Arch Neurol.* **2011**, *68*, 165.
4. Olanow, C. W.; Jankovic, J. *Mov Disord.* **2005**, *20*, 3.
5. Oldfield, V.; Keating, G. M.; Perry, C. M. *Drugs* **2007**, *67*, 1725.
6. Lecht, S.; Haroutianian, S.; Hoffman, A.; Lazarovici, P. *Ther. Clin. Risk Manag.* **2007**, *3*, 467.
7. (a) Aluf, Y.; Vaya, J.; Khatib, S.; Loboda, Y.; Finberg, J. P. M. *Neuropharmacology* **2013**, *65*, 48; (b) Van der Schyf, C. J.; Geldenhuys, W. J. *Int. Rev. Neurobiol.* **2011**, *100*, 107; (c) Weireb, O.; Amit, T.; Bar, O.; Youdim, M. B. H. *Int. Rev. Neurobiol.* **2011**, *100*, 191.
8. Binda, B. A.; Hubalek, F.; Li, M. *J. Med. Chem.* **2005**, *48*, 8148.
9. Gallagher, D. A.; Schrag, A. *CNS Drugs* **2008**, *22*, 563.
10. (a) Chahine, L. M.; Stern, M. B. *Int. Rev. Neurobiol.* **2011**, *100*, 151; (b) Weinreb, O.; Amit, T.; Riederer, P.; Youdim, M. B. H.; Mandel, S. A. *Int. Rev. Neurobiol.* **2011**, *100*, 127; (c) Zhu, W.; Xie, W.; Pan, T.; Jankovic, J.; Li, J.; Youdim, M. B. H.; Le, W. *J. Neurochem.* **2008**, *105*, 1970;

- (d) Am, O. B.; Amit, T.; Youdim, M. B. H. *Neurosci. Lett.* **2004**, 355, 169; (e) Meier-Davis, S. R.; Dines, K.; Arjmand, F. M.; Hamlin, R.; Huang, B.; Wen, J.; Christianson, C.; Shudo, J.; Nagata, T. *Cutan. Ocul. Toxicol.* **2012**, 31, 312.
11. Moussa, Y. B. H.; Ruth, L.; David, L.; Tirtsah, B. P. *EP 0436492 A2/1991*.
 12. Moussa, Y. B. H.; Haifa, Y.; John, P.; Finberg, M.; Ruth, L.; David, L.; Paskin, T. B.; Yellin, H. *US 5457133/1991*.
 13. Reddy, R. P.; Davies, H. M. L. *Org. Lett.* **2006**, 8, 5013.
 14. Boulton, L. T.; Lennon, I. C.; Bahar, E. *US 7476757 B2/2009*.
 15. Praveen, C.; Reddy, V. V.; Kumar, V.; Srinivasulu, R. *WO2010/059913 A2/2010*.
 16. Reddy, K. T.; Kumar, K. S.; Omprakash, G.; Dubey, P. K. *Der Pharma Chemica*, **2011**, 3, 110.
 17. (a) Sterling, J.; Herzig, Y.; Goren, T.; Finkelstein, N.; Lerner, D.; Goldenberg, W.; Miskolczi, I.; Molnar, S.; Rantal, F.; Tamas, T.; Toth, G.; Zagyva, A.; Zekany, A.; Lavian, G.; Gross, A.; Friedman, R.; Razin, M.; Huang, W.; Kraiss, B.; Chorev, M.; Youdim, M. B. *J. Med. Chem.* **2002**, 45, 5260; (b) Arie, G. L.; Igor, Z.; Victor, P.; Maxim, S.; Gennady, N. *WO02/068376 A1/2002*; (c) Pereira, C. S.; Salgado, S.; Aguiar, F. R.; Mera, X. G.; Borges, J. E. R. *Synlett*, **2013**, 837.
 18. Yang, J. W.; Chandler, C.; Stadler, M.; Kampen, D.; List, B. *Nature* **1981**, 37, 2091.
 19. Bal, B. S.; Childers, W. E.; Pinnick, H. W. *Tetrahedron* **2007**, 63, 2745.
 20. Roesner, S.; Casatejada, J. M.; Sonawane, R. P.; Aggarwal, V. K. *Org. Lett.* **2011**, 13, 5740.
 21. Ye, B.; Yao, Z. J.; Burke, T. R. *J. Org. Chem.* **1997**, 62, 5428.
 22. Glovanni, M.; Marcello, R.; Roberto, T.; Graziano, G.; Diego, G. *WO2010/049379 A1/2010*.
 23. Davis, F. A.; Szcwcyk, J. M. *Tetrahedron Lett.* **1998**, 39, 5951.
 24. Larock, R. C. *Comprehensive Organic Transformation*: VCH: New York, **1989**; pp 840.
 25. (a) Corey, E. J.; Gilman, N. W.; Ganem, B. E. *J. Am. Chem. Soc.* **1968**, 90, 5616; (b) Sundararaman, P. Walker, E. C.; Djerassi, C. *Tetrahedron Lett.* **1978**, 19, 1627; (c) Garegg, P. J.; Olcson, L.; Oscarson, S. *J. Org. Chem.* **1995**, 60, 2200; (d) Ogawa, T.; Matsui, M. *J. Am. Chem. Soc.* **1976**, 98, 1629; (e) Lee, K.; Kim, H.; Hong, J. *Angew. Chem. Int. Ed.* **2012**, 51, 5735.
 26. (a) *The Art of Drug Synthesis*; Johnson, D. S., Li, J. J., Eds.; John Wiley and Sons: Hoboken, NJ, 2007; (b) Zapf, A.; Beller, M. *Top. Catal.* **2002**, 19, 101; (c) Stetter, J.; Lieb, F. *Angew. Chem. Int. Ed.* **2000**, 39, 1724.
 27. (a) Gopinath, R.; Patel, B. K. *Org. Lett.* **2000**, 2, 577; (b) Gopinath, R.; Paital, A. R.; Patel, B. K. *Tetrahedron Lett.* **2002**, 43, 5123; (c) Gopinath, R.; Barkakaty, B.; Talukdar, B.; Patel, B. K. *J. Org. Chem.* **2003**, 68, 2944.
 28. Travis, B. R.; Sivakumar, M.; Hollist, G. O.; Borhan, B. *Org. Lett.* **2003**, 5, 1031.
 29. Sayama, S.; Onami, T. *Synlett* **2004**, 2739.
 30. Raj, I. V. P.; Sudalai, A. *Tetrahedron Lett.* **2005**, 46, 8303.
 31. Shaikh, T. M.; Emmanuvel, L. *Synth. Comm.* **2006**, 37, 2641.
 32. Karade, N. N.; Budhewar, V. H.; Katkar, A. N.; Tiwari, G. B. *Arkivoc* **2006**, 11, 162.
 33. Amati, A.; Dosualdo, G.; Zhao, L.; Bravo, A.; Fontana, F.; Minisci, F.; Bjorsvic, H. R. *Org. Process Res. Dev.* **1998**, 2, 261.
 34. Finney, E. E.; Ogawa, K. A.; Boydston, A. J. *J. Am. Chem. Soc.* **2012**, 134, 12374.

35. (a) De Sarkar, S.; Grimme, S.; Studer, A. *J. Am. Chem. Soc.* **2010**, *132*, 1190; (b) De Sarkar, S.; Studer, A. *Angew. Chem. Int. Ed.* **2010**, *48*, 9266; (c) Rose, C. A.; Zeitler, K. *Org. Lett.* **2010**, *12*, 4552; (d) Maki, B. E.; Chan, A.; Phillips, E. M.; Scheidt, K. A. *Tetrahedron* **2009**, *65*, 3102; (e) Noonan, C.; Baragwanath, L.; Connon, S. J. *Tetrahedron Lett.* **2008**, *49*, 4003; (f) Maki, B. E.; Scheidt, K. A. *Org. Lett.* **2008**, *10*, 4331; (g) Maki, B. E.; Chan, A.; Phillips, E. M.; Scheidt, K. A. *Org. Lett.* **2007**, *9*, 371; (h) Inoue, H.; Higashiura, K. *J. Chem. Soc., Chem. Commun.* **1980**, 549; (i) Reddy, R. S.; Rosa, J. N.; Veiros, L. F.; Caddick, S.; Gois, P. M. P. *Org. Biomol. Chem.* **2011**, *9*, 3126.
36. (a) Xin, Y. C.; Shi, S. H.; Xie, D. D.; Hui, X. P.; Xu, P. F. *Eur. J. Org. Chem.* **2011**, 6527; (b) Maji, B.; Vedachalan, S.; Ge, X.; Cai, S.; Liu, X.-W. *J. Org. Chem.* **2011**, *76*, 3016.
37. (a) Arde, P.; Ramanjaneyulu, B. T.; Reddy, V.; Saxena, A.; Anand, V. *Org. Biomol. Chem.* **2012**, *10*, 848; (b) Meng, J.-J.; Gao, M.; Wei, Y.-P.; Zhang, W.-Q. *Chem. Asian. J.* **2012**, *7*, 872.
38. (a) List, B. *Tetrahedron* **2002**, *58*, 5573; (b) Aleman, J.; Cabrera, S. *Chem. Soc. Rev.* **2013**, *42*, 774.
39. (a) Moore, J. L.; Rovis, T. *Top. Curr. Chem.* **2009**, *291*, 77; (b) Phillips, E. M.; Chan, A.; Scheidt, K. A. *Aldrichimica Acta* **2009**, *42*, 55; (c) Nair, V.; Vellalath, S.; Babu, B. P. *Chem. Soc. Rev.* **2008**, *37*, 2691; (d) Enders, D.; Niemeier, O.; Henseler, A. *Chem. Rev.* **2007**, *107*, 5606; (e) Marion, N.; Diez-Gonzalez, S.; Nolan, S. P. *Angew. Chem. Int. Ed.* **2007**, *46*, 2988; (f) Zeitler, K. *Angew. Chem. Int. Ed.* **2005**, *44*, 7506; (g) Enders, D.; Balensiefer, T. *Acc. Chem. Res.* **2004**, *37*, 534.
40. (a) De Sarkar, S.; Biswas, A.; Samanta, C. R.; Studer, A. *Chem. Eur. J.* **2013**, *19*, 4664; (b) Ryan, J. S.; Candish, L.; Lupton, D. W. *Chem. Soc. Rev.* **2013**, *42*, 4906; (c) Bugaut, X.; Glorius, F. *Chem. Soc. Rev.* **2012**, *41*, 351; (d) Nair, V.; Menon, R. S.; Biju, A. T.; Sinu, C. R.; Paul, R. R.; Jose, A.; Sreekumar, V. *Chem. Soc. Rev.* **2011**, *40*, 5336; (e) Chiang, P.-C.; Bode, J.W. in *N-Heterocyclic Carbenes*, The Royal Society of Chemistry, London, **2011**, pp. 399; (f) Grossmann, A.; Enders, D. *Angew. Chem., Int. Ed.* **2012**, *51*, 314; (g) Vora, H. U.; Wheeler, P.; Rovis, T. *Adv. Synth. Catal.* **2012**, *354*, 1617.
41. Mattson, A. E.; Bharadwaj, A. R.; Scheidt, K. A. *J. Am. Chem. Soc.* **2004**, *126*, 2314.
42. Chan, A.; Scheidt, K. A. *J. Am. Chem. Soc.* **2006**, *128*, 4558.
43. Chan, A.; Scheidt, K. A. *Org. Lett.* **2005**, *7*, 905.
44. Kawanaka, Y.; Phillips, E. M.; Scheidt, K. A. *J. Am. Chem. Soc.* **2009**, *131*, 18028.
45. Breslow, R. *J. Am. Chem. Soc.* **1958**, *80*, 3719.
46. Ugai, T.; Tanaka, S.; Dokawa, S. *Yakugaku Zasshi* **1943**, *63*, 296.
47. Stetter, H.; Kuhlmann, H. *Chem. Ber.* **1976**, *109*, 2890.
48. (a) Ragsdale, S. W. *Chem. Rev.* **2003**, *103*, 2333; (b) Kluger, R.; Tittmann, K. *Chem. Rev.* **2008**, *108*, 1797; (c) Chabriere, E.; Vernede, X.; Guigliarelli, B.; Charon, M. H.; Hatchikian, E. C.; Fontecilla-Camps, J. C. *Science* **2001**, *294*, 2559.
49. (a) Arduengo, A. J.; Krafczyk, R.; Schmutzler, R.; Craig, H. A.; Goerlich, J. R.; Marshall, W. J.; Unverzagt, M. *Tetrahedron* **1999**, *55*, 14523; (b) Scholl, M.; Ding, S.; Lee, C. W.; Grubbs, R. H. *Org. Lett.* **1999**, *1*, 953; (c) Enders, D.; Breuer, K.; Kallfass, U.; Balensiefer, T. *Synthesis* **2003**,

- 8, 1292; (d) Benhamou, L.; Chardon, E.; Lavigne, G.; Bellemin-lapponnaz, S.; Cesar, V. *Chem. Rev.* **2011**, *111*, 2705.
50. Phillips, E. M.; Riedrich, M.; Scheidt, K. A. *J. Am. Chem. Soc.* **2010**, *132*, 13179.
51. Maki, B. E.; Chan, A.; Scheidt, K. A. *Synlett* **2008**, 1306.

LIST OF PUBLICATIONS

1. A concise enantioselective synthesis of (+)-goniodiol and (+)-8-methoxygoniodiol *via* Co-catalyzed HKR of *anti*-(2*SR*, 3*RS*)-3-methoxy-3-phenyl-1,2-epoxypropane; **Kiran, I. N. C.**; Reddy, R. S.; Suryavanshi, G.; Sudalai, A. *Tetrahedron Lett.* **2011**, 52, 438.
2. CN-assisted oxidative cyclization of cyano cinnamates and styrene derivatives: a facile entry to 3-substituted chiral phthalides; Reddy, R. S.; **Kiran, I. N. C.**; Sudalai, A. *Org. Biomol. Chem.* **2012**, 10, 3655.
3. *N*-Heterocyclic carbene catalyzed esterification of aromatic aldehydes with alcohols under aerobic conditions; **Kiran, I. N. C.**; Lalwani, K.; Sudalai, A. *RSC Adv.* **2013**, 3, 1695.
4. Enantioselective synthesis of (-)-polysphorin analog *via* Co-catalyzed HKR of *syn*-(2*SR*, 3*SR*)-3-(methoxymethyl)oxy-3-(3,4,5-trimethoxyphenyl)-1,2-epoxypropane; **Kiran, I. N. C.**; Lalwani, K.; Sudalai, A. (*to be communicated*).
5. First enantioselective synthesis of new antitumour and antiinflammatory neolignan, surinamensinol B; Lalwani, K.; **Kiran, I. N. C.**; Sudalai, A. (*manuscript under preparation*).
6. Highly stereoselective synthesis of chiral 3-amino-1,2-diols *via* proline-catalyzed α -aminooxylation of β -aminoaldehydes; Venkataramasubramanian, V.; **Kiran, I. N. C.**; Sudalai, A. (Patent filed, *to be communicated*).
7. Enantioselective synthesis of cytoxazone epimers, taxol side chain and formal synthesis of *N*-thiolated-2-oxazolidinone; **Kiran, I. N. C.**; Venkataramasubramanian, V.; Sudalai, A. (Patent filed, *to be communicated*).
8. Organocatalytic enantioselective synthesis of rasagiline an anti-Parkinson drug; **Kiran, I. N. C.**; Sudalai, A. (*manuscript under preparation*).
9. A new process for the production of 3-substituted phthalides; Reddy, R. S.; **Kiran, I. N. C.**; Sudalai, A. *WO056340/2012*.
10. *N*-Heterocyclic carbene catalyzed esterification of aromatic aldehydes with alcohols under aerobic conditions; **Kiran, I. N. C.**; Lalwani, K.; Sudalai, A. *IN Ap. No 1685/DEL/2012*: 6/1/2012.
11. Cu-mediated annulation strategy for the effective synthesis of 3-substituted phthalides; Reddy, R. S.; Prasad, K. P.; **Kiran, I. N. C.**; Sudalai, A. *WOPCT /IN/0000051/2013*.
12. A single step enantioselective process for the preparation of 3-substituted chiral phthalides; Reddy, R. S.; **Kiran, I. N. C.**; Sudalai, A. *WO072830/2013*.