# SYNTHETIC AND MECHANISTIC STUDIES RELATED TO RIGID DERIVATIVES OF CYCLITOLS 

THESIS
SUBMITTED TO THE UNIVERSITY OF PUNE

FOR THE DEGREE OF

## DOCTOR OF PHILOSOPHY

IN

## CHEMISTRY

BY

## S. DEVARAJ

Division of Organic Chemistry (Synthesis)
National Chemical Laboratory

Pune-411 008

## Dedicated to my parents

 sister, brothersand uncles

## CERTIFICATE

This is to certify that the work incorporated in the thesis entitled "Synthetic and mechanistic studies related to rigid derivatives of cyclitols" submitted by $\mathbf{S}$. Devaraj was carried out by him under my supervision at the National Chemical Laboratory, Pune, India. Such materials obtained from other sources have been duly acknowledged in the thesis.

Date:
Division of Organic Chemistry (Synthesis)
(Dr. M. S. SHASHIDHAR)
National Chemical Laboratory
Research guide
Pune - 411008.

## DECLARATION

I hereby declare that the thesis entitled "Synthetic and mechanistic studies related to rigid derivatives of cyclitols" submitted for Ph. D. degree to the University of Pune has not been submitted by me for a degree to any other University.

Date:
Division of Organic Chemistry (Synthesis)
(S. DEVARAJ)

National Chemical Laboratory
Pune - 411008.

## ACKJOWLEDGEMENT

I wish to express my sincere gratitude to my research supervisor Dr. M. S. Shashidhar, for giving me the chance to work in this fascinating field of inositol chemistry. My interactions with him have improved my quality of research life. Apart from research, i am very thankful for his personal care because of which our group never felt that we are away from our family.

I acknowledge with deep sense of gratitude Dr. Mohan M. Bhadbhade, Mr. Rajesh G. Gonnade and Dr. V. G. Puranik for their fruitful discussions and valuable suggestions as well as for providing X-ray crystal structure analysis.

I would like to thank Dr. Ganesh Pandey, Dr. N.N. Joshi, Dr. K. N. Ganesh, Dr. Hazra and Dr. Pore for their timely help and constant encouragement. I would also like to thank Dr. Dhavale for his fruitful suggestions during my work presentations and also for recording some spectra.

A special thanks goes to Dr. Rajamohanan, Dr. Phalgune and Mrs. Kavitha from NMR division, Dr. V. G. Shah, Dr. Sujatha Biswas, Mrs. Sunitha Sawant, Mrs. Chitra Sanas, Senthil kumar from microanalysis and Ms. Annegiri from Library. I would also like to thank Dr. Mrs. Vidya Shashidhar and Dr. Mrs. Ranjana Bhadbhade for their help and encouragement.

I am very grateful to my GOD GIFTED friends and very fortunate in this life to have these friends, Malli, Easu, Subbu, Jayanthi, Thalai (Sankar), Venkatesh, Chithambaram, Balakrishnan, Ramesh, Mr. Marimuthu, Francis, Ramanujam, Sivaramakrishnan, Karthikeyan and my senior friends Mangalesh, Pasupathy, Murugan, C. Ramesh, Mr. Santhanam (Subbu’s father), Viji akka, Thiagu, Kitcha, Karuppayyan. I would also like to express my sincere thanks to my extremely good friends Srinivas Dumbre, Sanjay, Shivanad Pai, Subramanium, Bidhan, Nagendra, Girish, Prabal, Nishant, Swarup, Ravi, Kishore kesari, prasanna, Rajendra, Raman. I thank my roommate Sambhaji for his care and cooperation during my stay in the hostel.

A lots of thanks goes to my friends Selvakannan, Elangovan, Victor, Selvakumar, Kannan, Vijayaraj, Pradeep, M. Sankar, Sasi, Prathap, Suresh kumar, Thiru, Emmmanuel, for their timely help.

I thank my colleagues Dr. Praveen, Dr. Sureshan, Dr. Aditya, Sachin, Manas, Manoj, Madhuri, Rajendra, Shobana and Shailesh for maintaining a cheerful atmosphere in the lab and their timely help. I sincerely thank my great friend Murali for his brotherly affection and I am fortunate to have such person as my junior. I would also like to thank him for all his help during the final stages of my work. I also thank Sunilji and Moreji for their regular help in lab maintenance.

I would like to thank my PG professors Dr. Ragunathan, Dr. Srinivas, Dr. Rajagopalan, Dr. Rajkumar, UG professors Dr. Parthasarathy, Dr. Roopsing, Dr. Sathyamoorthy and the then HOD of the Chemistry Department (1997) and my school teachers Mr. Ganesan, Gangadaran, Kirubagaran without whose excellent guidance and help, it would have been impossible for me to reach this platform.

I would like to thank my PG seniors Thirumamagal, Sureshbabu, Peruncheralathan, Manikandan, Jesudass and my classmates Anniyappan, Mahesh for their constant support during my studies.

I would like to express my deepest gratitude for the constant support and love that I received from my parents, sister Sumathi, brothers Kumar, Ravi, uncles Rajavelu, Perumal, Nagappan and sisters-inlaw. I am very fortunate and glad to be one of them in our affectionate family. I would also like to thank my cousin Chandar, who is my best friend since my childhood, for his invaluable support. I thank my cousins Malini, Bhaskar, saravanan, Gopinath, Ramesh, Kishore, santosh, my nephew Sanjay, my niece Swetha and kutty chellam Keerthana for their eternal affection. I would also like to thank my cousin sisters Sujatha, Nalini, Santhanalakshmi and Sudha for their perpetual affection.

Finally, I thank Director, NCL for the infra-structural facilities and CSIR for the award of Research Fellowship.

## CONTENTS

Abbreviations ..... i
Synopsis of the Thesis ..... iv
List of publications ..... xiii
CHAPTER 1
General observations on the known methodologies for the O-substitution in myo-inositol, its triol and diol derivatives
1.1. Introduction ..... 1
1.2. Protection of myo-inositol derived triols ..... 10
1.2.1. Protection of myo-inositol derived $1,2,6$-triols ..... 10
1.2.2. Protection of myo-inositol derived $1,3,5$-triols ..... 11
1.2.3. Protection of myo-inositol derived 4,5,6-triol ..... 12
1.2.4. Protection of myo-inositol derived $1,2,5$-triols ..... 12
1.2.5. Protection of myo-inositol derived $2,4,6$-triols ..... 13
1.3. Protection of myo-inositol derived diols ..... 15
1.3.1. Protection of myo-inositol derived 1,2 -diols ..... 15
1.3.2. Protection of myo-inositol derived 3(1),6(4)-diols ..... 18
1.3.3. Protection of myo-inositol derived 1(3),5-diols ..... 19
1.3.4. Protection of myo-inositol derived 1,6 -diols ..... 19
1.3.5. Protection of myo-inositol derived 4(6),5-diols ..... 20
1.3.6. Protection of myo-inositol derived 2,6(4)-diols ..... 21
1.4. Conclusions ..... 24
1.5. References ..... 26

## CHAPTER 2: SECTION A

## Chelation controlled regiospecific O-substitution of myo-inositol orthoesters: convenient access to orthogonally protected myo-inositol derivatives

2A.1. Introduction ..... 32
2A.2. Results and Discussion ..... 35
2A.3. Conclusions ..... 55
2A.4. Experimental section ..... 56
2A.5. References ..... 71
CHAPTER2: SECTION B
Chelation controlled regioselective $\mathbf{O}$-substitution in myo-inositol derived
1,3,5-triol and diols
2B.1. Introduction ..... 111
2B.2. Results and Discussion ..... 112
2B.2.1. O-Alkylation of 2,4,6-tri-O-benzyl-myo-inositol ..... 112
2B.2.2. O-Alkylation / tosylation of diols. O-Tosylation of racemic 1-O-allyl-2,4,6-tri-O-benzyl-myo-inositol ..... 114
2B.2.3. O-Alkylation of racemic $1,2: 4,5-\mathrm{di}-O$-isopropylidene-myo-inositol ..... 114
2B.2.4. O-Alkylation of racemic 2,3,4,5-tetra-O-benzyl-myo-inositol ..... 115
2B.2.5. O-Alkylation of racemic 3,4,5,6-tetra-O-benzyl-myo-inositol ..... 117
2B.2.6. O-Alkylation of racemic 1,2-O-isopropylidene-3,6-di-O-benzyl- myo-inositol ..... 119
2B.3. Conclusions ..... 127
2B.4. Experimental section ..... 128
2B.5. References ..... 139

## CHAPTER 3

## Attempted synthesis of rigid, myo-inositol derived, triaxial 1,3,5-triols

3.1. Introduction ..... 161
3.2. Results and Discussion ..... 166
3.3. Conclusions ..... 179
3.4. Experimental section ..... 180
3.5. References ..... 191

## Chapter 1

General observations on the known methodologies for the $\mathbf{O}$-substitution in myo-inositol, its triol and diol derivatives

Chapter 2: Section A
Chelation controlled regiospecific $\mathbf{O}$-substitution of myoinositol orthoesters: convenient access to orthogonally protected myo-inositol derivatives

## Chapter 2: Section B

Chelation controlled regioselective O-substitution in myoinositol derived 1,3,5-triol and diols

Chapter 3
Attempted synthesis of rigid, myo-inositol derived, triaxial

> 1,3,5-triols

ABBREVIATIONS

| Ac | Acetyl |
| :---: | :---: |
| $\mathrm{Ac}_{2} \mathrm{O}$ | Acetic anhydride |
| AIBN | Azoisobutryonitrile |
| All | Allyl |
| AMOH | Acetylmandelic acid |
| anhyd | Anhydrous |
| Bn | Benzyl |
| BOC | tert-Butoxycarbonyl |
| BOM | Benzyloxymethyl |
| Bt | Butyryl |
| $\mathrm{n}-\mathrm{Bu}$ | Normal Butyl |
| t-Bu | tert-Butyl |
| Bz | Benzoyl |
| Calcd. | Calculated |
| Cat. | Catalytic |
| Conc | Concentrated |
| m-CPBA | m-Chloroperbenzoic acid |
| CSA | Camphorsulfonic acid |
| DAG | Diacylglycerol |
| DBU | 1,8-Diazabicyclo[5,4,0]undec-7-ene |
| DCC | Dicyclohexylcarbodiimide |
| DCM | Dichloromethane |
| DIBAL-H | Diisobutylaluminium hydride |
| DIPEA | Diisopropylethylamine |


| DMAP | 4-N,N-Dimethylaminopyridine |
| :---: | :---: |
| DMF | $N, N$-Dimethyl formamide |
| DMSO | Dimethyl sulfoxide |
| DSC | Differential scanning calorimetry |
| ee | Enantiomeric excess |
| eq | Equivalent |
| Et | Ethyl |
| EtOAc | Ethyl acetate |
| g | Gram |
| GPI | Glycosyl phosphatidyl inositol |
| h | Hour(s) |
| HPLC | High performance liquid chromatography |
| Hz | Hertz |
| $\operatorname{Ins}(1,4,5) \mathrm{P}_{3}$ | D-myo-inositol-1,4,5-trisphosphate |
| $\operatorname{Ins}(1,2-\mathrm{cyc}, 4,5) \mathrm{P}_{3}$ | D-myo-inositol-1,2-cyclic,4,5-trisphosphate |
| IR | Infrared |
| M | Molar |
| Me | Methyl |
| MeCN | Acetonitrile |
| MEM | 2-Methoxyethoxymethyl |
| MHz | Mega Hertz |
| min. | Minute(s) |
| mL | Milli liter |
| mmol | Milli Mole |
| mnt | Menthyl |


| mp . | Melting point |
| :---: | :---: |
| NIS | $N$-Iodosuccinimide |
| NMR | Nuclear Magnetic Resonance |
| ORTEP | Oak Ridge thermal ellipsoid plot |
| Ph | Phenyl |
| PMB | p-Methoxybenzyl |
| PNB | p-Nitrobenzyl |
| $i-\operatorname{Pr}$ | iso-propyl |
| psi | Per square inch |
| Pyr | Pyridine |
| $\mathrm{R}_{\mathrm{f}}$ | Retention factor |
| rt | Room temperature (23-30 ${ }^{\circ} \mathrm{C}$ ) |
| TBDPS | tert-Butyldiphenylsilyl |
| TBS | tert-Butyldimethylsilyl |
| Tf | Trifluoromethanesulfonyl |
| TFA | Trifluoroacetic acid |
| THF | Tetrahydrofuran |
| TIPDS | 1,3-(1,1,3,3-Tetraisopropyldisiloxanylidene) |
| TIPS | Triisopropylsilyl |
| TLC | Thin layer chromatography |
| Tr | Trityl (or) triphenylmethyl |
| Ts | p-Toluenesulfonyl |
| TsOH | $p$-Toluenesulfonic acid |
| UV | Ultraviolet |

## Synopsis of the Thesis

The thesis entitled 'Synthetic and mechanistic studies related to rigid derivatives of cyclitols' is divided in to three chapters. The first chapter consists of a review of the relevant literature pertaining to the chemistry of myo-inositol necessary to place the work described in the subsequent chapters in proper context. The second chapter consists of the experimental work done to develop a general methodology useful for the preparation of myo-inositol derivatives. The third chapter describes the attempts towards the preparation of rigid derivatives of myo-inositol to achieve the goal mentioned above.

Chapter 1: General observations on the known methodologies for the O substitution in myo-inositol, its triol and diol derivatives.

In the past two decades a large amount of research has been carried out on the chemistry and biochemistry related to inositols ${ }^{1}$, mainly due to the realization of the role played by phosphorylated myo-inositol derivatives in important biological phenomena such as cellular signal transduction, calcium mobilization, insulin stimulation, exocytosis and anchoring of certain proteins to the cell membranes. ${ }^{2}$ These developments in biology, with wide implications in the field of medicine, necessitated simpler methods for the synthesis of various inositol derivatives. myoInositol 1 being inexpensive and naturally abundant has served as a good starting material for the preparation of phosphoinositols, their derivatives and analogs as well as some natural products (for example polyoxin J, Nojirimycin, Sphingofungin D). ${ }^{3}$ Hence this chapter gives a survey of the literature pertaining to the reactions of myoinositol (hydroxyl groups) and its partially protected derivatives. Results presented in this chapter emphasize on the observed regioselectivity during the O-substitution reactions of myo-inositol as well as triols and diols derived from it (Chart 1).






11



Chart 1 PG = protecting group
Although, regio- and enantioselective reactions of myo-inositol hydroxyl groups are known to occur in living systems, there is no method for the regioselective O-substitution of any one of the six hydroxyl groups (of myo-inositol) exclusively and perhaps it is not easy to achieve this in the laboratory. Hence our approach was to prepare two myo-inositol based triols (14 and 15) and develop methods for the regioselective O -substitution in them in such a way that each step is regioselective to one of the hydroxyl groups. Such an approach in principle would allow regioselective O-substitution of all the six myo-inositol hydroxyl groups (17, Scheme 1). Since the triols, myo-inositol 1,3,5-orthoformate (14) and 2,4,6-tri-O-benzyl-myo-inositol (15) can be obtained easily in high yields, ${ }^{4}$ we have studied the relative reactivity of hydroxyl groups in these two triols and developed methods for the regioselective protection in these myo-inositol derivatives.


Chapter 2: Section A: Chelation controlled regiospecific O-substitution of myoinositol orthoesters: convenient access to orthogonally protected myo-inositol derivatives.

This section describes the work that led to the development of a general method for the sequential regiospecific O-substitution of all the three hydroxyl groups present in myo-inositol 1,3,5-orthoesters. This could be achieved utilizing the subtle differences in reactivity exhibited by alkali metal alkoxides of myo-inositol 1,3,5orthoesters, due to differences in their ability to form chelates. This method provides convenient access to orthogonally protected myo-inositol derivatives (Scheme 2). A comparison of the methylation of racemic 4-O-trityl-myo-inositol 1,3,5-orthoformate (32) in the presence of sodium or lithium ions showed that stabilization of the C4alkoxide by chelation with lithium overrides steric hindrance offered by the C6-axial substituent in deciding the regioselectivity during the nucleophilic O-substitution. Scheme 2 shows the summary of the results presented and discussed in this section which were rationalized based on the difference in the abilities of sodium and lithium ions to form chelates (35-38) with oxygen atoms in myo-inositol-1,3,5- orthoformate derivatives.

$R^{1} \quad R^{2}$ $20 \mathrm{H} \quad$ Alkyl $21 \mathrm{CH}_{3}$ Alkyl

$R^{1} \quad R^{3} \quad R^{4}$

| $\mathbf{2 6 H}$ | Alkyl | Alkyl | $\mathrm{R}^{1}$ | $R^{3}$ |
| :--- | :--- | :---: | :--- | :---: |
| $\mathbf{2 7} \mathrm{H}$ | Alkyl | Ts | $\mathbf{2 3 H}$ | Alkyl |
| $\mathbf{2 8 H}$ | Alkyl | Ac | $\mathbf{2 4} \mathrm{CH}_{3}$ | Alkyl |
| $\mathbf{2 9 H}$ | Ts | Alkyl | $\mathbf{2 5 H}$ | Ts |
| $\mathbf{3 0 H}$ | Ts | Ts |  |  |
| $\mathbf{3 1} \mathrm{CH}_{3}$ | Ts | Alkyl |  |  |


$\begin{array}{lc}\quad R^{1} & R^{3} \\ 23 \mathrm{H}^{2} & \text { Alkyl } \\ 24 \mathrm{CH}_{3} & \text { Alkyl } \\ 25 \mathrm{H} & \text { Ts }\end{array}$


14 H

$19 \mathrm{CH}_{3}$




M
35 Li
36 Na


37 Li
38 Na

Scheme 2. a) (i) n-BuLi (2eq) / THF; (ii) alkyl halide (2eq) / DMF; b) LiH, alkyl halide, DMF; c) NaH (2eq), alkyl halide (2eq), DMF; d) NaH (1eq), alkyl halide or TsCl, DMF; e) (i) n-BuLi (1eq) / THF; (ii) alkyl halide or TsCl / DMF; f) LiH, alkyl halide or TsCl or $\mathrm{Ac}_{2} \mathrm{O}$, DMF; g) n-BuLi / THF, Mel / DMF; h) NaH / THF, Mel / DMF.

Chapter 2: Section B: Chelation controlled regioselective O-substitution in myoinositol derived 1,3,5-triol and diols.

This section describes a study on the O-substitution in a myo-insoitol derived 1,3,5-triol 15 and a few myo-inositol derived diols (39, 44, 47, 52 and 57) via their alkoxides generated using either n-butyllithium or sodium hydirde. All the diols and triol studied in this section (Scheme 3) do not posess the orthoformate moiety and hence are not rigid. O-substitution in lithium alkoxides of the 1,3,5-triol $\mathbf{1 5}$ occurred exclusively at the C-1(3) hydroxyl group which has a vicinal cis-ether oxygen. As a result, both the C 1 and C3 hydroxyl groups in $\mathbf{1 5}$ could be protected sequentially without disturbing the C5-
hydroxyl group. However the same reactions carried out using the sodium alkoxide resulted in a mixture of products ( $\mathbf{3 9}$ and 40). Similar trend (O-substitution in lithium


Scheme 3. a) n-BuLi / THF, (b) AllBr / DMF; c) TsCl / DMF; d) NaH / THF; e) $\mathrm{Ac}_{2} \mathrm{O}$, pyridine; f) $\mathrm{BnBr} / \mathrm{DMF}$.
alkoxides exclusively at the hydroxyl group which has a vicinal cis-ether oxygen) was observed for the O-substitution of myo-inositol based non-vicinal diols 39 and 44.

In the case of vicinal diols ( $\mathbf{4 7}, 52$ and 57 ), reaction of lithium and sodium alkoxides with alkyl halides gave a mixture of products. The observed regioselectivity (for the O-substitution at a hydroxyl group next to a cis-oxygen) was much better in reactions where the alkoxide was generated with butyllithium as compared to the reaction where the alkoxide was generated with sodium hydride. Variation of temperature for the benzylation of the 1,2 -diol 57 revealed that at lower temperatures O-substitution at the axial C2-O-position was preferred. These results suggest that the reaction of inositol derived diols and triol with alkyl halides or sulfonly chloride can be directed to a hydroxyl group having a vicinal cis-oxygen atom, in the presence of lithium ions. The results show that the observed regioselectivity during the O-substitution in myo-inositol derivatives is sensitive to reactions conditions such as the presence of the metal ion (sodium or lithium), reaction temperature and the solvent used for the reaction, which support the involvement of chelates (62-67).

## Chapter 3: Attempted synthesis of rigid, myo-inositol derived, triaxial 1,3,5triols.

This chapter describes our attempts toward the synthesis of rigid myo-inositol derived triaxial 1,3,5-triols 70. Very few cyclohexane based triaxial 1,3,5-triols are known in the literature. ${ }^{5}$ These triols are known to complex metal ions and hence could have potential applications in separation and detection of metal ions. Also, pertaining to the chemistry of myo-inositol and its derivatives described in previous chapters, it was of interest to examine the regioselectivity of O-substitution reactions among the three axial hydroxyl groups in the 'axial-rich' configuration of myoinositol (as in 70). Hence we attempted to prepare myo-inositol derivatives locked
with a bridge between the C4 and C6 hydroxyl groups in the configuration having five axially disposed hydroxyl groups. The bridges we attempted to make are shown in Scheme 4.



$\mathrm{R}^{3}=$ myo-inositol moiety or a linker


We succeeded in making the bridge with a sulfite or a sulfate moiety (Scheme 5), but we were unable to cleave the orthoester without disturbing the sulfur-oxygen linkage. Structures of all the sulfur-bridged compounds (79-84) were established by single crystal X-ray diffraction method. The sulfite and the sulfate moiety were sensitive to acid hydrolysis (in the case of orthoformates 79, 80, 82) or prevented hydrogenolysis (in the case of orthobenzoates $\mathbf{8 1}, \mathbf{8 3}, \mathbf{8 4}$ ) or hydride reduction of the orthoester moiety. Reaction of myo-inositol orthoester derivatives with appropriate alkyl or acyl halides for the preparation of other bridged derivatives (71, 72, 73 and 75) resulted in mixture of products and there was no evidence for the formation of the desired products.




No reaction
Scheme 5. a) $\mathrm{SOCl}_{2}$, pyridine, DCM; b) $\mathrm{RuCl}_{3} . \mathrm{H}_{2} \mathrm{O}, \mathrm{NaIO}_{4}, \mathrm{CH}_{3} \mathrm{CN}, \mathrm{H}_{2} \mathrm{O}, \mathrm{DCM}$.
c) (i) iso- $\mathrm{BuNH}_{2}, \mathrm{MeOH}$; (ii) $\mathrm{NaH}, \mathrm{BnBr}$, DMF; d) DIBAL, DCM;
e) $\mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{H}_{2}, \mathrm{MeOH}$ or EtOAc.

### 1.1. Introduction

Centuries of research in chemistry resulted in an understanding of the chemical behavior of small organic molecules and enabled their characterization and classification depending on the functional groups present in them. This is essentially due to the exclusive reactions that each functional group undergoes in the presence of other functional groups in a small molecule. When several functional groups are present in the same molecule it is often difficult to carry out the desired reaction on a particular functional group due to interference from others. This led to the development of protecting groups, which aid in masking one functional group from a reagent or reaction condition while exposing the other functional groups present in the same molecule for the same reagent and hence bring about a functional group transformation, which forms the basis of organic synthesis. However, in molecules with the same kind of functional groups e.g., polyols (carbohydrates, cyclitols), polyamines, polyacids etc. bringing about a reaction at a particular site is an inordinate task. This is achieved in living systems with the help of enzymes, but understanding of organic reactions has not yet reached a point where this is possible in laboratory reactors.

New methodologies for the selective protection and deprotection of functional groups continue to appear in the literature regularly. ${ }^{1}$ Understanding the reasons that govern differences in the reactivity of different functional groups in the same molecule is the key for successful regioselective functionalization of complex organic molecules. Such an understanding becomes crucial while working with molecules having non-equivalent functional groups of the same kind (polyols, polyamines,
polyacids, etc). In syntheses involving cyclitols and carbohydrates, hydroxyl group protection is unavoidable.

Achieving selective reaction at one hydroxyl group in polyhydroxy compounds such as carbohydrates and cyclitols or their partially protected derivatives is a challenge and a topic of current research interest. ${ }^{2}$ This has implications for the synthesis of several classes of biologically or medicinally important compounds as exemplified by carbohydrates, oligosaccharides, inositols and their derivatives as well as natural products that can be synthesized from these polyols. ${ }^{3}$ Classical methods for the discrimination of hydroxyl groups in a polyol include (a) selective reaction of primary alcohols, in the presence of secondary and tertiary alcohols, with electrophiles; (b) preferential O-substitution of an equatorial hydroxyl group over that of an axial hydroxyl group in cyclic alcohols; (c) preferential hydrolysis of trans acetals over cis acetals of vicinal cyclic diols. ${ }^{4}$ Hydrolysis or reduction of orthoesters of diols and triols are also reported ${ }^{5}$ to be specific although their use in synthesis is restricted to certain category of compounds. However, when all the nonequivalent hydroxyl groups in a molecule are secondary or tertiary in nature, obtaining selectively derivatized products is a formidable task, due to various reasons including subtle differences in their reactivity and / or difficulty in separation of isomeric products and by-products.

Phosphorylated cyclitols and their analogs have attracted the attention of chemists and biologists in the last two decades due to their involvement in various biological phenomena such as cellular signal transduction, calcium mobilization, insulin stimulation, exocytosis, cytoskeletal regulation, intracellular trafficking of vesicles and anchoring of certain proteins to cell membranes (Scheme 1.1). ${ }^{3,6}$


Cyclitols have also been used as starting materials for the synthesis of natural products and their analogs having interesting biological and medicinal properties, ${ }^{3 \mathrm{~d}, 7}$ as chiral auxiliary or catalyst in asymmetric synthesis ${ }^{8}$ (Scheme 1.2).


10 Spicamycin, $\mathrm{R}^{3}=\mathrm{Me}_{2} \mathrm{CH}\left(\mathrm{CH}_{2}\right)_{\mathrm{n}} \mathrm{CONHCH}_{2} \mathrm{CO}$ - or
$\mathrm{MeCH}_{2} \mathrm{CH}_{2}\left(\mathrm{CH}_{2}\right)_{\mathrm{n}} \mathrm{CONHCH}_{2} \mathrm{CO}-(\mathrm{n}=8-14)$ :
11 Spicamycin Amino Nucleoside(SAN), R3 $=\mathrm{H}$
12 SPM VIII, $\mathrm{R}^{3}=\mathrm{Me}\left(\mathrm{CH}_{2}\right)_{10} \mathrm{CONHCH}_{2} \mathrm{CO}-$
$13 \mathrm{KRN} 5500, \mathrm{R}^{3}=\mathrm{ME}\left(\mathrm{CH}_{2}\right)_{8} \mathrm{CH}=\mathrm{CHCH}=\mathrm{CHCONHCH}_{2} \mathrm{CO}-$





Scheme 1.2. a) 25, t-BuOK ( 0.2 eq ), DCM, $-78^{\circ} \mathrm{C}, 15 \mathrm{~min}, 65 \%$ ( $96 \% \mathrm{ee}$ ); b) $\mathrm{TiCl}_{4}, \mathrm{Et}_{2} \mathrm{O},-7{ }^{\circ} \mathrm{C}, 2 \mathrm{~h}, 80-90 \%$.

Consequently, different methodologies have been developed for the regioselective protection and deprotection of inositol hydroxyl groups. Partially protected inositol derivatives can also be obtained from starting materials other than inositols (Scheme
1.3).


Scheme 1.3. a) Hydroxyl groups protection / deprotection; b) Ferrier reaction; c) $\mathrm{SmI}_{2}$-mediated cyclization; d) Pseudomonas putida-mediated hydroxylation followed by dihydroxylation reactions; e) Hydroxylation.

Since the rest of this thesis deals with the reactions of myo-inositol and its derivatives a brief description of its nomenclature is given below. Inositols are cyclohexane hexols. Nine isomers are known in the literature (Chart 1.1), among these isomers myo-, scyllo- and chiro- occur in nature, the myo-isomer being the most abundant. myo-Inositol has one axial and five equatorial hydroxyl groups. The carbon bearing the axial hydroxyl group is designated as C 2 and the other ring carbons can be numbered from C 1 to C 2 starting from a C 1 atom and proceeding around the ring in clockwise or anticlockwise fashion. According to convention, an anticlockwise
numbering in asymmetrically substituted myo-inositol leads to the configuration with D-prefix and the clockwise numbering gives the substituted myo-inositol an L-prefix (Chart 1.1). An IUB nomenclature ${ }^{9}$ allowing all biologically relevant compounds to be denoted as D-isomers has also been proposed.




43 cis-

44 тисо-

L48 L-chiro-


49 D-1 or L-3 substitution

50 L-1 or D-3 substitution

Chart 1.1

For convenience, in the rest of this thesis, numbering for the O -substituents ( R groups) in myo-inositol derivatives have been given superscript numbers corresponding to the carbon bearing that particular R-group, irrespective of the number of R groups present in the structure (for examples see Chart 1.2). All the structures of myo-inositol derivatives which represent a single enantiomer or a diastereomer have been numbered with D- or L-prefix to distinguish the other structures of myo-inositol derivatives which represent a racemic mixture.


Interest in the chemistry of inositols was revived in the past two decades due to the involvement of its phosphorylated derivatives in various cellular processes (Scheme 1.1). There are continued efforts towards the synthesis of naturally occurring phosphoinositols, their analogs, new cyclitol derivatives and other natural products with interesting biological and physical properties. Central to these efforts is the regioselective derivatization of inositol hydroxyl groups. The rest of this chapter gives a survey of the literature pertaining to selective reactions of myo-inositol and its partially protected derivatives necessary to put the work described in the following chapters in proper context.

Classical approach for the protection of myo-inositol hydroxyl groups involved the use of acetals of acetone, cyclohexanone, cyclopentanone, benzaldehyde, camphor etc. ${ }^{10}$ These methods of derivatization resulted in the formation of a mixture of products from which the desired ketal had to be separated (Scheme 1.4).


Scheme 1.4. a) (i) Dry acetone, anhyd $\mathrm{ZnCl}_{2}$, acetic acid, reflux, $9 \mathrm{~h} ; \mathrm{Ac}_{2} \mathrm{O}$, Pyr;
(ii) alcoholysis; b) cyclopentanone, TsOH, benzene, reflux, $6 \mathrm{~h}, 29 \%$.
c) 1-ethoxy cyclohexene, TsOH (cat), DMF (or DMSO).

Gigg et al ${ }^{11}$ discovered that the dibenzoate 59 (Scheme 1.5) had very low solubility in organic solvents and exploited this property for the easy isolation of $\mathbf{5 9}$ from a mixture of isomeric ketal dibenzoates. The 3,6-diol $\mathbf{6 0}$ was obtained by hydrolysis of the dibenzoate 59. This procedure for the isolation of the diisopropylidene diol did not require chromatography.


Scheme 1.5. a) 2,2-Dimethoxy propane, TsOH (cat), DMSO, $110^{\circ} \mathrm{C}$; b) $\mathrm{BzCl}, \mathrm{Pyr}$, filtration; c) $\mathrm{NaOH}, \mathrm{MeOH}$, reflux.

More recently, Bruzik et al ${ }^{12}$ prepared diastereomeric D-camphor ketals. This method also achieved the resolution of myo-inositol derivatives in addition to the protection of its hydroxyl groups. Ley et al ${ }^{13}$ reported a highly regioselective formation of bis dispoke acetal $\mathbf{6 1}$ in modest yields, on treatment of myo-inositol with bisdihydropyran (62) in the presence of CSA in DMF (Scheme 1.6).


Scheme 1.6. a) 62, CSA, DMF, $100^{\circ} \mathrm{C}$

Frost et al ${ }^{14}$ reported the protection of vicinal trans hydroxyl groups in myoinositol with 2,2,3,3,-tetramethoxybutane to obtain the 2,5-diol $\mathbf{6 4}$ in very good yield. Similar regioselective protection of the 1,3,4,6-hydroxyl groups can be achieved (63) using TIPDSCl although in lower yields (Scheme 1.7). ${ }^{15}$


Scheme 1.7. a) 2,2,3,3-Tetramethoxybutane, $(\mathrm{MeO})_{3} \mathrm{CH}, \mathrm{CSA}, \mathrm{MeOH}$, reflux, $79 \%$; b) TIPDSCI (2.5 eq), Pyr, rt.

Lee and Kishi reported ${ }^{16}$ the protection of C1-, C3- and C5-hydroxyl groups in myo-inositol as the orthoformate. The orthoformate $\mathbf{6 5}$ was obtained as a single product in good yield (Scheme 1.8).


Scheme 1.8. a) $(\mathrm{EtO})_{3} \mathrm{CH}, \mathrm{DMSO}, \mathrm{TsOH}, 100^{\circ} \mathrm{C}$

Attempts have also been made to achieve selective reaction at one or a few hydroxyl groups of myo-inositol by acylation. Benzoylation ${ }^{17}$ of myo-inositol with benzoyl chloride in pyridine at room temperature yielded the pentabenzoate $\mathbf{6 6}$ as the major product (Scheme 1.9). However, such approaches are of little use since chemically distinguishing the benzoates in subsequent synthetic steps becomes difficult.


Scheme 1.9. a) $\mathrm{BzCl}(3.5 \mathrm{eq}), \mathrm{Pyr}, 2$ h.

The reactions of myo-inositol described so far show that derivatization of myoinositol hydroxyl groups often lead to mixture of products and hence the yield of the desired myo-inositol derivative becomes poor. One of the exceptions is the formation of the orthoester $\mathbf{6 5}$ which can be obtained as a single product in more than $90 \%$ yield. ${ }^{16}$ However the consequence of the initial protection of myo-inositol hydroxyl groups as ketals is that further regioselective protection needs to be achieved among diols and triols which is easier than handling all the six hydroxyl groups (in inositol) at a time. After this is achieved, hydroxyl groups protected as ketals can be released to give diols, which can be selectively protected. This scheme of reactions although provides a route for the protection of the hydroxyl groups of myo-inositol, it often makes a synthetic scheme quite elaborate and low yielding. Also usually the method of preparation is specific to one or a small set of myo-inositol derivatives.

### 1.2. Protection of myo-inositol derived triols

### 1.2.1. Protection of myo-inositol derived 1,2,6-triols

Since these triols possess vicinal hydroxyl groups, the cis-diol moiety can be protected selectively as the corresponding $\operatorname{ketal}^{18}$ (72, Scheme 1.10 ) owing to its thermodynamic stability. Alkylation ${ }^{19}$ and silylation ${ }^{20}$ on these triols resulted in the preferential formation of the corresponding C1-hydroxyl group protected derivatives 73 and 74. These results show that C1-hydroxyl group can be protected selectively in 1,2,6-triols.


Scheme 1.10. a) 2,2-Dimethoxypropane, TsOH , acetone (dry), rt, 2 h ; b) (i) $\mathrm{Bu}_{2} \mathrm{SnO}$, benzene, reflux, 20 h , (ii) $\mathrm{Bu}_{4} \mathrm{NI}, \mathrm{AllBr}, 70^{\circ} \mathrm{C}, 2 \mathrm{~h}$; c) TBDPSCI, imidazole, DMF.

### 1.2.2. Protection of myo-inositol derived 1,3,5-triols

Alkylation, ${ }^{21}$ acylation ${ }^{22}$ and silylation ${ }^{23}$ of these triols result in predominant reaction at the $\mathrm{C} 1(3)$-hydroxyl group (Scheme 1.11). When the R groups at C 4 and C6 positions are different, a mixture of products is obtained. Miller et al ${ }^{24}$ reported regio and enantioselective monophosphorylation of C 1 - or C3-hydroxyl groups ( $>98 \%$ ee) of the triol 76 using low-molecular weight peptide-based catalysts. This reaction gives an easy access to enantiomeric myo-inositol-1-phosphates. The reactions in scheme 1.11 show that C5-hydroxyl group is the least reactive among the three hydroxyl groups in 1,3,5-triols.




Scheme 1.11. a) $\mathrm{BnBr}, \mathrm{Bu}_{4} \mathrm{NBr}, 2 \mathrm{M} \mathrm{NaOH}, \mathrm{DCM}$; b) $\mathrm{BzCl}, \mathrm{Pyr}$;
c) TBSCI , imidazole, DMF ; d) $\mathrm{CIPO}(\mathrm{OPh})_{2}, 2.5 \mathrm{~mol} \% 84, \mathrm{Et}_{3} \mathrm{~N}$, toluene, $0^{\circ} \mathrm{C}$;
e) $\mathrm{CIPO}(\mathrm{OPh})_{2}, 2 \mathrm{~mol} \% 85, \mathrm{Et}_{3} \mathrm{~N}$, toluene, $0^{\circ} \mathrm{C}$.

### 1.2.3. Protection of myo-inositol derived 4,5,6-triol

Relative configurations of the three hydroxyl groups in these triols are all trans and hence ketalization of these triols could lead to the formation of a mixture of ketals, unlike in the case of 1,2,6-triols (Scheme 1.10). Diacylation ${ }^{25}$ of the triol D86 (Scheme 1.12) with benzoyl chloride gave the 4,5-dibenzoate D87 while monobenzoylation gave a mixture of 4- and 5-benzoates D88 and D89. This is in contrast to the reactions of other triols and diols having the C5-hydroxyl group free where C5-hydroxyl group is the least reactive.


Scheme 1.12. a) $\mathrm{BzCl}(2 \mathrm{eq}), \mathrm{Pyr}$; b) $\mathrm{BzCl}(1.3 \mathrm{eq})$, Pyr.

### 1.2.4. Protection of myo-inositol derived $\mathbf{1 , 2 , 5 - t r i o l s}$

Phosphorylation ${ }^{26}$ of the triol $\mathbf{L 9 0}$ (Scheme 1.13) resulted in predominant phosphorylation at the C1-hydroxyl group, perhaps due to the steric hindrance for the reaction at C2- and C5-hydroxyl groups. Attempts at kinetic resolution of racemic myo-inositol derivatives $\mathbf{9 2 - 9 5}$ by benzoylation ${ }^{27}$ in the presence of chiral $\mathrm{Cu}(\mathrm{II})$ complex 97 resulted in moderate to good enantiomeric excess depending on the R group in the ester at the $\mathrm{C} 1(3)$-position (Scheme 1.13).



(D)
(D)


Scheme 1.13. a) 96, Pyr- $\mathrm{HBr}_{3}, \mathrm{Et}_{3} \mathrm{~N}, \mathrm{DCM}$; b) 97 , BzCl, DIPEA, $0^{\circ} \mathrm{C}$

### 1.2.5. Protection of myo-inositol derived 2,4,6-triols

myo-Inositol 1,3,5-orthoesters are a group of 2,4,6-triols which are frequently used for the preparation of inositol derivatives. They can be obtained exclusively in several gram quantities from myo-inositol. Literature reports show that one of the axial hydroxyl groups or the equatorial C2-hydroxyl group in orthoesters $(\mathbf{6 5}, 98)$ can be derivatized with moderate to good selectivity by varying the reaction conditions. For example, monoacylation ${ }^{28}$ of these triols in the presence of pyridine resulted in the formation of C 2 -esters as the major products while the use of triethylamine or sodium hydride as a base resulted in the formation of C4-esters (Scheme 1.14).

Exclusive acylation of the C2-hydroxyl group can also be effected via intramolecular acyl migration ${ }^{28 a}$ in C 4 -esters $(\mathbf{1 0 0} \rightarrow \mathbf{1 0 1})$. Similarly, alkylation ${ }^{29}$ of these triols in the presence of sodium hydride leads to the formation of the C4-ether $\mathbf{9 9}$ as the only product.


Scheme 1.14. a) NaH , alkyl halide, DMF ; b) NaH (1 eq) or $\mathrm{Et}_{3} \mathrm{~N}$, acyl halide, DMF; c) $\mathrm{NaH}(2 \mathrm{eq})$, acyl halide (1 eq), DMF; d) $\mathrm{NaH}, \mathrm{DMF}$.

Dialkylation ${ }^{29 a, 30}$ of the triol $\mathbf{6 5}$ with two equivalents of alkyl halide and two equivalents of a base like sodium hydride gave a mixture of products (Scheme 1.15) with symmetrical diether being the major product, but the isolated yields were poor. The unsymmetrical dibenzyl ether $\mathbf{1 0 6}$ could be obtained in good yields by benzylation ${ }^{31}$ of the monoether $99\left(\mathrm{R}^{4}=\mathrm{Bn}\right)$ under acidic conditions with benzyltrichloroacetamidate (see section 1.3.6, Scheme 1.27). Acylation ${ }^{32}$ or sulfonylation ${ }^{33}$ of myo-inositol orthoesters with two equivalents of acyl chloride or sulfonyl chloride in the presence of pyridine also resulted in the formation of unsymmetrical diesters. Symmetrical ditosylate $\mathbf{1 0 5}$ could be obtained as the major product by tosylation ${ }^{33}$ of $\mathbf{6 5}$ using sodium hydride as the base. These literature reports show that regiospecific alkylation or acylation of these 2,4,6-triols can be
carried out with modest to good yields, but dialkylation or diesterification leads to a mixture of products.


Scheme 1.15. a) $\mathrm{NaH}(2 \mathrm{eq}$ ), alkyl halide (2 eq), DMF; b) Acyl halide (2 eq) or $\mathrm{TsCl}(2 \mathrm{eq})$, Pyr; c) $\mathrm{NaH}(2 \mathrm{eq}), \mathrm{TsCl}(2 \mathrm{eq})$, DMF.

### 1.3. Protection of myo-inositol derived diols

### 1.3.1. Protection of myo-inositol derived 1,2-diols

Alkylation, ${ }^{18,34}$ acylation, ${ }^{34 a, b, 35}$ silylation, ${ }^{36}$ phosphorylation ${ }^{37}$ and tosylation ${ }^{35 b, 38}$ of 1,2-diols resulted in major reaction at the C1-hydroxyl group, as expected (Schemes $1.16-1.18$ ). Acylation ${ }^{39}$ with chiral acids was used for the resolution of racemic myo-inositol derived diols. Dibutyltin oxide was the reagent most frequently used for the selective functionalization of the C1-hydroxyl group in 1,2 -diols. Use of potassium hydroxide or sodium hydroxide for benzylation ${ }^{34 a}$ of $\mathbf{1 1 1}$ (Scheme 1.16) in benzene resulted in exclusive reaction at the C1-hydroxyl group. The above methodologies show that equatorially disposed C1-hydroxyl group is more reactive than the axially disposed C2-hydroxyl group as expected.

|  |  |  | a or b | $\rightarrow \mathrm{R}^{5} \mathrm{O} \underbrace{\mathrm{R}^{4} \mathrm{O}}_{\mathrm{O}^{6}}$ |
| :---: | :---: | :---: | :---: | :---: |
| $\mathrm{R}^{3}$ | $\mathrm{R}^{4}$ | $\mathrm{R}^{5}$ | $\mathrm{R}^{6}$ | 119 |
| 111 Bn | Bn | Bn | Bn | Alkyl $=\mathrm{Bn}$ or All or PMB or Me |
| 112 Bn | PMB | PMB | Bn |  |
| 113 All | All | All | Bn |  |
| 114 PMB | Bn | Bn | Bn |  |
| 115 Bn | Bn | PMB | PMB |  |
| 116 All | All | Bn | Bn |  |
| 117 Bn | Bn | Bn | All |  |
| 118 Bn ( | $\left(\mathrm{CH}_{3} \mathrm{O}\right.$ | $\left.\left.\mathrm{H}_{3}\right) \mathrm{C}\right)_{2}$ | $B n$ |  |

Scheme 1.16. a) Alkyl halide, KOH or NaOH , benzene, reflux, $50-52 \%$; b) $\mathrm{Bu}_{2} \mathrm{SnO}$, alkyl halide, $72-96 \%$.


| $\mathrm{R}^{3}$ | $\mathrm{R}^{4}$ | $\mathrm{R}^{5}$ | $\mathrm{R}^{6}$ |  | $\mathrm{R}^{1}$ | $\mathrm{R}^{3}$ | $\mathrm{R}^{4}$ | $\mathrm{R}^{5}$ | $\mathrm{R}^{6}$ (\%) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 111 Bn | Bn | Bn | Bn | 125 | Bz | Bn | Bn | Bn | Bn (68) |
| 112 Bn | PMB | PMB | Bn | 126 | Ac | Bn | Bn | Bn | Bn (76) |
| 115 Bn | Bn | PMB | PMB | 127 | Ts | Bn | Bn | Bn | Bn (79) |
| 118 Bn | $\left(\mathrm{CH}_{3} \mathrm{O}(\mathrm{C}\right.$ | $\left.\left.\mathrm{H}_{3}\right) \mathrm{C}\right)_{2}$ | Bn | D128 | (+)-AM | Bn | PMB | PMB | Bn |
| 120 Me | Me | Me | Me | D129 | (+)-AM | Bn | Bn | PMB | PMB |
| 121 Ac | Ac | Ac | Ac | D130 | (+)-AM | Bn | $\left(\mathrm{CH}_{3} \mathrm{O}\right.$ ( | $\left.\left.\mathrm{H}_{3}\right) \mathrm{C}\right)_{2}$ | Bn |
| 122 All | PMB | Bn | All | 131 | Ac | Me | Me | Me | Me (47) |
| 123 Bz | Bz | Bz | Bn | 132 | Ts | Ac | Ac | Ac | Ac (55) |
| 124 Bn | All | All | Bn | D133 (+)-AM |  | All | PMB | $B n$ | All |
|  |  |  |  | D134 | mntOAc | Bz | Bz | Bz | Bn |
|  |  |  |  | 135 | TBS | $B n$ | All | All | Bn (87) |




Scheme 1.17. a) Acyl chloride or TsCl, Pyr; b) (+)-AMOH, DCC, DMAP, DCM, $-15^{\circ} \mathrm{C}$ c) mntOAcCl, Pyr, $0^{\circ} \mathrm{C}$; d) TBSCl, 2,6-Lutidine; e) BzCl, Pyr, rt, 16-40 h.


Scheme 1.18. a) 148, $\mathrm{Pyr}^{1}-\mathrm{HBr}_{3} . \mathrm{Et}_{3} \mathrm{~N}$; b) 149, $\mathrm{Pyr}-\mathrm{HBr}_{3}, \mathrm{DCM} /$ Pyr / $E t_{3} \mathrm{~N}(1: 0.1: 0.05),-20^{\circ} \mathrm{C}, 2 \mathrm{~h}, 0^{\circ} \mathrm{C}, 0.5 \mathrm{~h}$.

However, acylation at the C2-hydroxyl group could be achieved using orthoesters. ${ }^{5 a}$, ${ }^{23,40}$ Recently Schultz et al ${ }^{40 b}$ reported the regioselective acylation of 1,2-diol at C2position via orthoester intermediate (Scheme 1.19).


Scheme 1.19. a) (MeO) $)_{3} \mathrm{CCH}_{2} \mathrm{CH}_{2} \mathrm{Me}, \mathrm{CSA}$, toluene, rt, 30 min ; b) MeOH (wet), rt, 2 h .
Frost et al ${ }^{41}$ reported the regioselective oxidation of the 1,2-diol 111 at C2position to get the inosose 153 .


Scheme 1.20. a) (i) $\mathrm{Bu}_{2} \mathrm{SnO}, \mathrm{MeOH}$, reflux; (ii) $\mathrm{Br}_{2}, \mathrm{DCM}, \mathrm{Bu}_{3} \mathrm{SnOMe}, 50 \%$.

### 1.3.2. Protection of myo-inositol derived 3(1),6(4)-diols

Most of the diols belonging to this category consist of diacetals of myoinositol. Alkylation ${ }^{42}$ and acylation ${ }^{39 e}$ of 3,6 -diols usually result in a mixture of products, the C3-O-substituted derivative being the major constituent (Scheme 1.21). Alkylation of dicyclohexylidene diol $\mathbf{5 6}$ exclusively at the C3-hydroxyl group could be carried out using barium oxide-barium hydroxide octahydrate as the base ${ }^{43}$ or by the gradual addition of sodium hydride ${ }^{44}$ at $0-5{ }^{\circ} \mathrm{C}$ to provide kinetic control. Yield for the tosylation ${ }^{45}$ of the diol $\mathbf{6 0}$ was reported to be quite low. These examples illustrate that the C3-hydroxyl group in 3,6-diols is more reactive and can be functionalized with electrophiles in preference to the C6-hydroxyl group.


Scheme 1.21. a) $\mathrm{NaH}, \mathrm{BnBr}, \mathrm{DMF}, 45 \%$ (154); b) $\mathrm{NaH}, \mathrm{BnBr}$, toluene, reflux, $60 \%$ (157); c) $\mathrm{BaO}, \mathrm{Ba}(\mathrm{OH})_{2} \cdot 8 \mathrm{H}_{2} \mathrm{O}, \mathrm{BnBr}, 70 \%(157)$; d) $\mathrm{BaO}, \mathrm{Ba}(\mathrm{OH})_{2} \cdot 8 \mathrm{H}_{2} \mathrm{O}$, AllBr, DMF; e) TsCl , Pyr, 30\% (161); f) Benzoylimidazole, CsF, DMF, rt, 2 h, 65\% (162).

### 1.3.3. Protection of myo-inositol derived 1(3),5-diols

C1-Hydroxyl group appears to be more reactive than the C5-hydroxyl group in 1,5 -diols as acylation ${ }^{46}$ and phosphorylation ${ }^{47}$ result in the formation of the corresponding C1-O-derivative (Scheme 1.22).



Scheme 1.22. a) PhOCSCI, DMAP, MeCN, benzene, $0^{\circ} \mathrm{C}$, 20 h ;
b) $(\mathrm{iPr})_{2} \mathrm{NP}(\mathrm{OBn})_{2}(1 \mathrm{eq}), 1 \mathrm{H}$-tetrazole, $\mathrm{DCM}, 23^{\circ} \mathrm{C}, 2 \mathrm{~h}, m-\mathrm{CPBA},-40^{\circ} \mathrm{C}, 1 \mathrm{~h}$.

### 1.3.4. Protection of myo-inositol derived 1,6-diols

The observed regioselectivity for the O-alkylation of 1,6-diols appears to be dependent on the reagents used. Dibutyltin oxide mediated benzylation ${ }^{48}$ of $\mathbf{1 6 7}$ (Scheme 1.23) is reported to occur exclusively at the C1-position in the presence of cesium fluoride while benzylation ${ }^{49}$ in the absence of cesium fluoride afforded minor amounts of the C6-ether along with the C1-ether. Allylation ${ }^{50}$ of the optically pure 1,6-diol D173 via the stannylene derivative gave the C1- and C6-allyl ethers in the ratio 3:1 while the same products were obtained in the ratio 7:1 (estimated by HPLC, total yield $56 \%$ ) on using sodium hydride as the base. Acylation ${ }^{51}$ of 1,6-diols in the presence of tertiary amines occurred predominantly at the C1-position.


Scheme 1.23. a) $\mathrm{Bu}_{2} \mathrm{SnO}, \mathrm{BnBr}$, toluene, $80 \%$ (169); b) $\mathrm{BzCl}, \mathrm{Pyr},-30^{\circ} \mathrm{C}$;
c) $\mathrm{Bu}_{2} \mathrm{SnO}, \mathrm{BnBr}, \mathrm{CsF}, 91-97 \%(171)$; d) $\left(\mathrm{Bu}_{3} \mathrm{Sn}\right)_{2} \mathrm{O}$, toluene, (-)-MntO-COCl, $\mathrm{NaHCO}_{3}$, $70 \%$; e) $\mathrm{Bu}_{2} \mathrm{SnO}$, AllBr, toluene; f) NaH , AllBr, THF.

### 1.3.5. Protection of myo-inositol derived 4(6),5-diols

Alkylation ${ }^{5 \mathrm{~b}, 52}$ of 4,5-diols results in the formation of a mixture of products, the C4-ether being the major constituent (Scheme 1.24). However, as observed in the case of 1,6-diols, the use of cesium fluoride resulted in the formation of C4(6)-ether (D179) exclusively. ${ }^{53}$ Benzoylation ${ }^{54}$ of the diol $\mathbf{1 8 3}$ gave a mixture of products but acylation ${ }^{37 \mathrm{a}}$ of the silyl protected diol $\mathbf{1 8 6}$ gave the C4-ester $\mathbf{1 8 7}$ in $84 \%$ yield. The better regioselectivity observed for the acylation of the silyl ether $\mathbf{1 8 6}$ could be due to the steric hindrance of the bulky silyl ether for the reaction at the C5-hydroxyl group.

$176 \mathrm{H} \quad \mathrm{Bn}(50)$
177 Bn H (39)



Scheme 1.24. a) $\mathrm{Bu}_{2} \mathrm{SnO}$, toluene, $\mathrm{BnBr}, \mathrm{Bu}_{4} \mathrm{NBr}, 70^{\circ} \mathrm{C}$; b) (i) $\mathrm{Bu}_{2} \mathrm{SnO}$, toluene;
(ii) $\mathrm{BnBr}, \mathrm{CsF}, \mathrm{DMF},-41$ to $23^{\circ} \mathrm{C}$; c) $\mathrm{NaH}, \mathrm{PMBCl}, \mathrm{DMF}$; d) $\mathrm{BzCl}, \mathrm{Pyr}$;
e) $\mathrm{CH}_{3} \mathrm{CO}\left(\mathrm{CH}_{2}\right)_{2} \mathrm{COOH}, \mathrm{DCC}$, DMAP.

### 1.3.6. Protection of myo-inositol derived 2,6(4)-diols

Fraser-reid et al reported an extensive study on the alkylation ${ }^{19,55}$ and acylation ${ }^{55}$ of the 2,6(4)-diols 73 and $\mathbf{1 8 8}$ (Scheme 1.25). Benzylation and allylation on the diols $\mathbf{7 3}$ and $\mathbf{1 8 8}$ resulted in predominant alkylation at the C2-hydroxyl group along with minor amount of dialkylated products. Acylation in the presence of pyridine however resulted in the formation of the C6-ester.


Scheme 1.25. a) $\mathrm{NaH}, \mathrm{BnBr}, \mathrm{Bu}_{4} \mathrm{NI}, \mathrm{DMF},-5$ to $0^{\circ} \mathrm{C}, 1-2 \mathrm{~h}$; b) $\mathrm{NaH}, \mathrm{AllBr}, \mathrm{Bu}_{4} \mathrm{NI}, \mathrm{DMF}, 0^{\circ} \mathrm{C}$,
 e) $\mathrm{BzCl}(3 \mathrm{eq}), \mathrm{DMAP}, \mathrm{Pyr}, 0^{\circ} \mathrm{C}, 1 \mathrm{~h}$.

They also carried out glycosidation ${ }^{56}$ of $2,6(4)$ diols (Scheme 1.26), since C2and / or C6-mono and diglycosylated inositols occur in inositides of glycosylphosphatidylinositols (GPIs) and lipoarabinomannans (LAMs), the biological "warheads" of malaria and tuberculosis cell surface oligosaccharides, respectively. Reaction of the diol 188 with n-pentenyl glycoside (NPG) 196 occurred at the C2hydroxyl group to give $\alpha$-mannosides 197 and 198 in the ratio 3:1 (65\% overall yield). In contrast when n-pentenyl orthoester 194 was used for glycosylation of the diol $\mathbf{1 8 8}$ the reaction occurred at the C6-hydroxyl group.


Scheme 1. 26. a) 188, NIS (1.3 eq), TBSOTf (cat), DCM, $0^{\circ} \mathrm{C}, 20 \mathrm{~min}$; b) $\mathbf{1 8 8}$, NIS (1.3 eq), TBSOTf (cat), DCM, rt, 10 min .

In myo-inositol-1,3,5-orthoester derived 2,6(4)-diols (Scheme 1.27), due to inversion of the inositol ring, C2-hydroxyl group is present in the equatorial
orientation and the C6(4)-hydroxyl group is present in the axial orientation (with respect to the myo-inositol ring). Benzylation ${ }^{29 \mathrm{a}}$ of the 2,6(4)-diol 199 with sodium hydride resulted in the formation of a mixture of products $(\mathbf{1 0 2}, \mathbf{1 0 6})$ in the ratio 5:1. Acid catalyzed benzylation ${ }^{31}$ of the 2,6(4)-diol 199 with benzyltrichloroacetimidate resulted in the formation of the C2-ether 106. Similar results were obtained on acylation ${ }^{28 b}$ of 2,4-diol $\mathbf{2 0 0}$ and silylation ${ }^{57}$ of the deoxy derivatives $\mathbf{2 0 1}$ and 202.


|  | $R^{4}$ | $R$ | $R^{2}$ | $R^{4}$ | $R^{6}$ |
| :--- | :--- | :--- | :--- | :--- | :--- |
| 199 H | OBn | 102 H | H | OBn | Bn |
| 200 H | OBOM | 106 H | Bn | OBn | H |
| 201 Bu | F | $\mathbf{2 0 3 H}$ | PNB | OBOM | H |
| 202 Bu | H | 204 Bu | TBS | F | H |
|  |  | 205 Bu | TBS | H | H |

Scheme 1. 27. a) Benzyl trichloroacetimidate, TfOH, DCM, cyclohexane, $81 \%$ (106); b) PNBCI, Pyr, DMAP, $0^{\circ} \mathrm{C}, 12 \mathrm{~h}, 78 \%$ (203); c) TBSOTf, Pyr, DCM, 0 to $23^{\circ} \mathrm{C}$, 75-100\% (204); d) TBSOTf, 2,6-lutidine, DMF, 81\% (205).

### 1.4. Conclusions

The literature available on the synthetic chemistry of myo-inositol and its derivatives suggests that there are no methods available for derivatizing a particular hydroxyl group selectively. There are only a few instances (listed below) of obtaining exclusively a single product on the reaction of myo-inositol or its partially protected derivatives. (a) protection of myo-inositol with 2,2,3,3,-tetramethoxybutane ${ }^{14}$ or TIPDS chloride ${ }^{15}$ (Scheme 1.7); (b) Formation of myo-inositol orthoesters ${ }^{16}$ from myo-inositol (Scheme 1.8); (c) enantioselective phosphorylation ${ }^{24}$ of the 1,3,5-triol 76 in the presence of certain low molecular weight peptides (Scheme 1.11); (d) mono acylation ${ }^{28 a}$ of myo-inositol orthoesters (Scheme 1.14); (e) mono alkylation ${ }^{29}$ of myo-
inositol orthoesters (Scheme 1.14); (f) alkylation ${ }^{48}$ of the 1,6-diol 167 using dibutyltin oxide in the presence of cesium fluoride (Scheme 1.23). The general trend of reactivity among hydroxyl groups observed in myo-inositol (or its derivatives) is $\mathrm{C} 1(\mathrm{C} 3)>\mathrm{C} 4(\mathrm{C} 6)>\mathrm{C} 5>\mathrm{C} 2$ except in myo-inositol 1,3,5-orthoesters, where the reactivity pattern is $\mathrm{C} 4>\mathrm{C} 2 \sim \mathrm{C} 6$. Although a general pattern of selectivity is observed in the reactivity of myo-inositol hydroxyl groups, the selectivity could be altered in some derivatives by manipulating the protecting groups. Differences in reactivity between axial and equatorial hydroxyl groups are reasonably well understood in terms of conformation and steric hindrance, but there are several exceptions to this. However, it is clear that relative reactivities of the inositol hydroxyl groups are often dependent on factors such as intramolecular and intermolecular interactions, especially hydrogen bonding and metal ion chelation effects which are influenced by reaction conditions and other reagents / solvents present in the reaction mixture. Furthermore, a chemical modification at a given position on the inositol ring could induce reactivity changes in other functional groups in the ring due to charge distributions, hydrogen bonding, conformational change, chelation, steric bulk, etc. which could result in change of relative reactivities of the hydroxyl groups. Hence it appears that there is a need for the development of a methodology for the efficient protection of myo-inositol hydroxyl groups wherein each step provides a single regioisomer. Such a method would provide easy and short routes for the protection of desired inositol hydroxyl groups and provide precursors for the synthesis of inositol derivatives. Such attempts carried out in our laboratory are described in the rest of this thesis.

### 1.5. References

1. Jarowicki, K.; Kocienski, P. J. Chem. Soc., Perkin Trans. 1 2001, 2109.
2. Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Das, T. Chem. Rev. 2003, 103, 4477.
3. For example see (a) Thomas, J. R.; Dwek, R. A.; Rademacher, T. W. Biochemistry 1990, 29, 5413; (b) Phosphoinositides: Chemistry, Biochemistry and Biomedical Applications, Bruzik, K. S. Ed.; ACS Symposium Series 718. American Chemical Society, Washington D. C. USA, 1999; (c) Chida, N.; Ogawa, S. Chem. Commun. 1997, 807; (d) Suzuki, T.; Suzuki, S. T.; Yamada, I.; Koashi, Y.; Yamada, K.; Chida, N. J. Org. Chem. 2002, 67, 2874.
4. For example see: (a) Stereochemistry of Organic Compounds by Eliel, E. L.; Wilen, S. H.; Mander, L. N. John Wiley and Sons, Inc. 2003; (b) Ravikumar, K. S.; Farquhar, D. Tetrahedron Lett. 2002, 43, 1367.
5. (a) Schlueter, U.; Lu, J.; Fraser-Reid, B. Org. Lett. 2003, 5, 255; (b) Garegg, P. J.; Berg, B. L.; Kvarnström, I.; Svensson, S. C. T. Carbohydr. Res. 1988, 173, 205.
6. The Inositol Phosphates. Chemical Synthesis and Biological Significance, Billington, D. C. VCH, New York, N.Y. 1993.
7. (a) Chida, N.; Ogawa, S. Chem. Commun. 1997, 807; (b) Tse, B.; Kishi, Y. J. Am. Chem. Soc. 1993, 115, 7892; (c) Gauthier, D. R.; Bender, S. L. Tetrahedron Lett. 1996, 37, 13.
8. (a) Akiyama, T.; Horiguchi, N.; Ida, T.; Ozaki, S. Chem. Lett. 1995, 975; (b) Akiyama, T.; Hara, M.; Fuchibe, K.; Sakamoto, S.; Yamaguchi, K. Chem. Commun. 2003, 1734.
9. Nomenclature Committee-IUB, Biochem. J. 1989, 258, 1.
10. (a) Angyal, S. J.; Macdonald, C. G. J. Chem. Soc. 1952, 686; (b) Angyal, S. J.; Tate, M. E.; Gero, S. D. J. Chem. Soc. 1961, 4116; (c) Garegg, P. J.; Iversen, T.; Johansson, R.; Lindberg, B. Carbohydr. Res. 1984, 130, 322; (d) Bruzik, K. S.; Salamończyk, G. M. Carbohydr. Res. 1989, 195, 67.
11. (a) Gigg, R.; Warren, C. D. J. Chem. Soc. (C). 1969, 2367; (b) Gigg, J.; Gigg, R.; Payne, S.; Conant, R. Carbohydr. Res. 1985, 142, 132.
12. Bruzik, к. s.; Tsai, M. D. J. Am. Chem. Soc. 1992, 114, 6361.
13. Downham, R.; Edwards, P. J.; Entwistle, D. A.; Hughes, A. B.; Kim, K. S.; Ley, S. V. Tetrahedron: Asymmetry 1995, 6, 2403.
14. Montchamp, J-L.; Tian, F.; Hart, M. E.; Frost, J. w. J. Org. Chem. 1996, 61, 3897.
15. Watanabe, Y.; Mitani, M.; Morita, T.; Ozaki, S. J. Chem. Soc., Chem. Commun. 1989, 482.
16. Lee, H. W.; Kishi, Y. J. Org. Chem. 1985, 50, 4402.
17. Watanabe, Y.; Shinohara, T.; Fujimoto, T.; Ozaki, S. Chem. Pharm. Bull. 1990, 38, 562.
18. Chen, J.; Dormán, G.; Prestwich, G. D. J. Org. Chem. 1996, 61, 393.
19. Jia, Z. J.; Olsson, L.; Fraser-Reid, B. J. Chem. Soc., Perkin Trans. 1 1998, 631.
20. Bruzik, K. S.; Kubiak, R. J. Tetrahedron Lett. 1995, 36, 2415.
21. Westerduin, P.; Willems, H. A. M.; van Boeckel, C. A. A. Carbohydr. Res. 1992, 234, 131.
22. Chung, S-K.; Chang, Y-T. Bioorg. Med. Chem. Lett. 1997, 7, 2715.
23. Dinkel, C.; Moody, M.; Traynor-Kaplan, A.; Schultz, C. Angew. Chem., Int. Ed. 2001, 40, 3004.
24. (a) Sculimbrene, B. R.; Miller, s. J. J. Am. Chem. Soc. 2001, 123, 10125; (b) Sculimbrene, B. R.; Morgan, A. J.; Miller, S. J. J. Am. Chem. Soc. 2002, 124, 11653.
25. Kubiak, R. J.; Bruzik, k. s. J. Org. Chem. 2003, 68, 960.
26. (a) Watanabe, Y.; Yamamoto, T.; Ozaki, s. J. Org. Chem. 1996, 61, 14; (b) Watanabe, Y.; Yamamoto, T.; Okazaki, T. Tetrahedron 1997, 53, 903.
27. Matsumura, Y.; Maki, T.; Tsurumaki, K.; Onomura, O. Tetrahedron Lett. 2004, 45, 9131.
28. (a) Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2000, 41, 4185; (b) Ozaki, S.; Koga, Y.; Ling, L.; Watanabe, Y.; Kimura, Y.; Hirata, M. Bull. Chem. Soc. Jpn. 1994, 67, 1058.
29. (a) Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I. M.; Vacca, J. P.; Jane deSolms, S.; Huff, J. R. J. Chem. Soc., Perkin Trans. 1 1989, 1423; (b) Billington, D. C.; Baker, R. J. Chem. Soc., Chem. Commun. 1987, 1011; (c) Jane deSolms, S.; Vacca, J. P.; Huff, J. R. Tetrahedron Lett. 1987, 28, 4503.
30. (a) Riley, A. M.; Murphy, C. T.; Lindley, C. J.; Westwick, J.; Potter, B. V. L. Bioorg. Med. Chem. Lett. 1996, 6, 2197; (b) Ballereau, S.; Poirier, S. N.; Guillemette, G.; Spiess, B.; Schlewer, G. J. Chem. Soc., Perkin Trans. 1 1998, 1859.
31. Baudin, G.; Glänzer, B. I.; Swaminathan, K. S.; Vasella, A. Helv. Chem. Acta. 1988, 71, 1367.
32. Banerjee, T.; Shashidhar, M. S. Tetrahedron Lett. 1994, 35, 8053.
33. Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2001, 42, 3037.
34. (a) Angyal, S. J.; Tate, M. E. J. Chem. Soc. 1965, 6949; (b) Nashed, M. A.; Anderson, L. Tetrahedron Lett. 1976, 17, 3503; (c) Roemer, S.; Stadler, C.;

Rudolf, M. T.; Jastorff, B.; Schultz, C. J. Chem. Soc., Perkin Trans. 1 1996, 1683;
(d) Gou, D-M.; Chen, C-S. J. Chem. Soc., Chem. Commun. 1994, 2125; (e) Aneja, S. G.; Parra, A.; Stoenescu, C.; Xia, W.; Aneja, R. Tetrahedron Lett. 1997, 38, 803; (f) Wang, D-s.; Chen, c-s. J. Org. Chem. 1996, 61, 5905; (g) Gou, D-M.; Chen, C-S. Tetrahedron Lett. 1992, 33, 721; (h) Cottaz, S.; Brimacombe, J. S.; Ferguson, M. A. J. J. Chem. Soc., Perkin Trans. 1 1993, 2945; (i) Riley, A. M.; Potter, B. V. L. Chem. Commun. 2000, 983.
35. (a) Ozaki, S.; Watanabe, Y.; Ogasawara, T.; Kondo, Y.; Shiotani, N.; Nishii, H.; Matsuki, T. Tetrahedron Lett. 1986, 27, 3157; (b) Solomons, K. R. H.; Freeman, S.; Poyner, D. R.; Yafai, F. J. Chem. Soc., Perkin Trans. 1 1996, 1845; (c) Seitz, S. P.; Kaltenbach III, R. F.; Vreekamp, R. H.; Calabrese, J. C.; Perrella, F. W. Bioorg. Med. Chem. Lett. 1992, 2, 171; (d) Fauq, A. H.; Kozikowski, A. P.; Ognyanov, V. I.; Wilcox, R. A.; Nahorski, S. R. J. Chem. Soc., Chem. Commun. 1994, 1301.
36. Dreef, C. E.; Elie, C. J. J.; Hoogerhout, P.; van der Marel, G. A.; van Boom, J. H. Tetrahedron Lett. 1988, 29, 6513.
37. (a) Watanabe, Y.; Hirofuji, H.; Ozaki, S. Tetrahedron Lett. 1994, 35, 123; (b) Watanabe, Y.; Tomioka, M.; Ozaki, S. Tetrahedron 1995, 51, 8969; (c) Reddy, K. K.; Rizo, J.; Falck, J. R. Tetrahedron Lett. 1997, 38, 4729.
38. Angyal, S. J.; Gilham, P. T.; Melrose, G. J. H. J. Chem. Soc. 1965, 5252.
39. (a) Chida, N.; Koizumi, K.; Kitada, Y.; Yokoyama, C.; Ogawa, S. J. Chem. Soc., Chem. Commun. 1994, 111; (b) Mills, S. J.; Potter, B. V. L. J. Chem. Soc., Perkin Trans. 1 1997, 1279; (c) Riley, A. M.; Potter, B. V. L. Tetrahedron Lett. 1999, 40, 2213; (d) Mills, s. J.; Potter, B. v. L. J. Org. Chem. 1996, 61, 8980; (e) Ozaki, S.;

Kondo, Y.; Nakahira, H.; Yamaoka, S.; Watanabe, Y. Tetrahedron Lett. 1987, 28, 4691.
40. (a) Andersch, P.; Schneider, M. P. Tetrahedron: Asymmetry 1996, 7, 349; (b) Dinkel, C.; Schultz, C. Tetrahedron Lett. 2003, 44, 1157.
41. Migaud, м. E.; Frost, J. w. J. Am. Chem. Soc. 1995, 117, 5154.
42. (a) Kulagowski, J. J. Tetrahedron Lett. 1989, 30, 3869; (b) Vacca, J. P.; Jane deSolms, S.; Huff, J. R.; Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I. M. Tetrahedron 1989, 45, 5679; (c) Liu, C.; Potter, B. V. L. Tetrahedron Lett. 1994, 35, 1605; (d) Liu, C.; Potter, B. v. L. J. Org. Chem. 1997, 62, 8335; (e) Gigg, J.; Gigg, R.; Payne, S.; Conant, R. J. Chem. Soc., Perkin Trans. 1 1987, 423.
43. Elie, C. J. J.; Verduyn, R.; Dreef, C. E.; Brounts, D. M.; van der Marel, G. A.; van Boom, J. H. Tetrahedron 1990, 46, 8243.
44. Aneja, R.; Aneja, S. G.; Parra, A. Tetrahedron Lett. 1996, 37, 5081.
45. Verónica, S-P.; Francisco Javier, M-M.; Rosalinda, C.; Angelina, F-P. J Carbohydr. Chem. 1997, 16, 1479.
46. Schmitt, L.; Spiess, B.; Schlewer, G. Tetrahedron Lett. 1998, 39, 4817.
47. Falck, J. R.; Krishna, U. M.; Katipally, K. R.; Capdevila, J. H.; Ulug, E. T. Tetrahedron Lett. 2000, 41, 4271.
48. (a) Yu, K-L.; Fraser-Reid, B. Tetrahedron Lett. 1988, 29, 979; (b) Liu, Y-C.; Chen, C-S. Tetrahedron Lett. 1989, 30, 1617.
49. Desai, T.; Gigg, J.; Gigg, R.; Payne, S. Carbohydr. Res. 1992, 225, 209.
50. Baeschlin, D. K.; Chaperon, A. R.; Green, L. G.; Hahn, M. G.; Ince, S. J.; Ley, S. V. Chem. Eur. J. 2000, 6, 172.
51. (a) Kratzer, B.; Mayer, T. G.; Schmidt, R. R. Tetrahedron Lett. 1993, 34, 6881;
(b) Mihai, C.; Mataka, J.; Riddle, S.; Tsai, M-D.; Bruzik, K. S. Bioorg. Med. Chem. Lett. 1997, 7, 1235.
52. Fauq, A. H.; Zaidi, J. H.; Wilcox, R. A.; Varvel, G.; Nahorski, S. R.; Kozikowski, A. P.; Erneux, C. Tetrahedron Lett. 1996, 37, 1917.
53. Gou, D-M; Liu, Y-C; Chen, C-S. Carbohydr. Res. 1992, 234, 51.
54. Chung, S-K.; Chang, Y-T.; Sohn, K-H. Chem. Commun. 1996, 163.
55. Anilkumar, G.; Jia, Z. J.; Kraehmer, R.; Fraser-Reid, B. J. Chem. Soc., Perkin Trans. 1 1999, 3591.
56. Anilkumar, G.; Nair, L. G.; Fraser-Reid, b. Org. Lett. 2000, 2, 2587.
57. (a) Biamonte, M. A.; Vasella, A. Helv. Chim. Acta. 1998, 81, 695; (b) Biamonte, M. A.; Vasella, A. Helv. Chim. Acta. 1998, 81, 688.

## 2A.1. Introduction

A survey of the literature (Chapter 1) revealed that although many myoinositol derivatives have been synthesized, there is no general method that is flexible enough to be adopted for the preparation of a variety of inositol derivatives. Also, most of the procedures used for the derivatization of inositol hydroxyl groups yield mixture of products from which the desired product is separated. ${ }^{1}$ The overall yield of the required O-protected myo-inositol derivative is usually not good due to: (a) formation of isomeric myo-inositol derivatives and / or (b) use of transient protecting groups (esters, sulfonates, allyl ether, etc.) which increase the number of steps and decrease the overall yield. Hence we have attempted to devise a methodology for the convenient preparation of inositol derivatives with the formation of minimum number of products. In other words, we set out to device methods for the regioselective derivatization of inositol hydroxyl groups. From previous experience in our laboratory as well as those reported in the literature, it was clear that regioselective derivatization of any one of the six hydroxyl groups in myo-inositol is not easy to achieve in the laboratory, although such reactions do occur in living cells. ${ }^{2}$ Hence we envisaged regioselective derivatization of two myo-inositol derived triols obtainable easily from myo-inositol that would allow us to investigate regioselective derivatization of all the myo-inositol hydroxyl groups. We chose myo-inositol-1,3,5orthoformate (65, a 2,4,6-triol) and 2,4,6-tri-O-benzyl-myo-inositol (76, a 1,3,5-triol) for our investigations. Development of methods for the regioselective derivatization of the hydroxyl groups in these two triols would in principle allow convenient access to a variety of O-protected myo-inositol derivatives (Scheme 2A.1). Regioselective functionalization of myo-inositol orthoesters are being attempted ${ }^{3}$ and appear to show
promise for the synthesis of biologically relevant molecules. The present section deals with the regioselective O-substitution in myo-inositol orthoesters.


Scheme 2A. 1
Formation of orthoesters of myo-inositol involves inversion of the carbocyclic ring. This results in five axial and one equatorial oxygen atoms on the carbocyclic ring. These orthoesters are in fact trioxa analogs of adamantane and hence have very rigid molecular frame. Due to the presence of two 1,3-diaxial hydroxyl groups which are strongly hydrogen bonded to each other, acidity of the C4- and C6-hydroxyl groups is higher than that of the equatorial C2-hydroxyl group (Scheme 2A.2).


Scheme 2A. 2
This difference in acidity of the hydroxyl groups of myo-inositol orthoesters can be exploited for their regioselective derivatization. The existence of hydrogen bond between the C4- and C6-hydroxyl groups in myo-inositol orthoesters was
established by NMR spectroscopy as well as X-ray crystal structures of myo-inositol orthoesters. ${ }^{4}$


Scheme 2A. 3

The differences in the observed reactivity of the three hydroxyl groups (and hence the observed regioselectivity) of myo-inositol orthoesters have been attributed to various factors ${ }^{3}$ shown in Scheme 2A. 3 and other parameters such as nature of the electrophile, ${ }^{3 a}$ strength of the base used, ${ }^{3 a, b, 5}$ solvent used for the reaction, ${ }^{3 b, c}$ presence or absence of metal ions, ${ }^{3 c, d}$ steric bulk of the reagent ${ }^{3 a, e}$ and the catalyst used. ${ }^{6}$ These variations were utilized to obtain either mono-O-substituted derivatives (99, 210-215, Scheme 2A.3) or di-O-substituted derivatives (216-219) in poor to good isolated yields. However, there is no general method for the orthogonal protection of the three hydroxyl groups of myo-inositol orthoesters (especially as ethers) which generates
only one regioisomer in each step, in respectable yields. Such a method would provide orthogonally protected myo-inositol orthoesters, which are the key intermediates for the synthesis of biologically important inositol derivatives. The following section describes a general method for the completely regioselective protection of the three secondary hydroxyl groups of myo-inositol orthoesters, utilizing the subtle differences in reactivity exhibited by its alkali metal alkoxides.

## 2A.2. Results and Discussion

Mono O-substitution of the axial C4-hydroxyl group in myo-inositol orthoesters can be achieved with very high regioselectivity. Mono O-alkylation ${ }^{3 \mathrm{c}}$ and O-acylation ${ }^{3 d}$ of myo-inositol orthoesters $\mathbf{6 5}$ and $\mathbf{9 8}$ in the presence of sodium hydride was thought to be due to the involvement of chelates such as $\mathbf{2 0 8}$ (Scheme 2A.3). ${ }^{4 \mathrm{c}, 7}$ However, acylation with acyl halides in the presence of tertiary amines (e.g. triethylamine, $\mathrm{pKa}_{\mathrm{H}}=10.8^{8}$ ) is also known ${ }^{3 \mathrm{a}, 5}$ to give the corresponding C4-Oacylated derivatives (Scheme 2A.4).


Scheme 2A.4. a) NaH , alkyl halide or acyl halide, DMF ; b) $\mathrm{Et}_{3} \mathrm{~N}$, acyl halide, DMF ;
c) NaH , DMF; d) $\mathrm{NaH}(2 \mathrm{eq}$ ), acyl halide (1 eq), DMF; e) TBSCI, imidazole, DMF; f) acyl halide, Pyr.

Weaker bases bring about substitution at the C2-O-position as exemplified by the benzoylation ${ }^{3 a}$ of $\mathbf{6 5}$ in the presence of pyridine $\left(\mathrm{pKa}_{\mathrm{H}}=5.2^{8}\right)$. Similarly, alkylation of $\mathbf{6 5}$ under phase transfer catalytic conditions ${ }^{9}$ and silylation ${ }^{3 f}$ of $\mathbf{6 5}$ in the presence of imidazole also result in the formation of the corresponding C2-O-substituted derivative as the major product. The predominant formation of the C2-O-substituted derivatives in the presence of weaker bases could be due to the relatively higher nucleophilicity of the $\mathrm{C} 2-\mathrm{OH}$ as compared to the $\mathrm{C} 4(6)-\mathrm{OH}$ under unionized conditions, due to the involvement of the latter in intramolecular H -bonding (Scheme 2A.2). Hence it was of interest to examine the regioselectivity observed (exclusive reaction at the C4-O-position) during the mono-O-substitution ${ }^{3 \mathrm{c}}$ of the triol ( $\mathbf{6 5}$ or $\mathbf{9 8}$ ) in the presence of other bases.

The triol 65 did not undergo alkylation with benzyl bromide in the presence of DBU. Benzylation of $\mathbf{6 5}$ in the presence of potassium carbonate resulted in exclusive alkylation at the C4-O-position to yield the benzyl ether 199 in $72 \%$ isolated yield (Scheme 2A.5).


Scheme 2A.5. a) DMF, $\mathrm{K}_{2} \mathrm{CO}_{3}$, BnBr .
This result along with the formation of C4-esters 214 in the absence of metal ions ${ }^{32}$ imply that chelation with metal ions is not a necessary condition for the mono-O-substitution at the C4-O-position in the triol 65. The observed regioselectivity can be rationalized by considering the intramolecular hydrogen bonding that is present ${ }^{4}$ in these triols, which results in an increase of the acidity of the C4(6)-hydroxyl group. ${ }^{3 \mathrm{~g}, 10}$ If the base used is strong enough to form even minor amounts of the anion
at the C4-O-position, which is also stabilized by intramolecular hydrogen bonding (207, Scheme 2A.3), reaction brought about by such bases result in substitution at the C4-O-position. Hence, during the mono-O-substitution of these triols ( 65 and 98 ), in our opinion, there is no need to invoke either chelation (Scheme 2A.3, 208) with metal ions or steric effect of the two axial hydroxyl groups ${ }^{3 a}$ to rationalize the experimental observations, since hydrogen present in the cis-axial hydroxyl group is a good acceptor of hydrogen bond which could stabilize the anion (Scheme 2A.3, 207). That the steric bulk of the electrophile does not contribute much to the observed regioslectivity for the reaction with orthoesters was evident by the fact that the C4trityl ether $\mathbf{2 2 3}$ could be obtained by tritylation of the triol $\mathbf{6 5}$ using sodium hydride as the base (Scheme 2A.6).


Scheme 2A.6. a) $\mathrm{NaH}, \mathrm{TrCl}, \mathrm{DMF}$.
Although O-alkylation ${ }^{3 c}$ of myo-inositol orthoesters (such as $\mathbf{6 5}$ or 98 ) with alkyl halides in the presence of one equivalent of sodium hydride resulted in exclusive reaction at the $\mathrm{C} 4(6)$-O-position, ${ }^{3 \mathrm{c}, 10}$ reaction of these triols with two equivalents of alkyl halide ${ }^{3 c, h, i}$ in the presence of two equivalents of sodium hydride resulted in the formation of a mixture of 2,4- and 4,6-diethers. However, isolated yield of the major product, the 4,6-diether $\mathbf{2 2 4}$ was poor (Scheme 2A.7).


Scheme 2A.7. a) NaH (2 eq), alkyl halide (2 eq), DMF.
Reaction of the C4-monoether (such as 99, Scheme 2A.4) with alkyl halide in the absence of metal ions provided the unsymmetrical diether (such as 225) as the major product. ${ }^{9}$ These results give credence to the postulated chelation ${ }^{3 \mathrm{c}}$ of the metal ion with 4,6-diaxial oxygens, during O-substitution of the axial monoether 99, since acidity of the two hydroxyl groups in this monoether is expected to be comparable in the absence of the strong intramolecular hydrogen bond ${ }^{4}$ as present in the corresponding triols ( 65 and 98). Vasella ${ }^{4 \mathrm{a}}$ has reported that the axial hydroxyl group in C4-monobenzyl ether $\mathbf{1 9 9}$ does form intramolecular hydrogen bond with the axial hydroxyl group, but this is not as strong as the intramolecular hydrogen bond between the two axial hydroxyl groups in the triol 65. Similarly crystal structure of C4-esters (such as 214, Scheme 2A.4) showed that the axial hydroxyl group is involved in intermolecular hydrogen bonding rather than in intramolecular hydrogen bonding. ${ }^{11}$

We postulated that if chelation of the metal ions was indeed operational during O-substitution reactions of C4-ethers (such as 199), the use of lithium alkoxides instead of sodium alkoxides must result in better regioselectivity, resulting in the formation of greater amounts of the 4,6-di-O-substituted derivatives $\mathbf{2 2 4}$ as compared to the 2,4-di-O-substituted derivatives $\mathbf{2 2 5}$ (Scheme 2A.7). This is expected since lithium, being smaller and a better ion for co-ordination ${ }^{12}$ with oxygen atoms, would stabilize the oxyanion at the C6(4)-O-position in axial C4(6)-O-monosubstituted myoinositol orthoesters (such as 99, Scheme 2A.4).

Reaction of the triol $\mathbf{6 5}$ with two equivalents of butyl lithium and benzyl bromide ( 2 eq ) in THF resulted in the formation of the C4-benzyl ether 199, the 4,6dibenzyl ether $\mathbf{1 0 2}$ and a trace of the tribenzyl ether $\mathbf{2 2 6}$ (Scheme 2A.8).


Scheme 2A.8. a) THF, n-BuLi, BnBr.
TLC analysis of the reaction mixture during the progress of the reaction revealed that the monobenzyl ether $\mathbf{1 9 9}$ did not get completely converted to the 4,6dibenzyl ether $\mathbf{1 0 2}$ even under reflux; however, formation of the 2,4-dibenzyl ether 106 (Scheme 1.15, Chapter 1) was not observed. Incomplete conversion of $\mathbf{1 9 9}$ to $\mathbf{1 0 2}$ could have been due to the formation of lithium-inositol derivative aggregates in THF which could retard further reaction. Literature reports show that lithium enolates do form aggregates in THF and the rate of the reaction of these aggregates with the alkyl halides is sluggish. ${ }^{13}$ Hence in future experiments DMF was used as a co-solvent to break the purported lithium-inositol derivative aggregates and increase the rate of the alkylation reaction.

Reaction of the triol $\mathbf{6 5}$ with two equivalents of butyl lithium and benzyl bromide (2 eq) in a mixture of THF and DMF resulted in the formation of the symmetrical diether $\mathbf{1 0 2}$ (Scheme 2A.9) and traces of the tribenzyl ether 226. Since we isolated the tri-O-benzyl ether 226 but no 2,4-di-O-benzyl derivative 106, it appears that $\mathbf{2 2 6}$ resulted from benzylation of the initially formed 4,6-dibenzyl ether 102.


Scheme 2A.9. a) n-BuLi (2 eq), THF, BnBr or AllBr in DMF; b) $\mathrm{DMF}, \mathrm{LiH}(4 \mathrm{eq}), \mathrm{BnBr}$ or AllBr .

Dibenzylation of the triol $\mathbf{6 5}$ with two equivalents of lithium hydride in DMF resulted in the formation of the C4-monobenzyl ether 199 and the 4,6-dibenzyl ether 102 but the reaction did not go to completion (as revealed by TLC analysis of the reaction mixture). But again, formation of the 2,4-dibenzyl ether 106 was not observed. When benzylation of $\mathbf{6 5}$ was carried out using four equivalents of lithium hydride in DMF, the symmetrical 4,6-dibenzyl ether $\mathbf{1 0 2}$ was obtained exclusively. Hence use of BuLi or lithium hydride for the benzylation of the triol $\mathbf{6 5}$ resulted in O substitution at the C4-hydroxyl group first followed by O-substitution at the C6hydroxyl group; O-substitution at the C2-hydroxyl group was not observed.

The 4,6-di-O-benzyl orthoacetate 227 and the di-O-allyl orthoformate $\mathbf{1 0 4}$ could also be obtained in good yields using BuLi or lithium hydride as a base for the alkylation of the corresponding triols (Scheme 2A.9). The structure of the dibenzyl orthoacetate 227 was confirmed by comparing its ${ }^{1} \mathrm{H}$ NMR spectrum with that of the dibenzyl orthoformate 102. The only difference observed between the ${ }^{1} \mathrm{H}$ NMR spectra of these two ethers was the presence of the orthoacetate methyl peak instead of the orthoformate proton peak. The diallyl ether 104 was characterized by comparison of its ${ }^{1} \mathrm{H}$ NMR spectrum with that reported ${ }^{3 i}$ in the literature. The literature method of preparation of $\mathbf{1 0 4}$ consisted of allylation of the triol $\mathbf{6 5}$ using
sodium hydride as the base, which resulted mixture of allyl ethers, from which the 4,6 -diallylether $\mathbf{1 0 4}$ was isolated in $44 \%$ yield. In general, known methods ${ }^{3 c, h, i}$ for the preparation of 4,6-di-O-substituted derivatives of triols $\mathbf{6 5}$ or $\mathbf{9 8}$ either resulted in a mixture of products or were restricted to the preparation of disulfonates ${ }^{3 b}$ or the dibenzyl ether $\mathbf{1 0 2}$ via a circuitous route. ${ }^{3 f, 14}$

Having obtained this interesting result on the di-O-substitution of the diaxial diols directly from a triol, we carried out O-alkylation of the C4-mono substituted derivatives of myo-inositol orthoesters using butyllithium or lithium hydride as a base, to investigate the generality of this reaction. In all such experiments only one regioisomer viz., the C4- and C6-di-O-substituted product was obtained. The results obtained for the regioselective C6-O substitution in a few C4-monosubstituted derivatives of myo-inositol orthoesters are listed in Scheme 2A.10.


Scheme 2A.10. a) n-BuLi / THF; b) DMF, LiH;
c) Alkyl halide or $\mathrm{Ac}_{2} \mathrm{O}$ or TsCl , DMF or $\mathrm{TsCl} / \mathrm{THF}$.

A comparison of the yield of symmetrical 4,6-di-O-substituted derivatives reveals that tosylation of the unsymmetrical diols $\mathbf{1 9 9}$ and $\mathbf{2 3 0}$ are low yielding compared to their alkylation and acylation reactions. However, in all the tosylation reactions carried out, the unsymmetrical 2,4-ditosylate 110 (Scheme 1.15, Chapter 1) was not formed. The low yield of symmetric ditosylates was perhaps due to the lesser polarity
(compared to DMF) of the solvent used for the reaction (THF). The use of a mixture of THF and DMF for the tosylation reactions resulted in the formation of the tritosylate 240. Reasons for these observations are not clearly understood.

Among the 4,6-di-O-substituted derivatives obtained, the dibenzyl ether $\mathbf{1 0 2}^{3 \mathrm{c}}$ and the ditosylate $\mathbf{1 0 5}^{15}$ are previously reported in the literature and hence their structure could be established by comparison of the physical and spectroscopic data available in the literature. The NMR spectra of symmetrically C4- and C6-di-Osubstituted derivatives of myo-inositol orthoesters are quite characteristic due to the presence of a mirror plane in the molecule.

Structure of the allyl ether $\mathbf{2 3 1}$ was confirmed by converting it to the known ${ }^{3 c}$ 2,4-dibenzyl ether 242 (Scheme 2A.11) and comparison of its (242) ${ }^{1} \mathrm{H}$ NMR spectrum with that reported in the literature. One of the characteristics of unsymmetrical benzyl ethers derived from myo-inositol orthoformate is that the two benzylic methylene protons of the axial benzyl ether appear as AB quartet and the benzylic methylene protons of the equatorial benzyl ether appears as a singlet in their ${ }^{1} \mathrm{H}$ NMR spectra. In the case of symmetrical 4,6-dibenzyl ether, benzylic protons of both the benzyl ether groups appear as AB quartet with the same chemical shift.


Scheme 2A.11. a) n-BuLi / THF, AllBr / DMF; b) NaH , BnBr, DMF.
The structure of the orthoformate 6-acetate $\mathbf{2 3 2}$ was confirmed by comparing the chemical shift of the acetate methyl peak in its ${ }^{1} \mathrm{H}$ NMR spectrum with the chemical shifts for the acetate methyl groups in the ${ }^{1} \mathrm{H}$ NMR spectra of known orthoester acetates 245-251 (Chart 2A.1). In the case of C2-acetates (equatorial), the acetate methyl peak appears in the range $\delta 2.20-2.26$ while in the case of $\mathrm{C} 4(6)$ acetates, acetate methyl peak appears in the range $\delta 1.88-2.11$. Since in the ${ }^{1} \mathrm{H}$ NMR spectrum of the acetate 232 the acetate methyl peak appeared at $\delta 1.86$ we concluded that the acetylation had occurred at the C6-hydroxyl group and not at the C2-hydroxyl group.

2.24

246

2.09



Chart 2A.1. Chemical shift of acetate methyl groups in the ${ }^{1} \mathrm{H}$ NMR spectra of acetate derivatives of myo-inositol orthoesters.

The structure of the 6-tosylate $\mathbf{2 3 3}$ was arrived at by comparing the chemical shift of the C6-ring proton (attached to the carbon bearing the tosylate group) in its ${ }^{1} \mathrm{H}$ NMR spectrum with the spectra of its analogs (Chart 2A.2). In the ${ }^{1} H$ NMR spectra of C4- and / or C6-tosylate derivatives of myo-inositol orthoesters the signal due to the (equatorial) hydrogen attached to C 4 and / or C6 always appeared at $\delta>5.00$ whereas for the C 2 -tosylate the signal due to the (axial) hydrogen attached to C 2 appeared at $\delta<5.00$. The appearance of a signal in the ${ }^{1} \mathrm{H}$ NMR spectrum of the tosylate 233 at $\delta 5.19$ showed that tosylation had occurred at the C6-hydroxyl group in 199. This is further supported by the ${ }^{1} \mathrm{H}$ NMR spectrum of the by-product ditosylate $\mathbf{2 3 8}$ in which one ring proton appears at $\delta 5.09$ and another proton appears at $\delta$ 4.93. In the ${ }^{1} \mathrm{H}$ NMR spectra of all these tosylate derivatives the most de-shielded
signals can be attributed to the hydrogen attached to the carbon carrying the tosylate group.


252


238



O Ts 240


110

105

Chart 2A.2. Chemical shift of the $\mathrm{C} 2-\mathrm{H}$ and $\mathrm{C} 6(4)-\mathrm{H}$ in the ${ }^{1} \mathrm{H}$ NMR spectra of tosyltate derivatives of myo-inositol orthoformates.

The structure of the C6-allyl ether 234 was confirmed by solvolysis of the tosylate followed by acetylation, which gave the racemic 2,4-diacetate 256 (Scheme 2A.12). The structure of the unsymmetrical diacetate 256 was established by its ${ }^{1} \mathrm{H}$ NMR spectrum which showed two separate singlets for the acetyl methyl groups. If the product of allylation of the racemic 4 -tosylate $\mathbf{2 2 9}$ was the corresponding 2-allyl ether, solvolysis of the tosylate followed by acetylation would have resulted in the formation of the symmetric diacetate $\mathbf{2 5 9}$ which would exhibit a single peak for the two acetyl methyl groups in its ${ }^{1} \mathrm{H}$ NMR spectrum.


Scheme 2A.12. a) AllBr, LiH, DMF; b) $\mathrm{NaOMe}, \mathrm{MeOH}$, reflux; c) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr.

The structure of the orthoacetate allyl ether $\mathbf{2 3 5}$ was confirmed by comparison of its ${ }^{1} \mathrm{H}$ NMR spectrum with the orthoformate analog 234. The two ${ }^{1} \mathrm{H}$ NMR spectra were similar except that the orthoacetate methyl peak appeared (at $\delta 1.42$ ) in the spectrum of $\mathbf{2 3 5}$ instead of the peak for the orthoformate proton (at $\delta 5.44$ ) in the spectrum of 234.

We also tested the possibility of carrying out the O-substitution of the unsymmetrical 2,4-diol having an ester group at the C4-position (such as 260, Scheme 2A.13). Earlier work ${ }^{3 \mathrm{~d}}$ in our laboratory had shown that C 4 -esters undergo facile isomerization to the corresponding C2-esters when treated with sodium hydride in DMF.


Scheme 2A.13. a) $\mathrm{NaH}, \mathrm{DMF}, 5 \mathrm{~min}$; b) LiH, DMF, 30 min .
Since lithium alkoxides are less reactive as compared to the corresponding sodium alkoxides (through which the isomerization of the C 4 -ester to the C 2 -ester
proceeds), we wondered whether this would allow us to obtain good yield of the axial C6-ethers on alkylation of the racemic C4-benzoate $\mathbf{2 6 0}$ in the presence of lithium hydride. The fact that the lithium alkoxide is less reactive than the corresponding sodium alkoxide was indicated by the experimental observation that the C4-benzoate 260 underwent complete isomerization to the corresponding C2-benzoate 261 in 5 $\min$ (or less) on treatment with sodium hydride while the reaction time was about 30 $\min$ in the presence of lithium hydride (Scheme 2A.13). However, the weaker basicity of lithium hydride (as compared to that of sodium hydride) as a reason for the slower rate of conversion (of the C4-benzoate $\mathbf{2 6 0}$ to the C2-benzoate 261) observed cannot be ruled out. In any case, irrespective of the cause for the difference in rates observed, slower isomerization of the C4-benzoate $\mathbf{2 6 0}$ in the presence of lithium hydride prompted us to attempt its benzylation. Butyllithium could not be used for this reaction to generate the required lithium alkoxide since alkyllithium is known ${ }^{16}$ to react with esters to give the corresponding alkyl ketone.


Scheme 2A.14. a) DMF, $\mathrm{BnBr}, \mathrm{LiH}$; b) DMF, $\mathrm{BnBr}, \mathrm{NaH} ; \mathrm{c}) \mathrm{DMF}, \mathrm{NaH}, \mathrm{BnBr}$.

Benzylation of the C4-benzoate $\mathbf{2 6 0}$ using sodium hydride as a base is known $^{11}$ to yield a mixture of benzyl ethers 262 and 263 in the ratio 1.3:1 (based on isolated yields), provided sodium hydride was added to a solution of $\mathbf{2 6 0}$ and benzyl bromide in DMF (Scheme 2A.14). Addition of benzyl bromide to a mixture of $\mathbf{2 6 0}$ and sodium hydride results in the exclusive formation of the benzyl ether 263.

Reaction of benzyl bromide with the C4-benzoate $\mathbf{2 6 0}$ using lithium hydride as the base yielded 262 and 263 in the ratio 2:1 (based on isolated yields). Although formation of the undesired product 263 was suppressed on using lithium hydride instead of sodium hydride, the isolated yield of the desired product $\mathbf{2 6 2}$ remained same. It is interesting to see that benzylation at the C2-O-position did not occur; this is perhaps because, formation of the C2-alkoxide results in immediate migration of the C4-O-benzoyl group (which is an intramolecular reaction) to the C2-O-position rather than its benzylation (which is an intermolecular reaction). These results also seem to support our contention that in the presence of metal ions, C4-alkoxide is formed preferentially due to their stabilization by chelate formation.

In a separate experiment, we also observed hydrolysis of the benzoate during work-up of the reaction mixture (by quenching lithium hydride with ice and evaporation of DMF at higher temperature $75-80^{\circ} \mathrm{C}$ ) and isolated 263 (12\%) and 199 (73\%). The product $\mathbf{1 9 9}$ can arise directly by the hydrolysis of the benzoate in $\mathbf{2 6 2}$ or 263 and in principle also by hydrolysis of the axial benzoate in $\mathbf{2 6 0}$ to generate the triol 65 followed by its benzylation. The latter route for the formation of 199 can be ruled out since this is possible only during the reaction (before quenching with ice) and if so, we should have obtained comparable yield of $\mathbf{1 9 9}$ in all the experiments irrespective of the work-up procedure. However, these results clearly showed that the use of lithium hydride as a base to bring about C6-O-substitution in C4-esters of myoinositol orthoesters is not of much synthetic utility.

A perusal of the results described in this section shows that, the experimental protocol for best results for the C6-O-substitution in C4-O-protected myo-inositol $1,3,5$-orthoester derivatives is the generation of the lithium alkoxide in dry THF followed by the addition of a solution of the appropriate electrophile in dry DMF.

This method is sufficiently flexible to allow the preparation of 4,6-diethers and any combination of ether, ester (alkylation followed by acylation - see preparation of the acetate 232, Scheme 2A.10) and sulfonate derivatives. This method is not suitable for the preparation of 4,6-diesters since alkoxides of C4(6)-monoesters (214, Scheme 2A.4) undergo isomerization to the corresponding C 2 -esters ${ }^{3 \mathrm{~d}}$ faster than substitution at the C6-O-position. The versatility of this method was demonstrated by the rapid and efficient preparation of an unsymmetrical tri-O-substituted derivative 264 (Scheme 2A.15) from the racemic C4-benzyl ether 199 in a one pot reaction. Hence starting from myo-inositol, an orthogonally protected 2,4,6-tri-O-substituted derivative can be obtained in four steps, wherein each step gives only one product / regioisomer. This method should allow modification of the hydroxyl groups at C 2 , C4 and C6 positions to obtain other inositol derivatives.


Scheme 2A.15. a) DMF, NaH, BnBr; b) n-BuLi / THF, AllBr / DMF; c) NaH, Mel.

We also investigated the extent to which steric hindrance due to the presence of 4,6-diaxial substituents (in $\mathbf{6 5}, \mathbf{9 8}$ and their derivatives) dictate the course of the reaction and contribute to the observed regioselectivity. To understand this, we carried out (a) reaction of the triol $\mathbf{6 5}$ with bulky electrophiles (TBSCl) and (b) Osubstitution of myo-inositol orthoester derivatives carrying a bulky protecting group (trityl) at the C4-O-position, with a small electrophile (methyl iodide).


Scheme 2A.16. a) n-BuLi, TBSCI, THF; b) LiH (4eq), TBSCI (2eq), DMF; c) n-BuLi / THF, BnBr / DMF.

Reaction of the triol $\mathbf{6 5}$ with TBSCl (1 eq.) / BuLi led to the formation of the C4-O-protected derivative 265 as the major product (Scheme 2A.16), and the corresponding C2-silyl ether was not formed. Silylation of $\mathbf{6 5}$ in the presence of metals (palladium) ${ }^{17}$ those are not basic enough to form the C 4 -oxyanion result in a mixture of products consisting of different regioisomeric silyl ethers. Reaction of the triol 65 with two equivalents of $\mathrm{TBSCl} / \mathrm{LiH}$ led to the formation of the racemic 2,4-di-O-silyl derivative 266. This is in contrast to the corresponding reaction of the triol 65 with alkyl halides (not having bulky alkyl groups), which afforded the 4,6-diethers as the major product (Scheme 2A.9). The difference in the observed regioselectivity in these two reactions can be attributed to the steric hindrance offered by the C4-OTBS group for the entry of another bulky TBS group at the C6-O-position.


Scheme 2A.17. a) n-BuLi / THF, Mel / DMF; b) NaH / THF, Mel / DMF.

Reaction of the racemic trityl ether 223 (Scheme 2A.17) with butyllithium followed by methyl iodide gave a mixture of products consisting of isomeric monomethyl ethers 267 and 268 as well as the dimethyl ether 269. The racemic C6ether 267 was formed in higher proportion compared to the corresponding C2-methyl ether $\mathbf{2 6 8}(\mathbf{2 6 7 : 2 6 8}=61: 39)$. The use of sodium hydride for the same reaction also resulted in a mixture of products (267-269) but $\mathbf{2 6 8}$ was present in larger proportion as compared to $\mathbf{2 6 7}(\mathbf{2 6 7 : 2 6 8}=11: 89)$.

The structure of the two monomethyl ethers 267 and 268 were established by unambiguous synthesis of $\mathbf{2 6 8}$ starting from the known ditosylate $\mathbf{1 0 5}$ (Scheme 2A.18). ${ }^{15}$ The ditosylate 105 was methylated using sodium hydride and methyl iodide and the tosyl groups in $\mathbf{2 5 3}$ were cleaved with sodium methoxide in methanol under reflux. The diol $\mathbf{2 7 0}$ so obtained was tritylated to give 268 .


Scheme 2A.18. a) NaH , Mel, DMF; b) $\mathrm{NaOMe}, \mathrm{MeOH}$, reflux; c) $\mathrm{NaH}, \mathrm{TrCl}, \mathrm{DMF}$.
In principle, reaction of myo-inositol orthoester derived 2,4-diols (such as 99, Scheme 2A.4) with butyllithium or sodium hydride can result in the formation of two anions 271 and 273 (Scheme 2A.19). Higher isolated yields of the 4,6-disubstituted products on the reaction of myo-inositol orthoester derived 2,4-diols with electrophiles in the presence of metal hydrides (Scheme 2A.10) suggest that $\mathbf{2 7 1}$ is preferentially formed (see results on alkylation of the C4-benzoate Scheme 2A.14). This could be a result of greater stabilization of the anion 271 by metal ions due to chelation (272) with two diaxially oriented oxygen atoms as compared to the
stabilization of the anion 273 due to chelation (274) with the oxygen atoms in axial equatorial orientation.


Scheme 2A.19. a) NaH or LiH or n -BuLi.
This is analogous to the formation of a strong intramolecular hydrogen bond between the 4,6-diaxial hydroxyl groups in myo-inositol orthoesters (e.g. 65) which has been observed in their crystals, ${ }^{4 \mathrm{a}, \mathrm{b}}$ while the cis-1,2-hydroxyl groups result in a comparatively weaker intramolecular hydrogen bond. ${ }^{4 a, c}$ Increase in the yield of the 4,6-di-O-substituted derivatives and non formation of the 2,4-di-O-substituted derivatives (Scheme 2A. 10 and Scheme 2A.17) in the BuLi assisted reaction as compared to the NaH assisted reaction suggests that $\mathrm{Li}^{+}$stabilizes the anion $\mathbf{2 7 1}$ better than $\mathrm{Na}^{+}$does.

It is of relevance to compare the results of alkylation of the C4-trityl ether $\mathbf{2 2 3}$ (Scheme 2A.17) and the corresponding benzyl ether $\mathbf{1 9 9}$ in the presence of $\mathrm{Na}^{+}$and $\mathrm{Li}^{+}$, which could shed light on the relative ease of formation / stabilization of anions 271 and 273 under the conditions of alkylation. Since monobenzylation of the triol $\mathbf{6 5}$ with $\mathrm{NaH} / \mathrm{BnBr}$ yields $\mathbf{1 9 9}$ exclusively, it can be inferred that dibenzylation of the triol 65 proceeds exclusively via the 4-benzyl ether 199 to yield a mixture of products consisting of 2,4 and 4,6 diethers (Scheme 2A.7) in the ratio 1:5 and minor amount of the tribenzyl ether 226. ${ }^{3 c}$ On increasing the steric bulk of the alkyl group at the C4-O-
position with a trityl group, predominant methylation (using sodium hydride as the base) took place at the C2-O-position (to give 268, Scheme 2A.17) which reflects the steric crowding at the C6 position. Alkylation of the benzyl ether 199 with BuLi / BnBr on the other hand provided the 4,6-diether $\mathbf{1 0 2}$ exclusively while methylation of the trityl ether $\mathbf{2 2 3}$ with MeI / BuLi gave the C6-methylether $\mathbf{2 6 7}$ as the major product (unlike in the reaction assisted by $\mathrm{Na}^{+}$). These results give credence to our postulate that in $\mathrm{Li}^{+}$assisted reactions stabilization of the anion 271 by chelation plays a dominant role over steric effect of the trityl group. However, yet another possibility which cannot be ruled out is the inability of the larger $\mathrm{Na}^{+}$to chelate at the 4,6positions (in 223) due to the presence of bulky trityl group while the smaller $\mathrm{Li}^{+}$can still form a chelate at the 4,6 positions due to its smaller size. These results strongly suggest that stabilization of the oxyanion by chelation with lithium (272) overrides steric hindrance offered by the cis-axial substituent in deciding the regioselectivity during the nucleophilic O -substitution in myo-inositol orthoesters. Hence it appears that the major factor that controls regioselectivity during the di-O-substitution of myoinositol orthoesters in the presence of metal ions is the extent of chelation that exists between the metal ion and the two di-axial oxygen atoms. It is also interesting to note that myo-inositol derivatives were indeed found to bind lithium ions better than sodium ions as postulated above, by the picrate extraction experiments. ${ }^{18}$ Several myo-inositol derivatives tested bound lithium picrate 10-100 times better than sodium picrate. These results support our suggestion that the extent of chelation of metal ions by inositol derivatives is a major factor in deciding the observed regioselectivity for the O-substitution reactions shown in Scheme 2A.10.

We also carried out a search of the literature to see if any systematic study had been carried out to investigate the effect of counter ions on organic reactions.

Although butyllithium had been used as a base to generate lithium alkoxides from the corresponding alcohols ${ }^{19}$ their reaction with alkyl halides has not been investigated in detail. However, retardation of the rate of anionic oxy-Claisen rearrangement of $\alpha$ allyloxy ketones due to the nature of counter ion is reported. ${ }^{20}$ This study showed that rate of oxy-Claisen rearrangement was dependent on the strength of the metal oxygen bond in enolates 276; lithium enolate was the least reactive and the potassium enolate was the most reactive (Scheme 2A.20).


Scheme 2A.20. a) Metal hydride, toluene.

## 2A. 3 Conclusions

We have developed a general method for the orthogonal O-substitution of the three hydroxyl groups of myo-inositol orthoesters wherein each step in the scheme of reactions proceed with very high regioselectivity. We have also shown that chelation of the metal ions with inositol orthoester derivatives play a major role in deciding the regioselectivity in these reactions. These results coupled with other regioselective reactions reported in the literature provide convenient access to any O -substituted myo-inositol orthoester derivative.

## 2A.4. Experimental section

General. All the solvents used were purified according to literature procedures. ${ }^{21}$ All the reactions were carried out in an atmosphere of argon. Dry DMF and dry THF were used as solvents in all the experiments involving metal hydrides or n-butyllithium. Sodium hydride used in experiments was $60 \%$ suspension in mineral oil. A stock solution of n-butyllithium (1.4-1.7 M) in dry hexanes was prepared and used in all the experiments. myo-Inositol derivatives $\mathbf{6 5},{ }^{3 \mathrm{f}} \mathbf{9 8},{ }^{22} \mathbf{1 9 9},{ }^{3 \mathrm{c}} \mathbf{2 2 9}^{3 \mathrm{~b}}$ were prepared according to literature procedures. Thin layer chromatography was performed on E. Merck precoated $60 \mathrm{~F}_{254}$ plates and the spots were rendered visible either by shining UV light or by charring the plates with concd $\mathrm{H}_{2} \mathrm{SO}_{4}$ or chromic acid. Column chromatographic separations (silica gel, 100-200 mesh) and flash column chromatographic separations (silica gel, 230-400 mesh) were carried out with light petroleum-ethyl acetate mixtures as eluent. 'Usual work-up' implies washing of the organic layer with water followed by brine, drying over anhydrous sodium sulfate and removal of the solvent under reduced pressure using a rotary evaporator. Compounds previously reported in the literature were characterized by comparison of their melting points and / or ${ }^{1} \mathrm{H}$ NMR spectra with the reported data. IR spectra were recorded in $\mathrm{CHCl}_{3}$ solution or as nujol mull or as neat on a Shimadzu FTIR-8400 or Perkin Elmer 16 spectrophotometer. NMR spectra were recorded on Bruker ACF 200 spectrometer (at 200 MHz for ${ }^{1} \mathrm{H}$ and 50.3 MHz for ${ }^{13} \mathrm{C}$ ) unless otherwise mentioned. Chemical shifts ( $\delta$ ) reported are referred to internal tetramethylsilane. Microanalytical data were obtained using a Carlo-Erba CHNS-0 EA 1108 elemental analyzer. All the melting points reported are uncorrected and were recorded using a Büchi B-540 electrothermal melting point apparatus. All the asymmetrically substituted myo-inositol
derivatives reported are racemic; however, only one of the enantiomers is shown in all the schemes.

Racemic 4-O-benzyl-myo-inositol 1,3,5-orthoformate (199). The triol 65 ( 0.100 g , 0.53 mmol ) and benzyl bromide ( $0.63 \mathrm{~mL}, 5.26 \mathrm{mmol}$ ) were dissolved in DMF ( 4 $\mathrm{mL})$ and stirred with potassium carbonate $(0.727 \mathrm{~g}, 5.26 \mathrm{mmol})$ at $106^{\circ} \mathrm{C}$ for 26 h . The reaction mixture was cooled to ambient temperature, diluted with ethyl acetate and worked up as usual. The known ${ }^{3 \mathrm{c}}$ benzyl ether 199 ( $0.106 \mathrm{~g}, 72 \%$ ) was isolated as a gum after filtration of the crude product on a short column of silica gel.

Racemic 4-O-trityl-myo-inositol 1,3,5-orthoformate (223). To a solution of the triol $65(1.00 \mathrm{~g}, 5.30 \mathrm{mmol})$ in DMF $(15 \mathrm{~mL})$ was added sodium hydride $(0.233 \mathrm{~g}$, 5.83 mmol ) and stirred for 0.5 h . A solution of trityl chloride ( $1.630 \mathrm{~g}, 5.83 \mathrm{mmol}$ ) in DMF ( 8 mL ) was added drop-wise and the mixture stirred for 1 h . The reaction mixture was worked up with ethyl acetate and the crude product was purified by filtration over a short column of silica gel to obtain $223(0.900 \mathrm{~g}, 40 \%)$ as a solid.

Data for 223:
mp. $217-218^{\circ} \mathrm{C}$.
IR ( $v, \mathrm{CHCl}_{3}$ ): 3350-3700 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.60(\mathrm{~m}, 15 \mathrm{H}), 5.28(\mathrm{~s}, 1 \mathrm{H}), 4.53-4.62(\mathrm{~m}, 1 \mathrm{H}), 4.16-4.40$ $(\mathrm{m}, 3 \mathrm{H}), 3.87\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=9.8 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), 3.69-3.77(m, 1H), 3.02-3.20 ( $\mathrm{m}, 2 \mathrm{H}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 142.6,128.4,128.2,128.0,102.3,89.8,74.1,73.1,70.1,67.7$, 67.4, 60.9 .

Elemental analysis calcd for $\mathrm{C}_{26} \mathrm{H}_{24} \mathrm{O}_{6}$ : C, 72.21 ; H, 5.59. Found: C, 72.19; H, 5.73.
Racemic 4-O-(p-toluenesulfonyl)-myo-inositol 1,3,5-orthoacetate (230). To а solution of the triol $\mathbf{9 8}^{22}(0.650 \mathrm{~g}, 3.20 \mathrm{mmol})$ in dry DMF ( 6 mL ) sodium hydride
$(0.154 \mathrm{~g}, 3.84 \mathrm{mmol})$ was added and stirred for 0.5 h . Then a solution of tosyl chloride $(0.730 \mathrm{~g}, 3.84 \mathrm{mmol})$ in DMF ( 6 mL ) was added in one lot and stirred for 0.5 h. The reaction mixture was worked up with ethyl acetate as usual and the solid obtained was filtered through a short column of silica gel to get $\mathbf{2 3 0}(0.837 \mathrm{~g}, 73 \%)$ as a colorless solid.

## Data for 230:

mp. 137-139 ${ }^{\circ} \mathrm{C}$.
IR ( $v$, nujol): 3078-3533 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.83(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.8 \mathrm{~Hz}), 7.40(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.8 \mathrm{~Hz}), 5.06-5.18(\mathrm{~m}$, $1 \mathrm{H}), 4.47-4.60(\mathrm{~m}, 1 \mathrm{H}), 4.15-4.31(\mathrm{~m}, 2 \mathrm{H}), 3.96-4.13(\mathrm{~m}, 2 \mathrm{H}), 2.90-3.15$ (broad s, 1H, $\mathrm{D}_{2} \mathrm{O}$ exchangeable), 2.30-2.80 (br, $1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), $2.48(\mathrm{~s}, 3 \mathrm{H}), 1.42(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\left(\mathrm{CD}_{3}\right)_{2} \mathrm{CO}\right): \delta 145.8,133.7,130.4,128.1,108.9,75.1,74.4,73.2,69.7$, 66.8, 58.7, 23.8, 20.9.

Elemental analysis calcd for $\mathrm{C}_{15} \mathrm{H}_{18} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 50.27 ; \mathrm{H}, 5.06$. Found: C, 50.08; H, 5.06. Synthesis of 4,6-di-O-substituted-myo-inositol 1,3,5-orthoesters.

Procedure A. The required triol $\mathbf{6 5}$ or $\mathbf{9 8}(1 \mathrm{mmol})$ was taken in THF $(5 \mathrm{~mL})$ and n butyllithium ( 2.3 mmol ) was added at $0^{\circ} \mathrm{C}$. A solution of the appropriate alkyl halide ( 2.2 mmol ) in DMF ( 1 mL ) was added and the mixture stirred for $36-45 \mathrm{~h}$. The resulting mixture was then worked up with ethyl acetate. The products were separated by column chromatography using 10-20\% ethyl acetate - light petroleum as the eluent.

Procedure B. The required triol 65 or $\mathbf{9 8}(1 \mathrm{mmol})$ was taken in DMF $(2-3 \mathrm{~mL})$ and lithium hydride ( 4 mmol ) was added at ambient temperature. A solution of the appropriate alkyl halide ( 2.2 mmol ) in DMF $(1 \mathrm{~mL})$ was added and the mixture was
stirred for $36-45 \mathrm{~h}$. The reaction mixture was then worked up with ethyl acetate and the products were separated as in procedure A.

Procedure C. The required 4-O-substituted myo-inositol orthoester ( 1 mmol ) was taken in dry THF ( 4 mL ) and cooled to $0^{\circ} \mathrm{C}$. n-Butyllithium ( 1.5 mmol ) was added drop-wise using a syringe followed by a solution of the required alkyl halide, acyl halide or sulfonyl halide ( 1.2 mmol ) in DMF ( 2 mL ). The reaction mixture was stirred for 22-28 h and worked up with ethyl acetate and the products were separated as in procedure A .

Procedure D. The required 4-O-substituted myo-inositol orthoester ( 1 mmol ) was taken in DMF ( $3-4 \mathrm{~mL}$ ) and stirred for 2 h with lithium hydride ( 2 mmol ). A solution of the alkyl halide in DMF ( 2 mL ) was added and the mixture stirred for $36-40 \mathrm{~h}$. The reaction mixture was then worked up with ethyl acetate and the products were separated as in procedure A.

4,6-Di-O-benzyl-myo-inositol 1,3,5-orthoformate (102). The triol 65 ( $0.190 \mathrm{~g}, 1$ mmol ) was benzylated using benzyl bromide ( $0.376 \mathrm{~g}, 2.20 \mathrm{mmol}$ ) and n-butyllithium (Procedure A) to obtain the known $\mathbf{1 0 2}^{3 \mathrm{c}}(0.296 \mathrm{~g}, 80 \%)$ and tribenzyl ether $\mathbf{2 2 6}^{3 \mathrm{c}}$ ( $0.055 \mathrm{~g}, 12 \%$ ).
mp. (for 102) $118-120^{\circ} \mathrm{C}$, $\left(\right.$ Lit. $\left.{ }^{3 \mathrm{c}} \mathrm{mp} .124-125^{\circ} \mathrm{C}\right)$.
mp. (for 226) $103-105^{\circ} \mathrm{C}$, (Lit. ${ }^{3 \mathrm{c}} \mathrm{mp} .102-104^{\circ} \mathrm{C}$ ).
4,6-Di-O-allyl-myo-inositol 1,3,5-orthoformate (104). The triol 65 ( $0.190 \mathrm{~g}, 1$ $\mathrm{mmol})$ was allylated using allyl bromide $(0.278 \mathrm{~g}, 2.30 \mathrm{mmol})$ and lithium hydride (Procedure B) to obtain $104(0.251 \mathrm{~g}, 93 \%)$ as a gum.

## Data for 104:

IR $\left(v, \mathrm{CHCl}_{3}\right): 3250-3700 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR (500 MHz, $\left.\mathrm{CDCl}_{3}\right): \delta 5.82-5.93(\mathrm{~m}, 2 \mathrm{H}), 5.48(\mathrm{~s}, 1 \mathrm{H}), 5.30(\mathrm{dd}, 2 \mathrm{H}, \mathrm{J}=1.4$, $17.0 \mathrm{~Hz}), 5.20(\mathrm{dd}, 2 \mathrm{H}, \mathrm{J}=1.4,10.0 \mathrm{~Hz}), 4.39-4.44(\mathrm{~m}, 1 \mathrm{H}), 4.29(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=3.2 \mathrm{~Hz})$, 4.18-4.23 (m, 2H), 4.03-4.15 (m, 5H), $3.14\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=10 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 134.3,117.5,103.5,73.8,73.2,70.7,68.0,61.5$.
Elemental analysis Calcd for $\mathrm{C}_{13} \mathrm{H}_{18} \mathrm{O}_{6}$ : C, 57.77; H, 6.71. Found: C, 57.60; H, 7.04. 4,6-Di-O-(p-toluenesulfonyl)-myo-inositol 1,3,5-orthoformate (105). The racemic 4-tosylate 229 ( $0.500 \mathrm{~g}, 1.45 \mathrm{mmol}$ ) was taken in THF ( 8 mL ) and n-butyllithium $(1.60 \mathrm{mmol})$ was added at $-78^{\circ} \mathrm{C}$. A solution of tosyl chloride $(0.332 \mathrm{~g}, 1.75 \mathrm{mmol})$ in THF ( 2 mL ) was added and the mixture stirred for 2 h during which the reaction temperature was allowed to come to ambient temperature. The resulting mixture was then worked up with ethyl acetate. The known ditosylate 105 ( $0.412 \mathrm{~g}, 57 \%$ ) and tritosylate $\mathbf{2 4 0}$ ( $0.284 \mathrm{~g}, \mathbf{3 0 \%}$ ) were separated by column chromatography.
mp. (for 105) $181-182^{\circ} \mathrm{C}$, (Lit. ${ }^{15} \mathrm{mp} .183-185^{\circ} \mathrm{C}$ ).
mp. (for 240) $102-104^{\circ} \mathrm{C}$, (Lit. $\left.{ }^{15} \mathrm{mp} .103-105^{\circ} \mathrm{C}\right)$.
4,6-Di-O-benzyl-myo-inositol $\mathbf{1 , 3 , 5 - o r t h o a c e t a t e}$ (227). The triol 98 ( $0.204 \mathrm{~g}, 1$ mmol ) was benzylated using benzyl bromide ( $0.359 \mathrm{~g}, 2.20 \mathrm{mmol}$ ) and n-butyllithium (Procedure A) to obtain $227(0.268 \mathrm{~g}, 70 \%)$ as a solid and the monobenzyl ether $\mathbf{2 2 8}$ $(0.040 \mathrm{~g}, 14 \%)$ as a gum.

Data for 227:
mp. $88-90^{\circ} \mathrm{C}$.
IR ( $v, \mathrm{CHCl}_{3}$ ): 3300-3600 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.28(\mathrm{~s}, 10 \mathrm{H}), 4.62(\mathrm{AB} \mathrm{q}, 4 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}), 4.08-4.46(\mathrm{~m}, 6 \mathrm{H})$, $3.04\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), $1.44(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 137.7,128.5,127.9,127.7,109.3,73.8,73.6,71.7,67.9,60.4$, 24.4.

Elemental analysis calcd for $\mathrm{C}_{22} \mathrm{H}_{24} \mathrm{O}_{6}$ : C, 68.74; H, 6.29. Found: C, 68.97; H, 6.14.

## Data for 228:

${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.15-7.53(\mathrm{~m}, 5 \mathrm{H}), 4.66(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=2 \mathrm{~Hz}), 4.34-4.50(\mathrm{~m}, 2 \mathrm{H})$, 4.15-4.31 (m, 3H), 3.96-4.10 (m, 1H), 3.67 (d, 1H, J = 9.7 Hz, $\mathrm{D}_{2} \mathrm{O}$ exchangeable), 3.00-3.35 (broad s, $1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), $1.43(\mathrm{~s}, 3 \mathrm{H})$.

4,6-Di-O-benzyl-myo-inositol 1,3,5-orthoformate (102). The monobenzyl ether 199 $(0.283 \mathrm{~g}, 1 \mathrm{mmol})$ was benzylated by procedure C to obtain the known ${ }^{3 \mathrm{c}}$ dibenzyl ether 102 ( $0.328 \mathrm{~g}, 89 \%$ ) and the tribenzyl ether $226(0.037 \mathrm{~g}, 8 \%)$.
mp. (for 102) $118-120^{\circ} \mathrm{C}$.
Racemic 4-O-benzyl-6-O-allyl-myo-inositol 1,3,5-orthoformate (231). The monobenzyl ether $199(0.283 \mathrm{~g}, 1 \mathrm{mmol})$ was allylated by procedure C to obtain the 6-allyl ether 231 ( $0.256 \mathrm{~g}, 80 \%$ ) and the diallyl ether 236 ( $0.036 \mathrm{~g}, 10 \%$ ) as gums.

## Data for 231:

IR ( $v, \mathrm{CHCl}_{3}$ ): $3220-3600 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.33(\mathrm{~s}, 5 \mathrm{H}), 5.75-6.00(\mathrm{~m}, 1 \mathrm{H}), 5.47(\mathrm{~s}, 1 \mathrm{H}), 5.16-5.35(\mathrm{~m}, 2 \mathrm{H})$, $4.62(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.8 \mathrm{~Hz}), 4.38-4.49(\mathrm{~m}, 1 \mathrm{H}), 4.26-4.37(\mathrm{~m}, 2 \mathrm{H}), 3.95-4.25(\mathrm{~m}$, $5 \mathrm{H}), 3.22\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=11.2 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 137.9,134.3,128.7,128.1,127.8,117.8,103.6,73.9,73.3$, 71.7, 71.0, 68.0, 61.7 .

Elemental analysis Calcd for $\mathrm{C}_{17} \mathrm{H}_{20} \mathrm{O}_{6}$ : C, 63.74; H, 6.29. Found: C, 63.83; H, 6.47.

## Data for 236:

${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.30(\mathrm{~s}, 5 \mathrm{H}), 5.75-6.10(\mathrm{~m}, 2 \mathrm{H}), 5.50(\mathrm{~s}, 1 \mathrm{H}), 5.10-5.40(\mathrm{~m}, 4 \mathrm{H})$, $4.58(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}), 4.20-4.45(\mathrm{~m}, 5 \mathrm{H}), 3.90-4.20(\mathrm{~m}, 5 \mathrm{H})$.

The monoallyl ether 231 obtained above was converted to the $\mathrm{known}^{3 \mathrm{c}}$ racemic dibenzyl ether $\mathbf{2 4 2}$ as follows: To a solution of $\mathbf{2 3 1}(0.050 \mathrm{~g}, 0.20 \mathrm{mmol})$ in DMF ( 1
$\mathrm{mL})$, sodium hydride $(0.016 \mathrm{~g}, 0.40 \mathrm{mmol})$ was added and stirred for 5 min . Benzyl bromide $(0.068 \mathrm{~g}, 0.40 \mathrm{mmol})$ was added and the mixture stirred for 0.5 h and worked up as usual to obtain 242 ( $0.062 \mathrm{~g}, 97 \%$ ).

Racemic 4-O-benzyl-6-O-acetyl-myo-inositol 1,3,5-orthoformate (232). The monobenzyl ether 199 ( $0.286 \mathrm{~g}, 1 \mathrm{mmol}$ ) was acetylated (Procedure D, reaction time $4 \mathrm{~h})$, with acetic anhydride $(0.125 \mathrm{~g}, 1.20 \mathrm{mmol})$ to obtain the racemic $232(0.230 \mathrm{~g}$, $70 \%$ ) and the diacetate $237(0.057 \mathrm{~g}, 15 \%)$ as white solids.

## Data for 232:

mp. $102-104{ }^{\circ} \mathrm{C}$.
IR (v, $\left.\mathrm{CHCl}_{3}\right): 3300-3570,1737 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.16-7.45(\mathrm{~m}, 5 \mathrm{H}), 5.51(\mathrm{~s}, 1 \mathrm{H}), 5.35-5.46(\mathrm{~m}, 1 \mathrm{H}), 4.62-4.72$ $(\mathrm{m}, 1 \mathrm{H}), 4.57(\mathrm{~s}, 2 \mathrm{H}), 4.05-4.43(\mathrm{~m}, 4 \mathrm{H}), 3.31\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=11.8 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), $1.86(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 170.2,137.5,128.8,128.4,127.7,103.5,73.6,72.8,72.2,68.6$, 66.6, 61.6, 20.9.

Elemental analysis Calcd for $\mathrm{C}_{16} \mathrm{H}_{18} \mathrm{O}_{7}$ : C, $59.62 ; \mathrm{H}, 5.63$. Found: C, 59.91; H, 5.65. Data for 237:
mp. $116-118^{\circ} \mathrm{C}$
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.30-7.45(\mathrm{~m}, 5 \mathrm{H}), 5.58(\mathrm{~s}, 1 \mathrm{H}), 5.40(\mathrm{td}, 1 \mathrm{H}, \mathrm{J}=1.5,3.9 \mathrm{~Hz})$, 5.31-5.36 (m, 1H), 4.65-4.72 (m, 1H), 4.59 (AB q, 2H, J = 11.3 Hz), 4.40-4.46 (m, $1 \mathrm{H}), 4.29-4.39(\mathrm{~m}, 2 \mathrm{H}), 2.23(\mathrm{~s}, 3 \mathrm{H}), 1.88(\mathrm{~s}, 3 \mathrm{H})$.

Racemic 4-O-benzyl-6-O-(p-toluenesulfonyl)-myo-inositol $\mathbf{1 , 3 , 5}$-orthoformate (233). The monobenzyl ether 199 ( $0.278 \mathrm{~g}, 1 \mathrm{mmol}$ ) was sulfonylated (Procedure D, reaction time 4 h ), with $p$-toluenesulfonyl chloride $(0.228 \mathrm{~g}, 1.20 \mathrm{mmol})$ to obtain the 6-tosylate $\mathbf{2 3 3}$ as a solid ( $0.283 \mathrm{~g}, 65 \%$ ) and the ditosylate $\mathbf{2 3 8}(0.060 \mathrm{~g}, 10 \%)$ as gum.

## Data for 233:

mp. $160-162^{\circ} \mathrm{C}$.
IR ( $v, \mathrm{CHCl}_{3}$ ): 3300-3600 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.75(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 7.20-7.40(\mathrm{~m}, 7 \mathrm{H}), 5.43(\mathrm{~s}, 1 \mathrm{H}), 5.13-$ $5.25(\mathrm{~m}, 1 \mathrm{H}), 4.59(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}), 4.40-4.47(\mathrm{~m}, 1 \mathrm{H}), 4.27-4.39(\mathrm{~m}, 1 \mathrm{H})$, 4.15-4.24 (m, 1H), 3.96-4.14 (m, 2H), $3.11\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), 2.44 (s, 3H).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 145.7,137.4,133.1,130.3,128.7,128.1,127.9,103.3,73.0$, 72.9, 72.8, 72.3, 71.8, 67.5, 60.9, 21.9.

Elemental analysis Calcd for $\mathrm{C}_{21} \mathrm{H}_{22} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 58.06$; H, 5.10. Found: C, 58.10 ; H, 4.80 .

## Data for 238:

${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.79(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 7.73(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 5.44(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}$ $=1.4 \mathrm{~Hz}), 5.09(\mathrm{td}, 1 \mathrm{H}, \mathrm{J}=1.5,3.9 \mathrm{~Hz}), 4.90-4.96(\mathrm{~m}, 1 \mathrm{H}), 4.53(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.8$ $\mathrm{Hz}), 4.35-4.41(\mathrm{~m}, 1 \mathrm{H}), 4.25(\mathrm{td}, 1 \mathrm{H}, \mathrm{J}=1.5,3.4 \mathrm{~Hz}), 4.13-4.20(\mathrm{~m}, 1 \mathrm{H}), 4.04-4.11$ (m, 1H), $2.46(\mathrm{~s}, 3 \mathrm{H}), 2.43(\mathrm{~s}, 3 \mathrm{H})$.

Racemic 4-O-(p-toluenesulfonyl)-6-O-allyl-myo-inositol 1,3,5-orthoformate (234). The racemic 4-tosylate $\mathbf{2 2 9}^{\mathbf{3 b}}(0.344 \mathrm{~g}, 1 \mathrm{mmol})$ was allylated (Procedure D), with allyl bromide ( $0.145 \mathrm{~g}, 1.20 \mathrm{mmol}$ ) to obtain $234(0.278 \mathrm{~g}, 72 \%)$ and the diallyl ether $239(0.026 \mathrm{~g}, 6 \%)$ as gums.

## Data for 234:

IR ( $v, \mathrm{CHCl}_{3}$ ): $3200-3600 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.81(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 7.37(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 5.75-6.00(\mathrm{~m}$, $1 \mathrm{H}), 5.44(\mathrm{~s}, 1 \mathrm{H}), 5.05-5.38(\mathrm{~m}, 3 \mathrm{H}), 4.35-4.50(\mathrm{~m}, 1 \mathrm{H}), 3.95-4.34(\mathrm{~m}, 6 \mathrm{H}), 3.18(\mathrm{~d}$, $1 \mathrm{H}, \mathrm{J}=10.3 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), $2.47(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): ~ \delta 145.8,133.9,130.3,128.2,118.1,103.3,72.8,72.3,70.8,67.5$, 60.9, 21.9.

Data for 239:
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.82(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.6 \mathrm{~Hz}), 7.38(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.2 \mathrm{~Hz}), 5.75-6.10(\mathrm{~m}$, $2 \mathrm{H}), 5.49(\mathrm{~s}, 1 \mathrm{H}), 5.05-5.40(\mathrm{~m}, 5 \mathrm{H}), 3.80-4.45(\mathrm{~m}, 9 \mathrm{H}), 2.47(\mathrm{~s}, 3 \mathrm{H})$.

The monoallyl ether $\mathbf{2 3 4}$ obtained above was converted to the known ${ }^{3 c} 4$-allyl ether 255 as follows: The racemic $234(0.050 \mathrm{~g}, 0.13 \mathrm{mmol})$ and sodium methoxide ( 0.070 $\mathrm{g}, 1.30 \mathrm{mmol})$ were refluxed in dry methanol $(2 \mathrm{~mL})$ for 2 h . The reaction mixture was cooled to ambient temperature and worked up as usual to obtain racemic $\mathbf{2 5 5}$ $(0.026 \mathrm{~g}, 87 \%)$ as a gum. The monoallyl ether $\mathbf{2 5 5}(0.026 \mathrm{~g})$ was characterized as its diacetate $256(0.031 \mathrm{~g}, 89 \%)$ obtained by acetylation with acetic anhydride $(0.046 \mathrm{~g}$, 0.45 mmol ) in pyridine ( 1 mL ) for 12 h .

Data for 256:
mp. $87-89^{\circ} \mathrm{C}$ (colorless solid).
IR ( $v, \mathrm{CHCl}_{3}$ ): $1742 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right): \delta 5.79-5.94(\mathrm{~m}, 1 \mathrm{H}), 5.57(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=1.0 \mathrm{~Hz}), 5.38-5.43$ $(\mathrm{m}, 1 \mathrm{H}), 5.31-5.35(\mathrm{~m}, 1 \mathrm{H}), 5.19-5.29(\mathrm{~m}, 2 \mathrm{H}), 4.61-4.66(\mathrm{~m}, 1 \mathrm{H}), 4.35-4.40(\mathrm{~m}, 1 \mathrm{H})$, 4.27-4.33 (m, 2H), 4.03-4.10 (m, 2H), $2.22(\mathrm{~s}, 3 \mathrm{H}), 2.06(\mathrm{~s}, 3 \mathrm{H})$.

Elemental analysis Calcd for $\mathrm{C}_{14} \mathrm{H}_{18} \mathrm{O}_{8}$ : C, 53.50; H, 5.77. Found: C, 53.70; H, 5.73.
Racemic 4-O-(p-toluenesulfonyl)-6-O-allyl-myo-inositol 1,3,5-orthoacetate (235). The racemic 4-tosylate 230 ( $0.358 \mathrm{~g}, 1 \mathrm{mmol}$ ) was allylated (Procedure D), with allyl bromide ( $0.146 \mathrm{~g}, 1.20 \mathrm{mmol}$ ) to obtain $235(0.246 \mathrm{~g}, 62 \%)$ and diallyl ether 241 $(0.039 \mathrm{~g}, 9 \%)$ as gums.

Data for 235:
IR ( $v, \mathrm{CHCl}_{3}$ ): 3300-3590 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.82(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}) .7 .38(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 5.75-6.00(\mathrm{~m}$, $1 \mathrm{H}), 5.15-5.40(\mathrm{~m}, 2 \mathrm{H}), 5.00-5.12(\mathrm{~m}, 1 \mathrm{H}), 4.34-4.39(\mathrm{~m}, 1 \mathrm{H}), 3.94-4.28(\mathrm{~m}, 6 \mathrm{H})$, 2.89-3.10 (broad s, 1H, $\mathrm{D}_{2} \mathrm{O}$ exchangeable), $2.47(\mathrm{~s}, 3 \mathrm{H}), 1.42(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 145.6,134.0,133.4,130.3,128.2,117.9,109.4,73.5,73.2$, 72.9, 72.7, 70.8, 67.5, 60.0, 24.3, 21.8.

Elemental analysis calcd for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 54.26 ; \mathrm{H}, 5.57$. Found: C, 54.12; H, 5.55. Data for 241:
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.82(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 7.38(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.8 \mathrm{~Hz}), 5.75-6.10(\mathrm{~m}$, $2 H), ~ 5.15-5.40(\mathrm{~m}, 4 \mathrm{H}), 4.95-5.10(\mathrm{~m}, 1 \mathrm{H}), 3.95-4.35(\mathrm{~m}, 8 \mathrm{H}), 3.75-3.85(\mathrm{~m}, 1 \mathrm{H})$, $2.47(\mathrm{~s}, 3 \mathrm{H}), 1.43(\mathrm{~s}, 3 \mathrm{H})$.

## Reaction of racemic 4-O-benzoyl-myo-inositol 1,3,5-orthoformate with lithium hydride and benzyl bromide.

Procedure E. To a solution of the diol $\mathbf{2 6 0}(0.294 \mathrm{~g}, 1 \mathrm{mmol})$ and benzyl bromide ( $0.13 \mathrm{~mL}, 1.10 \mathrm{mmol}$ ) in DMF ( 3 mL ) lithium hydride ( $0.016 \mathrm{~g}, 2.00 \mathrm{mmol}$ ) was added and the mixture stirred for 24 h at ambient temperature. The reaction mixture was worked up with ethyl acetate as the solvent and the products were separated by column chromatography $263(0.073 \mathrm{~g}, 19 \%), 262(0.151 \mathrm{~g}, 39 \%)$ and $199(0.062 \mathrm{~g}$, $22 \%$ ).

Procedure F. To a solution of the $\operatorname{diol} 260(0.250 \mathrm{~g}, 0.85 \mathrm{mmol})$ and benzyl bromide ( $0.11 \mathrm{~mL}, 0.94 \mathrm{mmol}$ ) in DMF ( 3 mL ) lithium hydride ( $0.014 \mathrm{~g}, 1.70 \mathrm{mmol}$ ) was added and the mixture stirred for 24 h at ambient temperature. The reaction mixture was quenched with ice and DMF was evaporated at $75-80^{\circ} \mathrm{C}$ under reduced pressure and worked up as usual with ethyl acetate. The products were separated by column chromatography to obtain 263 ( $0.039 \mathrm{~g}, 12 \%$ ) and 199 ( $0.174 \mathrm{~g}, 73 \%$ ).

## Data for 262:

mp. $104-106^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.88(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.1 \mathrm{~Hz}), 7.45-7.56(\mathrm{~m}, 1 \mathrm{H}), 7.10-7.31(\mathrm{~m}, 7 \mathrm{H})$, 5.71-5.79 (m, 1H), $5.57(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=1.1 \mathrm{~Hz}), 4.62-4.71(\mathrm{~m}, 1 \mathrm{H}), 4.54(\mathrm{~s}, 2 \mathrm{H}), 4.31-4.46$ $(\mathrm{m}, 3 \mathrm{H}), 4.20-4.28(\mathrm{~m}, 1 \mathrm{H}), 3.10-3.50$ (broad $\mathrm{s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable).

## Data for 263:

mp. $98-100^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.17(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=6.9 \mathrm{~Hz}), 7.55-7.67(\mathrm{~m}, 1 \mathrm{H}), 7.43-7.54(\mathrm{~m}, 2 \mathrm{H})$, $7.38(\mathrm{~s}, 5 \mathrm{H}), 5.50-5.58(\mathrm{~m}, 2 \mathrm{H}), 4.75(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.4 \mathrm{~Hz}), 4.56-4.63(\mathrm{~m}, 1 \mathrm{H})$, 4.41-4.55 (m, 3H), 4.31-4.39(m, 1H), $3.86\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=9.9 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable $)$.

Racemic 2-O-methyl-4-O-benzyl-6-O-allyl-myo-inositol 1,3,5-orthoformate (264). Racemic 199 ( 0.281 g , 1 mmol ) was taken in THF ( 4 mL ) and cooled to $0^{\circ} \mathrm{C}$. n Butyllithium ( $1.1 \mathrm{~mL}, 1.50 \mathrm{mmol}$ ) was added slowly followed by allyl bromide ( 0.145 $\mathrm{g}, 1.20 \mathrm{mmol})$ in dry DMF ( 2 mL ) and the mixture stirred for 22 h . Then excess sodium hydride ( $0.100 \mathrm{~g}, 2.50 \mathrm{mmol}$ ) and methyl iodide ( $0.355 \mathrm{~g}, 2.50 \mathrm{mmol}$ ) were added and the mixture stirred for 4 h and worked up with ethyl acetate. The triether 264 ( $0.267 \mathrm{~g}, 80 \%$ ) was separated (as gum) by flash column chromatography.

## Data for 264:

${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.50(\mathrm{~m}, 5 \mathrm{H}), 5.75-6.05(\mathrm{~m}, 1 \mathrm{H}), 5.51(\mathrm{~s}, 1 \mathrm{H}), 5.14-5.40$ $(\mathrm{m}, 2 \mathrm{H}), 4.63(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.7 \mathrm{~Hz}), 4.24-4.50(\mathrm{~m}, 5 \mathrm{H}), 4.00-4.23(\mathrm{~m}, 2 \mathrm{H}), 3.78(\mathrm{~s}$, $1 \mathrm{H}), 3.48(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 137.9,134.4,128.7,128.1,127.8,117.9,103.4,74.2,74.0$, $71.9,71.1,70.2,70.0,69.6,68.3,56.9$.

Elemental analysis Calcd for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{O}_{6}$ : C, 64.66; H, 6.63. Found: C, 64.76; H, 6.89.

Racemic 4-O-(t-butyldimethylsilyl)-myo-inositol 1,3,5-orthoformate (265). The triol $65(0.500 \mathrm{~g}, 2.63 \mathrm{mmol})$ was silylated (Procedure A) in THF ( 25 mL ) using n butyllithium ( $1.7 \mathrm{~mL}, 2.63 \mathrm{mmol}$ ) and a solution of $t$-butyldimethylsilyl chloride ( $0.476 \mathrm{~g}, 3.15 \mathrm{mmol}$ ) in THF ( 5 mL ). The known racemic 265 ( $0.584 \mathrm{~g}, 73 \%$ ) was isolated by column chromatography.
mp. $74-76^{\circ} \mathrm{C}$, (Lit. ${ }^{17} \mathrm{mp} .73-74^{\circ} \mathrm{C}$ ).
Racemic 2,4-di-O-(t-butyldimethylsilyl)-myo-inositol 1,3,5-orthoformate (266). The triol $65(0.150 \mathrm{~g}, 0.80 \mathrm{mmol})$ was silylated (Procedure B) in dry DMF ( 2 mL ) using lithium hydride $(0.260 \mathrm{~g}, 3.20 \mathrm{mmol})$ and $t$-butyldimethylsilyl chloride $(0.253 \mathrm{~g}$, $1.68 \mathrm{mmol})$ to obtain the known racemic $266(0.174 \mathrm{~g}, 52 \%)$ as a solid.
mp. $79-81^{\circ} \mathrm{C}$, $\left(\right.$ Lit. $\left.{ }^{3 \mathrm{n}} \mathrm{mp} .75-78{ }^{\circ} \mathrm{C}\right)$.
Methylation of racemic 4-O-trityl-myo-inositol 1,3,5-orthoformate (223). Racemic 223 ( $0.600 \mathrm{~g}, 1.40 \mathrm{mmol}$ ) was methylated (Procedure C) in THF ( 6 mL ) using n-butyllithium ( $1 \mathrm{~mL}, 1.68 \mathrm{mmol}$ ) and methyl iodide $(0.12 \mathrm{~mL}, 1.68 \mathrm{mmol})$ in DMF ( 1 mL ) to obtain the 6-methyl ether $267(0.147 \mathrm{~g}, 24 \%)$, the 2-methyl ether 268 $(0.036 \mathrm{~g}, 6 \%)$ and the dimethyl ether $269(0.183 \mathrm{~g}, 29 \%)$.

Data for 267:
mp. $165-166^{\circ} \mathrm{C}$.
IR ( $v$, nujol): 3323-3541 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.43-7.55(\mathrm{~m}, 6 \mathrm{H}), 7.23-7.40(\mathrm{~m}, 9 \mathrm{H}), 5.29(\mathrm{~s}, 1 \mathrm{H}), 4.43-4.52$ $(\mathrm{m}, 1 \mathrm{H}), 4.14-4.24(\mathrm{~m}, 2 \mathrm{H}), 4.02-4.10(\mathrm{~m}, 1 \mathrm{H}), 3.72-3.83(\mathrm{~m}, 1 \mathrm{H}), 3.46(\mathrm{~s}, 3 \mathrm{H}), 3.08-$ $3.18(\mathrm{~m}, 1 \mathrm{H}), 2.77-2.94$ (broad $\mathrm{s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 143.8,128.6,128.0,127.5,102.8,88.0,75.8,73.7,72.4,69.0$, 68.0, 61.3, 57.6.

Elemental analysis calcd for $\mathrm{C}_{27} \mathrm{H}_{26} \mathrm{O}_{6}$ : C, 72.63; H, 5.87. Found: C, 72.30; H, 6.10.

## Data for 268:

mp. $191-193{ }^{\circ} \mathrm{C}$.
IR ( $v$, nujol): 3304-3541 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.25-7.50(\mathrm{~m}, 15 \mathrm{H}), 5.32(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=1.1 \mathrm{~Hz}), 4.54-4.62(\mathrm{~m}, 1 \mathrm{H})$, 4.31-4.45 (m, 2H), 3.79-3.89 (m, $2 \mathrm{H}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), 3.57-3.65 (m, 1 H$)$, $3.48(\mathrm{~s}, 3 \mathrm{H}), 3.41-3.51(\mathrm{~m}, 1 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 142.7,128.5,128.2,128.1,102.3,89.9,71.1,70.3,69.9,69.0$, 68.1, 67.8, 56.9.

Elemental analysis calcd for $\mathrm{C}_{27} \mathrm{H}_{26} \mathrm{O}_{6}$ : C, 72.63 ; H, 5.87. Found: C, 72.43; H, 5.44.
Data for 269:
mp. $251-253{ }^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.45-7.55(\mathrm{~m}, 5 \mathrm{H}), 7.20-7.40(\mathrm{~m}, 10 \mathrm{H}), 5.34(\mathrm{~s}, 1 \mathrm{H}), 4.44-4.52$ $(\mathrm{m}, 1 \mathrm{H}), 4.28-4.36(\mathrm{~m}, 1 \mathrm{H}), 4.08-4.15(\mathrm{~m}, 1 \mathrm{H}), 3.99-4.07(\mathrm{~m}, 1 \mathrm{H}), 3.74-3.80(\mathrm{~m}, 1 \mathrm{H})$, $3.53(\mathrm{~s}, 3 \mathrm{H}), 3.31(\mathrm{~s}, 3 \mathrm{H}), 3.07-3.14(\mathrm{~m}, 1 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 143.9,128.6,128.1,127.5,102.8,88.0,75.9,70.1,69.7,69.4$, 69.2, 68.5, 57.7, 56.7 .

Elemental analysis calcd for $\mathrm{C}_{28} \mathrm{H}_{28} \mathrm{O}_{6}$ : C, 73.03; H, 6.13. Found: C, 72.99; H, 6.10. The proportion of 267 and 268 formed on using butyllithium and sodium hydride for the methylation of $\mathbf{2 2 3}$ were estimated as follows: Racemic 223 ( $0.100 \mathrm{~g}, 0.23 \mathrm{mmol})$ was methylated (Procedure C) in THF ( 1 mL ) using n-butyllithium ( $0.15 \mathrm{~mL}, 0.25$ $\mathrm{mmol})$ and methyl iodide $(0.02 \mathrm{~mL}, 0.25 \mathrm{mmol})$ in DMF $(0.18 \mathrm{~mL})$. The mixture of monoethers 267 and $268(0.042 \mathrm{~g})$ was separated by column chromatography and their proportion $(\mathbf{2 6 7 : 2 6 8}=61: 39)$ estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy. Use of sodium hydride $(0.010 \mathrm{~g}, 0.25 \mathrm{mmol})$ instead of butyllithium in the above experiment
provided a mixture of the two ethers $(0.053 \mathrm{~g})$ in the ratio 267:268 $=11: 89$ and the dimethyl ether 269 ( 0.030 g. 28\%).

Racemic 2-O-methyl-4-O-trityl-myo-inositol 1,3,5-orthoformate (268). To a solution of the ditosylate $105(0.300 \mathrm{~g}, 0.60 \mathrm{mmol})$ in DMF, sodium hydride $(0.036 \mathrm{~g}$, $0.90 \mathrm{mmol})$ was added followed by methyl iodide $(0.06 \mathrm{~mL}, 0.90 \mathrm{mmol})$. The reaction mixture was stirred for 15 min and worked up with ethyl acetate as usual to get 2-O-methyl-4,6-di-O-(p-toluenesulfonyl)-myo-inositol 1,3,5-orthoformate (253, $0.287 \mathrm{~g}, 93 \%)$.

Data for 253:
mp. $141-143{ }^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.84(\mathrm{~d}, 4 \mathrm{H}, \mathrm{J}=8.2 \mathrm{~Hz}), 7.39(\mathrm{~d}, 4 \mathrm{H}, \mathrm{J}=8.2 \mathrm{~Hz}), 5.44(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}$ $=1.6 \mathrm{~Hz}), 5.12(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=4 \mathrm{~Hz}), 4.26-4.37(\mathrm{~m}, 2 \mathrm{H}), 4.13-4.22(\mathrm{~m}, 1 \mathrm{H}), 3.59-3.68(\mathrm{~m}$, 1H), 3.42 ( $\mathrm{s}, 3 \mathrm{H}$ ), 2.48 ( $\mathrm{s}, 6 \mathrm{H})$.

The methyl ether $\mathbf{2 5 3}(0.450 \mathrm{~g}, 0.90 \mathrm{mmol})$ and sodium methoxide $(0.486 \mathrm{~g}, 9 \mathrm{mmol})$ were refluxed in dry methanol : THF (4:1) for 6 h . The solvents were evaporated and the solid obtained was chromatographed over silica gel using 1:1 ethyl acetate - light petroleum as eluent to obtain 2-O-methyl-myo-inositol 1,3,5-orthoformate (270, 0.124 g, 69\%).

Data for 270:
mp. $179-181^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\left(\mathrm{CD}_{3}\right)_{2} \mathrm{CO}\right): \delta 5.38(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=1.2 \mathrm{~Hz}), 4.98-5.10$ (broad s, $2 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), 4.39-4.52 (m, 2H), 4.24-4.31 (m, 2H), 4.15-4.23 (m, 1H), 3.75-3.82 (m, 1H), 3.46 (s, 3H).

To a solution of the diol $\mathbf{2 7 0}$ obtained above $(0.080 \mathrm{~g}, 0.40 \mathrm{mmol})$ in DMF was added sodium hydride $(0.024 \mathrm{~g}, 0.60 \mathrm{mmol})$ and trityl chloride $(0.167 \mathrm{~g}, 0.60 \mathrm{mmol})$ and the
reaction mixture stirred at ambient temperature for 1 h and worked up as usual with ethyl acetate. Column chromatography over silica gel gave the trityl ether 268 (0.054 g, 31\%).
mp. $191-193{ }^{\circ} \mathrm{C}$.

## 2A.5. References

1. Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Das, T. Chem. Rev. 2003, 103, 4477.
2. For example see Rossier, M. F.; Capponi, A. M.; Vallotton, M. B. Molecular and Cellular Endocrinology 1988, 57, 163.
3. (a) Flores-Mosquera, M.; Martín-Lomas, M.; Chiara, J. L. Tetrahedron Lett. 1998, 39, 5085; (b) Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2001, 42, 3037; (c) Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I. M.; Vacca, J. P.; deSolms, S. J.; Huff, J. R. J. Chem. Soc., Perkin Trans. 1 1989, 1423; (d) Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2000, 41, 4185; (e) Li, C.; Vasella, A. Helv. Chim. Acta. 1993, 76, 211; (f) Lee, H. W.; Kishi, Y. J. Org. Chem. 1985, 50, 4402; (g) Biamonte, M. A.; Vasella, A. Helv. Chim. Acta. 1998, 81, 695; (h) Riley, A. M.; Murphy, C. T.; Lindley, C. J.; Westwick, J.; Potter, B. V. L. Bioorg. Med. Chem. Lett. 1996, 6, 2197; (i) Ballereau, S.; Poirier, S. N.; Guillemette, G.; Spiess, B.; Schlewer, G. J. Chem. Soc., Perkin Trans. 1 1998, 1859; (j) Ozaki, S.; Koga, Y.; Ling, L.; Watanabe, Y.; Kimura, Y.; Hirata, M. Bull. Chem. Soc. Jpn. 1994, 67, 1058; (k) Billington, D. C.; Baker, R. J. Chem. Soc., Chem. Commun. 1987, 1011; (1) Jane deSolms, S.; Vacca, J. P.; Huff, J. R. Tetrahedron Lett. 1987, 28, 4503; (m) Banerjee, T.; Shashidhar, M. S. Tetrahedron Lett. 1994, 35, 8053; (n) Craig, B. N.; Janssen, M. U.; Wickersham, B. M.; Rabb, D. M.; Chang, P. S.; O'Leary, D. J. J. Org. Chem. 1996, 61, 9610.
4. (a) Uhlmann, P.; Vasella, A. Helv. Chim. Acta. 1992, 75, 1979; (b) Bhosekar, G.; Murali, C.; Gonnade, R. G.; Shashidhar, M. S.; Bhadbhade, M. M. Cryst. Growth. Des. 2005, 5, 1977; (c) Bernet, B; Vasella, A. Helv. Chim. Acta. 2000, 83, 995; (d) Fierman, M.; Nelson, A.; Khan, S. I.; Barfield, M.; O’Leary. D. J. Org. Lett. 2000, 2, 2077.
5. Chung, S-K.; Chang, Y-T.; Lee, J. W.; Ji, Y-K. Korean J. Med. Chem. 1997, 7, 82.
6. Andersch, P.; Schneider, M. P. Tetrahedron: Asymmetry 1993, 4, 2135.
7. Also see Praveen, T.; Das, T.; Sureshan, K. M.; Shashidhar, M. S.; Samanta, U.; Pal, D.; Chakrabarti, P. J. Chem. Soc., Perkin Trans. 2 2002, 358.
8. Organic Chemistry, Clayden, J.; Greeves, N.; Warren, S.; Wothers, P.; Oxford University press, New York, N.Y. 2001, pp. 199-202.
9. Baudin, G.; Glänzer, B. I.; Swaminathan, K. S.; Vasella, A. Helv. Chim. Acta. 1988, 71, 1367.
10. Also see (a) Zapata, A.; Bernet, B.; Vasella, A. Helv. Chim. Acta. 1996, 79, 1169; (b) Das, T.; Praveen, T.; Shashidhar, M. S. Carbohydr. Res. 1998, 313, 55.
11. (a) Sureshan. K. M. Ph.D. thesis, University of Pune, January, 2002; (b) XIV Carbohydrate Conference, Indian Institute of Technology, Madras, Chennai, December 16-17, 1999.
12. (a) Chemistry of the Metal Chelate Compounds, Martell, A. E.; Calvin, M. Prentice Hall, New York, N.Y. Chapter-5, pp.181-206. (b) Comprehensive Coordination Chemistry, Vol. 3. Wilkinson, G. (Ed.), Pergamon press, Chapter-23, pp.1-80.
13. Streitwieser, A.; Husemann, M.; Kim, Y.-J. J. Org. Chem. 2003, 68, 7937.
14. Falck, J. R.; Barma, D. K.; Venkataraman, S. K.; Baati, R.; Mioskowski, C. Tetrahedron Lett. 2002, 43, 963.
15. Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Gonnade, R. G.; Bhadbhade, M. M. Carbohydr. Res. 2002, 337, 2399.
16. Trost, B. M.; Godleski, S. A.; Belletire, J. L. J. Org. Chem. 1979, 44, 2052.
17. Chung, M-K.; Orlova, G.; Goddard, J. D.; Schlaf, M.; Harris, R.; Beveridge, T. J.; White, G.; Hallet, F. R. J. Am. Chem. Soc. 2002, 124, 10508.
18. Devaraj, S.; Shashidhar, M. S.; Dixit, S. S. Tetrahedron 2005, 61, 529.
19. Mukaiyama, T.; Shintou, T.; Fukumoto, K. J. Am. Chem. Soc. 2003, 125, 10538.
20. Koreeda, M.; Luengo, J. I. J. Am. Chem. Soc. 1985, 107, 5572.
21. Purification of Laboratory Chemicals, Perrin, D. D.; Armarego, W. L. F. $2^{\text {nd }}$ edition, Pergamon Press, Oxford, UK, 1988.
22. Garrett, S. W.; Liu, C.; Riley, A. M.; Potter, B. V. L. J. Chem. Soc., Perkin Trans. 1 1998, 1367.


















































${ }^{1}$ H NMR spectra for the mixture of products 267 and 268 obtained on methylation of the racemic diol 223 with
a) butyllithium and methyl iodide.

b) sodium hydride and methyl iodide.




## 2B.1. Introduction

In the previous section a novel methodology for the orthogonal protection of the three hydroxyl groups of myo-inositol 1,3,5-orthoesters was described. Although it is likely that good regioselectivity observed was due to the inherent rigidity of the trioxaadamantane frame work of these orthoesters, it was curious to see if the regioselectivity of O-substitution reactions would get affected in partially protected myo-inositol derivatives in the absence of the orthoester moiety, on using sodium and lithium alkoxides. Hence we carried out O-substitution reactions of partially protected myo-inositol derivatives by generating their alkoxides using butyllithium and compared the results with the coresponding reaction where sodium hydride was used to generate the alkoxides. Discovery of reagents and reaction conditions to obtain good regioselectivity in O-substitution reactions of partially protected myo-inositol derivatives would be very useful, as seen in the chapter 1 of this thesis. This is especially appropriate since the work described in the previous section allowed sequential regioselective protection of C2-, C4- and C6-hydroxyl groups of myoinositol and the availability of a general method for the regioselective protection of C1-, C3- and C5-hydroxyl groups would allow modification of all the six hydroxyl groups of myo-inositol at will (Scheme 2B.1).


In myo-inositol derived 1,3,5-triols (278), C1- and C3-hydroxyl groups have one cis and one trans vicinal oxygen atom, whereas the C5-hydroxyl group has both trans vicinal oxygen atoms. This difference is known to lead to some regioselectivity in the
reactions of these hydroxyl groups (Section 1.2.2, Chapter 1) but the known methods are not good enough. Hence we chose a symmetric 1,3,5-triol (278, $\left.R^{2}=R^{4}=R^{6}=B n\right)$ for our investigation since the regioselective monosubstitution at C1- as well as C3positions would lead to a single racemic product. A similar reaction of the myo-inositol derived 1,3,5-triols carrying different substituents at C4- and C6-positions would lead to two different diastereomeric products (Scheme 2B.2).


The following section deals with the orthogonal protection of the triol 76 (Scheme 2B.3) and the regioselective protection of some diols derived from myo-inositol, using butyllithium as well as sodium hydride as base under identical conditions. We have carried out O-substitution in a few myo-inositol derived diols to see the generality as well as to provide better method for the regiospecific protection of diols known to be useful for the preparation of inositol derivatives. We have chosen vicinal as well as non-vicinal diols having hydroxyl groups with different relative orientations.

## 2B.2. Results and discussion

## 2B.2.1. O-Alkylation of 2,4,6-tri-O-benzyl-myo-inositol

Reaction of the triol $7{ }^{1}$ with one equivalent of butyllithium followed by one equivalent of allyl bromide resulted in mono allylation at the C1-hydroxyl group. This was confirmed by the ${ }^{1} \mathrm{H}$ NMR spectrum of the diacetate derivative $\mathbf{2 8 4}$ obtained by the acetylation of the monoallyl ether 282. The ${ }^{1} \mathrm{H}$ NMR spectrum of the diacetate $\mathbf{2 8 4}$ showed two distinct acetate methyl peaks at $\delta 1.93$ and 1.85 in the ratio $1: 1$ because of
its unsymmetrical nature. The ${ }^{1} \mathrm{H}$ NMR spectrum of the tribenzyl triacetate $\mathbf{2 8 5}$ showed two peaks at $\delta 1.94$ and 1.87 in the ratio $2: 1$ because of the symmetry of the molecule. This clearly showed that allylation had taken place at the $\mathrm{C} 1(3)$-hydroxyl group. Had the allylation of the triol 76 taken place at the C5-hydroxyl group, the resulting diacetate 287 would have shown only one peak for the acetate methyl hydrogens. Presence of a mixture of C1- and C5-allyl ethers (282 and 286) would have been indicated by unequal integral ratios for the acetate methyl group signals in the ${ }^{1} \mathrm{H}$ NMR spectrum of a mixture of acetates 284 and 287.


Scheme 2B.3. a) n-BuLi, THF; (b) AllBr, DMF; c) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr; d) NaH , THF. * Estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy.

Allylation of the triol 76 with one equivalent of sodium hydride and allyl bromide resulted in a mixture of products which was further acetylated. The ratio of the two allyl ethers $\mathbf{2 8 4}$ and $\mathbf{2 8 7}$ in the mixture was estimated to be $1: 1$ by comparing the integrals of the acetate methyl peaks in its ${ }^{1} \mathrm{H}$ NMR spectrum.

2B.2.2. O-Alkylation / tosylation of diols. O-Tosylation of racemic 1-O-allyl-2,4,6-tri-O-benzyl-myo-inositol


Scheme 2B.4. a) n-BuLi, THF; (b) TsCl, DMF; c) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr.

Reaction of the racemic 3,5-diol $\mathbf{2 8 2}$ with one equivalent of butyllithium followed by one equivalent of tosyl chloride resulted in the corresponding C3-tosylate 288. The ${ }^{1} \mathrm{H}$ NMR spectrum of the acetate obtained from the tosylate $\mathbf{2 8 8}$ showed a single acetate methyl peak at $\delta 1.75$. This confirmed that tosylation in 282 had occurred selectively at the C3-hydroxyl group. Hence O-substitution in lithium alkoxides of the triol 76 allows the sequential substitution of both C1- and C3-hydroxyl groups that are cis to the C2oxygen leaving the C5-hydroxyl group undisturbed.

## 2B.2.3. O-Alkylation of racemic 1,2:4,5-di-O-isopropylidene-myo-inositol

Benzylation of the racemic 3,6-diol $\mathbf{6 0}^{2}$ with one equivalent of butyllithium and one equivalent of benzyl bromide (or sulfonylation with tosyl chloride) resulted in the corresponding C3-O-substituted derivative in good yield (Scheme 2B.5). No reaction was observed at the C6-hydroxyl group of $\mathbf{6 0}$.


Scheme 2B.5. a) n-BuLi, THF; b) BnBr, DMF; c) TsCl, DMF; d) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr.

Reaction of the racemic 3,6-diol $\mathbf{6 0}$ with electrophiles (alkyl halides, sulfonyl halides, acyl halides) under a variety of reaction conditions is reported ${ }^{3}$ to give a mixture of products. Benzylation of the racemic 3,6 -diol $\mathbf{6 0}$ with sodium hydride and benzyl bromide is known ${ }^{3}$ to give a mixture of monobenzyl ethers $(\mathbf{1 5 4}, \mathbf{1 5 5})$ and the dibenzyl ether 156 (15\%) from which the monobenzyl ether 154 (56\%) was isolated by crystallization. Reaction of the racemic 3,6 -diol $\mathbf{6 0}$ with tosyl chloride is reported ${ }^{4}$ to give the 3-tosylate $\mathbf{1 6 1}$ in $30 \%$ yield. A comparison of these results with the present work clearly shows that better regioselectivity during O-substitution in the diol $\mathbf{6 0}$ can be achieved by generating the alkoxide with butyllithium. Use of lithium hydride as a base for the benzylation of the racemic 3,6-diol $\mathbf{6 0}$ gave a mixture of products (as indicated by TLC). This reaction was carried out at a higher temperatue $\left(55^{\circ} \mathrm{C}\right)$ due to the low basicity of lithium hydride.

## 2B.2.4. O-Alkylation of racemic 2,3,4,5-tetra-O-benzyl-myo-inositol

The racemic 1,6-diol 295 was prepared by a slightly modified procedure reported ${ }^{5}$ for the preparation of an enantiomer D297 of the 1,6-diol (Scheme 2B.6, $292 \rightarrow \rightarrow$ D297). The racemic diol 295 was prepared from the triol 292 by first protecting the vicinal diols as the corresponding isopropylidene derivative. The ketal 293 was prepared by the reaction of the triol $\mathbf{2 9 2}$ with 2,2-dimethoxypropane in the presence of CSA. Our initial attempt to obtain this ketal ${ }^{5}$ by using TsOH failed. The resulting isopropylidene derivative 293 was benzylated at the C3-hydroxyl group using sodium hydride and benzyl bromide. The isopropylidene group in the crude product 294 was cleaved by acid hydrolysis which afforded the racemic 1,6-diol 295.


Diastereomeric mixture
Scheme 2B.6. a) 2,2-Dimethoxypropane, CSA, DCM, reflux; b) $\mathrm{NaH}, \mathrm{BnBr}, \mathrm{DMF}$; c) $\mathrm{HCl}, \mathrm{MeOH}, \mathrm{DCM}$; e) $\mathrm{AcCl}, \mathrm{DCM}, \mathrm{MeOH}$.

Preparation of racemic 1,6-diol 295 from the dicyclohexylidene derivative of myoinositol is reported. ${ }^{6}$ We did not follow this route due to practical difficulties in this method and the lower overall yield of 295 .

Allylation of the racemic 1,6-diol 295 using butyllithium as the base to generate the alkoxide yielded the corresponding C1-allyl ether 298 as the major product (Scheme 2B.7). The mixture of monoallyl ethers 298 and 299 was converted to the mixture of the corresponding acetates and their ratio ( $\mathbf{3 0 0}: \mathbf{3 0 1}=80: 20$ ) was estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy. Use of sodium hydride to generate the alkoxide for allylation of the 1,6-diol 295 gave a mixture of monoethers in the ratio $2: 1$. This was revealed by the ${ }^{1} \mathrm{H}$ NMR spectrum of the mixture of acetates $(\mathbf{3 0 0}: \mathbf{3 0 1}=66: 33)$. These results show that the regioselectivity for the allylation at the C1-hydroxyl group in 1,6-diol (using DMF as the solvent) is better when lithium alkoxide is used for the reaction. Allylation of one of the enatiomers of the 1,6-diol D297 (Chapter 1, section 1.3.4) in the presence of sodium hydride in THF is reported ${ }^{7}$ to give a mixture of C1- and C6-ethers in the ratio 7:1 in an overall yield of $56 \%$.


Scheme 2B.7. a) n-BuLi, THF; b) AllBr, DMF; c) NaH , THF; d) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr.

* Estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy.


## 2B.2.5. O-Alkylation of racemic 3,4,5,6-tetra-O-benzyl-myo-inositol

Benzylation of the racemic 1,2 -diol $\mathbf{1 1 1}^{8}$ (Scheme 2B.8) in DMF at $0{ }^{\circ} \mathrm{C}$, using butyllithium to generate the alkoxide, gave the pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ as the major product along with a small amount of the hexabenzyl ether 302. The mixture of isomeric pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ was seperated by column chromatography and a portion of this mixture was acetylated to obtain a mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8} .{ }^{1} \mathrm{H}$ NMR spectrum of the mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ showed that the two pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ were present in more or less equal amounts. This was revealed by a comparison of the integrals of the acetate methyl group peaks in the ${ }^{1} \mathrm{H}$ NMR spectrum of the mixture of acetates $\mathbf{3 0 7}$ and 308. The peak corresponding to the C 2 -acetate was identified by comparison with the ${ }^{1} \mathrm{H}$ NMR spectrum of the pentabenzyl-2-acetate $\mathbf{3 0 7}$ (acetate methyl peak at $\delta 2.17$ ) synthesized following a literature procedure. ${ }^{8}$


Scheme 2B.8. a) n-BuLi, THF; b) BnBr, DMF; c) NaH , THF; d) AllBr, DMF; e) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr.

* Estimated by 1H NMR spectroscopy.

Since the pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ had the same R $_{f}$ value on TLC (eluent: $15 \%$ ethyl acetate / petroleum ether) we initially thought that it was a single isomer and obtained good quality crystals by crystallization from a mixture of ethyl acetate and light petroleum. Analysis of one of the crystals revealed it to be the C2-benzyl ether 304 (Figure 2B.1). However, as explained above, the product actually consisted of a mixture of two isomeric pentabenzyl ethers.


Figure 2B.1. ORTEP of 304

Benzylation of the racemic 1,2-diol $\mathbf{1 1 1}$ was also carried out at $-78^{\circ} \mathrm{C}$ and $75^{\circ} \mathrm{C}$ using BuLi to generate the alkoxide. These reactions also gave a mixture of pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ but the relative proportion of the equatorial C1-benzyl ether increased at higher temperatures (Scheme 2B.8). Benzylation of the racemic 1,2diol 111 using sodium hydride as the base at $0{ }^{\circ} \mathrm{C}$ followed by acetylation gave a mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ in the ratio 64:36. This experiment showed that benzylation was predominant at the C 1 -hydroxyl group in contrast to the benzylation when butyllithium was used as the base. We also carried out the allylation on the racemic diol $\mathbf{1 1 1}$ (using butyllithium as the base) at $0^{\circ} \mathrm{C}$ and estimated the ratio of products (67:33) (after acetylation of the mixture of monoallyl ethers 305 and 306) by ${ }^{1} \mathrm{H}$ NMR spectroscopy.

## 2B.2.6. O-Alkylation of racemic 1,2-O-isopropylidene-3,6-di-O-benzyl-myo-inositol

 Allylation of the racemic 4,5-diol 311 ${ }^{3 \mathrm{a}}$ (Scheme 2B.9) by sequential reaction with butyllithium and allyl bromide resulted in a mixture of products. The mixture of monoethers $\mathbf{3 1 2}$ and $\mathbf{3 1 3}$ was converted to the corresponding acetates $\mathbf{3 1 4}$ and $\mathbf{3 1 5}$ and their ratio (314:315 $=50: 50$ ) was estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy. We did not carry out the alkylation of the racemic diol $\mathbf{3 1 1}$ using the corresponding sodium alkoxide since the reaction of its lithium alkoxide yielded a $1: 1$ mixture of products. Also alkylation of a diol 180 (1-deoxy-1-fluoro-2,3-isopropylidene-4-O-benzyl-myoinositol) which is similar to $\mathbf{3 1 1}$ using sodium hydride is reported ${ }^{9}$ to give a mixture of both the possible isomeric ethers.

Scheme 2B.9. a) n-BuLi, THF; b) AllBr, DMF; c) $\mathrm{Ac}_{2} \mathrm{O}$, Pyr; d) PMBCI, NaH, DMF.

* Estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy.

A comparison of the results on O-substitution of the triol 76 and the diols $\mathbf{6 0}, \mathbf{1 1 1}$, 282, 295 and 311 clearly shows that O-substitution in their lithium alkoxides preferentially takes place at a hydroxyl group that has a neighbouring cis-oxygen atom while the same reactions using sodium alkoxides are not that selective. This is supported by the fact that alkylation of the racemic 4,5-diol $\mathbf{3 1 1}$ (with butyllithium and allyl bromide) in which there is no adjacent cis-oxygen to either of the hydroxyl groups result in equal amounts of both the possible allyl ethers. Chelation assisted regioselective O -substitution of vicinal diols (using heavier metals) ${ }^{10}$ have earlier been reported (Scheme 2B.10).


319

322 (63\%)


323 (61\%)


321


Scheme 2B.10. a) $\mathrm{Bu}_{2} \mathrm{SnO}$ (or) $\mathrm{Bu}_{2} \mathrm{Sn}(\mathrm{OMe})_{2}$; b) (i) $\mathrm{R}^{1} \mathrm{X}$, (ii) $\mathrm{H}_{2} \mathrm{O}$; c) $\mathrm{NaH}, \mathrm{NiCl}_{2}, \mathrm{BnBr}, \mathrm{THF}, \mathrm{DMF}$.

The results presented in Schemes 2B. 3 - 2B. 5 and 2B. 7 - 2B. 9 can be rationalized based on the relative stability of the alkoxides (Scheme 2B.11) generated by the reaction of the myo-inositol derived diols and triol with butyllithium or sodium hydride.

Scheme 2B. 11 shows all the possible alkoxides that can be generated by the reaction of the myo-inositol derived triol (76) and the diols (60, 111, 282, 295 and 311) with butyllithium or sodium hydride. We can presume that the metal ions in all these alkoxides are chelated to neighboring oxygen atoms present in the molecule. As seen in Section A of this chapter, the alkoxides of these diols and triols would be present as an equilibrium mixture of the possible chelates. The relative proportion of individual chelates present in a reaction mixture at a given temperature would depend on their relative stability.







| 359 Li | 361 Li |
| :--- | :--- |
| 360 Na | 362 Na |



Scheme 2B.11. a) DMF, n-BuLi; b) DMF, NaH
We can assume that the O-substitution reaction takes place principally through these single molecular entities and the amount of the molecular aggregates present in the reaction mixture is negligible. This is because the O-substitution reaction in THF was
very sluggish while the same reaction in DMF - THF mixture proceeded smoothly (Section 2A. 2 of this chapter).

The factors that could contribute to the relative stability of the alkoxides $\mathbf{3 2 5 - 3 7 0}$ are (a) the nature of the metal ion involved and (b) the relative orientation of the vicinal oxygen atoms in a myo-inositol derivative, which can chelate with the metal ion. In reactions involving lithium alkoxides, it is conceivable that the chelation from a vicinal cis-oxygen atom is better than the chelation from a vicinal trans-oxygen atom. ${ }^{11}$ Hence the relative stability of lithium chelates is expected to decrease in the order 325 $>\mathbf{3 2 7} \sim 329$ for 76; 331 $>333 \sim 335 \sim 337$ for 282; 339 $>\mathbf{3 4 1}$ 343~345 for $\mathbf{6 0}$; 347 $>349 \sim 351 \sim 353$ for 295; 355~357~359 $>361$ for 111; 363~365~367~369 for 311. This implies larger proportion of the O-substitution reaction proceeding through the chelates $\mathbf{3 2 5}, \mathbf{3 3 1}, \mathbf{3 3 9}, \mathbf{3 4 7}, \mathbf{3 5 5}, \mathbf{3 5 7}$ or $\mathbf{3 5 9}$ and hence formation of the products resulting from O-substitution at a hydroxyl group which has a vicinal cis-oxygen atom. The relative stability of the chelates formed from sodium alkoxide is not expected to be very different since sodium ion is not as good a chelator as lithium ion. Hence reaction of sodium alkoxide with alkyl halides result in poorer regioselectivity.

In the case of the racemic 1,2-diol 111, the alkoxide at the C2-position is stabilized by two vicinal cis-oxygen atoms ( $\mathbf{3 5 5}$ and 357) , whereas the C1-alkoxide is stabilized by chelation due to one vicinal cis-oxygen atom (359) and one vicinal trans-oxygen atom (361). This could be the reason for the formation of larger amounts of the C 2 -ether from lithium alkoxides. Perhaps with increase in temperature the difference in the stability of lithium chelates become narrower leading to the formation of relatively larger amounts of the C 1 -ether at higher temperatures. As discussed above, the relative stability of the chelates formed from sodium alkoxide is not expected to be very different and this results in the formation of larger amounts of the C 1 -ether due to steric
reasons (reaction at an axial hydroxyl group versus reaction at an equatorial hydroxyl group). These results seem to suggest that the O-substitution reaction in the diol $\mathbf{1 1 1}$ does depend on the realtive stability of the chelates since reducing the relative stability of chelates either by increasing the temperature (in the case of lithium chelates) or by changing the metal ion (sodium chelates) leads to similar trend (more reaction at the equatorial C1-hydroxyl group) in observed regioselectivity (Scheme 2B.8).

The relative stability of chelates (or the strength of chelation) depicted in Scheme 2B. 11 is analogous to the relative strength of intramolecular hydrogen bonding in polyhydroxy cyclohexanes which is known ${ }^{11 \mathrm{ab}}$ to decrease as we go from 1,2-cis diols to 1,2-trans diols. Regioselectivity in the reactions of vicinal diols have earlier been correlated with the strength of intramolecular hydrogen bonding in carbohydrate derivatives. Knapp and coworkers reported ${ }^{12}$ the regioselective triflation of carbohydrate derived vicinal diols 319, 320, 371 and 372 (Scheme 2B.12) having hydroxyl groups in different relative orientations. Among the two vicinal hydroxyl groups, exclusive triflation took place regioselectively at the hydroxyl group which was cis to the vicinal ether oxygen ( $\mathbf{3 1 9}$ and 320). When both the hydroxyl groups were either cis or trans to the vicinal ether oxygen (371 and 372), then the triflation gave a mixture of products.


Scheme 2B.12. a) $\mathrm{Tf}_{2} \mathrm{O}$, Pyr, DCM, $-20^{\circ} \mathrm{C}$. * Estimated by ${ }^{1} \mathrm{H}$ NMR specrtrpscopy
They also investigated hydrogen bonding interactions between the hydroxyl groups in compounds ( $\mathbf{3 1 9}, \mathbf{3 2 0}, \mathbf{3 7 1}$ and $\mathbf{3 7 2}$ ) subjected to triflation, by ${ }^{1} \mathrm{H}$ NMR spectroscopy (Chart 2B.1). The carbohydrate derived vicinal diols 319, 320, 371 and 372 with a neighbouring cis oxygen atom showed larger coupling constant ( $7-10 \mathrm{~Hz}$, implying stronger hydrogen bond) than those that did not have a neighboring cis oxygen atom (coupling constant 1-3 Hz implying weaker hydrogen bond). Hence they were able to correlate the observed regioselectivity (for triflation) among the carbohydrate derived vicinal diols with the strength of the intramolecular hydrogen bonding interaction in carbohydrate derived diols $\mathbf{3 1 9 ,} \mathbf{3 2 0}, \mathbf{3 7 1}$ and $\mathbf{3 7 2}$.


319

$J_{A B}=8.95 \mathrm{~Hz}$


320

$J_{A B}=2.36 \mathrm{~Hz} \quad J_{A B}=2.25 \mathrm{~Hz}$
372
371

## Chart 2B. 1

A comparison of the regioselectivity observed in the reactions of myo-inositol orthoester derivatives (Section 2A) and the less rigid derivatives discussed in this section reveals that the orthoester derivatives exhibited better regioselectivity in their O-substitution reactions. This can again be explained based on the differences in the strength of chelation of lithium ions by 1,3-diaxial oxygen atoms (in myo-inositol orthoesters) as against 1,2-cis or 1,2-trans oxygen atoms in myo-inositol derivatives. Again, this correlates with the relative strength of the intramolecular hydrogen bonding between the 1,3-diaxial diol in myo-inositol orthoesters ${ }^{1 \mathrm{lb}, \mathrm{c}}$ and the diols investigated in the present section.

## 2B.3. Conclusions

We have developed a method for the preferential O-substitution of hydroxyl groups that have an adjacent cis oxygen atom in myo-inositol derived triol and diols. This methodology has the potential to be useful for the selective derivatization of hydroxyl groups in polyols and their derivatives. We did not carry out acylation in the present study (Section B) due to the potential of hydroxyl esters to undergo inter or intra molecular acyl migration with ease, ${ }^{13}$ which would hamper conclusions on the observed regioselectivity. The results described in this section in principle, should provide a convenient method for the preparation of myo-inositol derivatives carrying orthogonal protecting groups at C1-, C3- and C5-positions and hence allow their modification. The results presented in both the sections of this chapter provide convenient methods for the modification of all the six hydroxyl groups of myo-inositol and hence aid in the preparation of a variety of cyclitol derivatives (Scheme 2B.13).


## 2B.4. Experimental section

General: General experimental procedures are same as in section A of this chapter except for the following: A stock solution of n-butyllithium ( $0.82-1.62 \mathrm{M}$ ) in dry hexanes was prepared and used in all the experiments. myo-Inositol derivatives $\mathbf{6 0},{ }^{2 \mathrm{a}, \mathrm{b}}$ $\mathbf{7 6},{ }^{1} \mathbf{1 1 1},{ }^{8} \mathbf{3 0 7},{ }^{8} \mathbf{3 1 0}{ }^{3 \mathrm{a}}$ were prepared according to literature procedures. Compounds $\mathbf{2 8 4}, \mathbf{2 8 5}, \mathbf{2 9 0}, 291$ were not characterized rigorously since they were not used further in any experiment.

General procedure for the $\mathbf{O}$-alkylation and O -sulfonylation of myo-inositol derived triol and diols.

The required myo-inositol derived triol or diol ( $0.5-1 \mathrm{mmol}$ ) was dissolved in dry THF (3-6 mL) and cooled to $0^{\circ} \mathrm{C}$. n -Butyllithium (1.0-1.2 mmol) was added drop-wise using a syringe followed by a solution of the required alkyl halide or sulfonyl halide (1.2-1.5 mmol) in DMF $(0.5-1 \mathrm{~mL})$ at $0^{\circ} \mathrm{C}$. The reaction mixture was stirred for $20-$ 56 h at room temperature and worked up as usual with ethyl acetate. The products were separated by column chromatography using 10-20\% ethyl acetate - light petroleum or light petroleum - DCM mixtures as eluents. In some experiments, the ratio of the mixture of ethers formed was estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy.

The same procedure was followed for the reactions where sodium hydride was used to generate the alkoxides, but the ratio of the products was estimated by ${ }^{1} \mathrm{H}$ NMR spectroscopy.

## Racemic 1-O-allyl-2,4,6-tri-O-benzyl-myo-inositol (282).

The $1,3,5$-triol $76(0.450 \mathrm{~g}, 1 \mathrm{mmol})$ was allylated as in the general procedure (reaction time 28 h ), using butyllithium ( $1.46 \mathrm{~mL}, 1.20 \mathrm{mmol}$ ) and allyl bromide $(0.182 \mathrm{~g}, 1.50 \mathrm{mmol})$ to obtain the racemic 1-allyl ether $282(0.351 \mathrm{~g}, 71 \%)$ as a gum and the diallyl ether $\mathbf{2 8 3}$ ( $0.089 \mathrm{~g}, 17 \%$ ) as solid, after column chromatography.

## Data For 282:

IR ( $v$, neat): 3196-3641 $\mathrm{cm}^{-1}$.
${ }^{1}$ H NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.45(\mathrm{~m}, 15 \mathrm{H}), 5.80-6.05(\mathrm{~m}, 1 \mathrm{H}), 5.26-5.39(\mathrm{~m}, 1 \mathrm{H}), 5.14-$ $5.24(\mathrm{~m}, 1 \mathrm{H}), 4.68-5.04(\mathrm{~m}, 6 \mathrm{H}), 4.10-4.18(\mathrm{~m}, 2 \mathrm{H}), 4.04(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=2.4 \mathrm{~Hz}), 3.85(\mathrm{t}$, $1 \mathrm{H}, \mathrm{J}=9.4 \mathrm{~Hz}), 3.68(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.4 \mathrm{~Hz}), 3.51(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=9.0 \mathrm{~Hz}), 3.35(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=$ $2.4,9.8 \mathrm{~Hz}), 2.52\left(\mathrm{~s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), $3.33\left(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=5.9 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 134.6,130.5,124.3,124.2,123.9,123.6,123.5,112.8,73.5$, 72.9, 72.3, 71.3, 70.8, 68.0, 67.3.

Elemental analysis calcd for $\mathrm{C}_{30} \mathrm{H}_{34} \mathrm{O}_{6}$ : C, 73.45; H, 6.99. Found: C, 73.07; H, 6.89. Data for 283:
mp. $93-95^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.26-7.50(\mathrm{~m}, 15 \mathrm{H}), 5.81-6.03(\mathrm{~m}, 2 \mathrm{H}), 5.33-5.38(\mathrm{~m}, 1 \mathrm{H}), 5.25-$ $5.30(\mathrm{~m}, 1 \mathrm{H}), 5.18-5.23(\mathrm{~m}, 1 \mathrm{H}), 5.13-5.17(\mathrm{~m}, 1 \mathrm{H}), 4.72-4.97(\mathrm{~m}, 6 \mathrm{H}), 4.10(\mathrm{dt}, 4 \mathrm{H}$, $\mathrm{J}=1.5,5.3 \mathrm{~Hz}), 4.02-4.06(\mathrm{~m}, 1 \mathrm{H}), 3.82-3.95(\mathrm{~m}, 2 \mathrm{H}), 3.42-3.55(\mathrm{~m}, 1 \mathrm{H}), 3.25(\mathrm{dd}$, $2 \mathrm{H}, \mathrm{J}=2.4,9.7 \mathrm{~Hz}$ ), 2.43-2.53 (broad s, 1H).

## Racemic 1-O-allyl-2,4,6-tri-O-benzyl-3,5-di-O-acetyl-myo-inositol (284).

The racemic allyl ether $282(0.030 \mathrm{~g}, 0.06 \mathrm{mmol})$ was acetylated using acetic anhydride $(0.06 \mathrm{~mL}, 0.60 \mathrm{mmol})$ and pyridine $(1 \mathrm{~mL})$ overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave the diacetate 284 (0.034 g, $98 \%$ ) as a gum.

## Data for 284:

$\mathbf{I R}\left(v, \mathrm{CHCl}_{3}\right): 1745 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.43(\mathrm{~m}, 15 \mathrm{H}), 5.81-6.03(\mathrm{~m}, 1 \mathrm{H}), 5.26-5.31(\mathrm{~m}, 1 \mathrm{H}), 5.16-$ $5.26(\mathrm{~m}, 1 \mathrm{H}), 5.09(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.6 \mathrm{~Hz}), 4.81-4.93(\mathrm{~m}, 2 \mathrm{H}), 4.74(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2.5,10.2$
$\mathrm{Hz}), 4.52-4.70(\mathrm{~m}, 4 \mathrm{H}), 4.15(\mathrm{dt}, 2 \mathrm{H}, \mathrm{J}=1.4,5.5 \mathrm{~Hz}), 4.05-4.11(\mathrm{~m}, 1 \mathrm{H}), 3.88-4.03$ $(\mathrm{m}, 2 \mathrm{H}), 3.48(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2.2,9.8 \mathrm{~Hz}), 1.93(\mathrm{~s}, 3 \mathrm{H}), 1.85(\mathrm{~s}, 3 \mathrm{H})$.

## 1,3,5-tri-O-acetyl-2,4,6-tri-O-benzyl-myo-inositol (285).

The $1,3,5$-triol $76(0.050 \mathrm{~g}, 0.11 \mathrm{mmol})$ was acetylated using acetic anhydride ( 0.3 $\mathrm{mL}, 3.30 \mathrm{mmol}$ ) and pyridine ( 1 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave the triacetate 285 ( $0.063 \mathrm{~g}, 98 \%$ ).

## Data for 285:

mp. 117-119 ${ }^{\circ} \mathrm{C}$.
IR ( $v, \mathrm{CHCl}_{3}$ ): $1749 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.19-7.44(\mathrm{~m}, 15 \mathrm{H}), 5.19(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.6 \mathrm{~Hz}), 4.94(\mathrm{dd}, 2 \mathrm{H}, \mathrm{J}=$ $2.5,10.2 \mathrm{~Hz}), 4.53-4.71(\mathrm{~m}, 6 \mathrm{H}), 4.11(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=2.5 \mathrm{~Hz}), 4.03(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=10 \mathrm{~Hz}), 1.94$ (s, 6H), 1.87 (s, 3H).

## Racemic 1-O-allyl-2,4,6-tri-O-benzyl-3-O-(p-toluenesulfonyl)-myo-inositol (288).

The racemic allyl ether $\mathbf{2 8 2}(0.254 \mathrm{~g}, 0.52 \mathrm{mmol})$ was sulfonylated as in the general procedure (reaction time 15 h ), using butyllithium ( $0.76 \mathrm{~mL}, 0.62 \mathrm{mmol}$ ) and $p$ toluenesulfonyl chloride $(0.148 \mathrm{~g}, 0.78 \mathrm{mmol})$ to obtain the racemic 3-tosylate $\mathbf{2 8 8}$ $(0.179 \mathrm{~g}, 53 \%)$ as a gum, after column chromatography.

## Data for 288:

IR ( $v$, neat): 3216-3630 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.73(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8 \mathrm{~Hz}), 7.05-7.48(\mathrm{~m}, 17 \mathrm{H}), 5.75-6.01(\mathrm{~m}, 1 \mathrm{H})$, 5.24-5.37 (m, 1H), 5.13-5.23 (m, 1H), 4.66-4.96 (m, 4H), 4.55 (AB q, 2H, J = 11.4 $\mathrm{Hz}), 4.37(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2.3,10.2 \mathrm{~Hz}), 4.25(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=2.4 \mathrm{~Hz}), 4.08(\mathrm{dt}, 2 \mathrm{H}, \mathrm{J}=1.5,5.5$ $\mathrm{Hz}), 3.84(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=9.3 \mathrm{~Hz}), 3.44(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.4 \mathrm{~Hz}), 3.32(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2.3,9.8$ $\mathrm{Hz}), 2.43\left(\mathrm{~s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), $2.34(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 141.3,136.3,135.1,131.1,130.4,126.3,125.0,124.8,124.4$, $124.2,124.1,113.5,75.3,74.0,73.6,73.3,73.2,71.9,71.8,71.6,71.3,68.1,18.1$.

Elemental analysis calcd for $\mathrm{C}_{37} \mathrm{H}_{40} \mathrm{O}_{8} \mathrm{~S} .2 .3 \mathrm{H}_{2} \mathrm{O}: \mathrm{C}, 64.76$; H, 6.55. Found: C, 64.38; H, 6.07.

## Racemic

## 1-O-allyl-2,4,6-tri-O-benzyl-3-O-(p-toluenesulfonyl)-5-O-acetyl-myo-

 inositol (289).The racemic 3-tosylate $288(0.020 \mathrm{~g}, 0.03 \mathrm{mmol})$ was acetylated using acetic anhydride ( $0.04 \mathrm{~mL}, 0.46 \mathrm{mmol}$ ) and pyridine ( 0.7 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave the acetate $\mathbf{2 8 9}(0.021 \mathrm{~g}, 99 \%)$ as gum.

## Data for 289:

IR ( $v$, neat): 3193-3520, $1747 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.74(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.4 \mathrm{~Hz}), 7.02-7.54(\mathrm{~m}, 17 \mathrm{H}), 5.82-$ $6.01(\mathrm{~m}, 1 \mathrm{H}), 5.29-5.41(\mathrm{~m}, 1 \mathrm{H}), 5.19-5.28(\mathrm{~m}, 1 \mathrm{H}), 4.80-5.06(\mathrm{~m}, 4 \mathrm{H}), 4.56-4.65(\mathrm{~m}$, $1 \mathrm{H}), 4.32-4.46(\mathrm{~m}, 4 \mathrm{H}), 4.14(\mathrm{dt}, 2 \mathrm{H}, \mathrm{J}=1.5,5.5 \mathrm{~Hz}), 3.93(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=9.9 \mathrm{~Hz})$, $3.44(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2.2,9.6 \mathrm{~Hz}), 2.34(\mathrm{~s}, 3 \mathrm{H}), 1.75(\mathrm{~s}, 3 \mathrm{H})$.

Elemental analysis calcd for $\mathrm{C}_{39} \mathrm{H}_{42} \mathrm{O}_{9} \mathrm{~S} . \mathrm{H}_{2} \mathrm{O}: \mathrm{C}, 66.46 ; \mathrm{H}, 6.29$. Found: C, $66.78 ; \mathrm{H}$, 6.04.

Reaction of 2,4,6-tri-O-benzyl-myo-inositol (76) with sodium hydride / allyl bromide.

The $1,3,5$-triol $76(0.100 \mathrm{~g}, 0.22 \mathrm{mmol})$ was allylated as in the general procedure (reaction time 2 h ), using sodium hydride ( $0.011 \mathrm{~g}, 0.27 \mathrm{mmol}$ ) and allyl bromide $(0.040 \mathrm{~g}, 0.33 \mathrm{mmol})$ to obtain a mixture of allyl ethers. This mixture was acetylated using acetic anhydride $(0.3 \mathrm{~mL})$ and pyridine $(0.6 \mathrm{~mL})$ overnight at room temperature. The mixture of acetates $\mathbf{2 8 4}$ and $\mathbf{2 8 7}$ was obtained $(0.121 \mathrm{~g})$ after usual work up with
ethyl acetate. Ratio of the peak integrals at 1.93 and 1.85 in ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{2 8 4}$ and $\mathbf{2 8 7}$ were present in the ratio 50:50.

## Racemic 1,2:4,5-di-O-isopropylidene-3-O-benzyl-myo-inositol (154).

The racemic 1,4-diol $\mathbf{6 0}(0.260 \mathrm{~g}, 1 \mathrm{mmol})$ was benzylated as in the general procedure (reaction time 38 h ) using butyllithium ( $1.34 \mathrm{~mL}, 1.10 \mathrm{mmol}$ ) and benzyl bromide $(0.205 \mathrm{~g}, 1.20 \mathrm{mmol})$ to obtain the racemic 3-benzyl ether $154(0.292 \mathrm{~g}, 83 \%)$ and the dibenzyl ether $156(0.038 \mathrm{~g}, 9 \%)$, after column chromatography.
mp. (for 154) $159-161{ }^{\circ} \mathrm{C}\left(\right.$ Lit. $^{2 \mathrm{c}} \mathrm{mp} .162-163{ }^{\circ} \mathrm{C}$ ).
mp. (for 156) $146-148{ }^{\circ} \mathrm{C}\left(\right.$ Lit. $\left.^{2 \mathrm{c}} \mathrm{mp} .150-153^{\circ} \mathrm{C}\right)$.

## Racemic 1,2:4,5-di-O-isopropylidene-3-O-benzyl-6-O-acetyl-myo-inositol (290).

The racemic benzyl ether $154(0.020 \mathrm{~g}, 0.06 \mathrm{mmol})$ was acetylated using acetic anhydride $(0.03 \mathrm{~mL}, 0.30 \mathrm{mmol})$ and pyridine $(0.7 \mathrm{~mL})$ at room temperature $(12 \mathrm{~h})$. The acetate $290(0.022 \mathrm{~g}, 98 \%)$ was obtained after usual work up with ethyl acetate.

Data for 290:
mp. $170-172{ }^{\circ} \mathrm{C}\left(\right.$ Lit. $\left.{ }^{14} \mathrm{mp} .183-185^{\circ} \mathrm{C}\right)$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.55(\mathrm{~m}, 5 \mathrm{H}), 5.27(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=6.8,11.2 \mathrm{~Hz}), 4.86(\mathrm{AB} \mathrm{q}$, $2 \mathrm{H}, \mathrm{J}=12.2 \mathrm{~Hz}), 4.30(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=4.4 \mathrm{~Hz}), 4.15(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.3 \mathrm{~Hz}), 4.02(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=$ 4.8, 7.3 Hz ), $3.78(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=4.4,10.3 \mathrm{~Hz}), 3.34(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=9.3,11.2 \mathrm{~Hz}), 2.12(\mathrm{~s}$, $3 \mathrm{H}), 1.60(\mathrm{~s}, 3 \mathrm{H}), 1.49(\mathrm{~s}, 3 \mathrm{H}), 1.44(\mathrm{~s}, 3 \mathrm{H}), 1.34(\mathrm{~s}, 3 \mathrm{H})$.

Racemic 1,2:4,5-di-O-isopropylidene-3-O-(p-toluenesulfonyl)-myo-inositol (161). The racemic 1,4-diol $\mathbf{6 0}(0.261 \mathrm{~g}, 1 \mathrm{mmol})$ was sulfonylated as in the general procedure (reaction time 2 h ) using butyllithium ( $1.34 \mathrm{~mL}, 1.10 \mathrm{mmol}$ ) and $p$-toluenesulfonyl chloride ( $0.209 \mathrm{~g}, 1.10 \mathrm{mmol})$. The tosylate $161(0.283 \mathrm{~g}, 68 \%)$ was isolated as white solid by column chromatography using DCM as eluent. Since spectral data for the tosyl derivative $\mathbf{1 6 1}$ is not reported in the literature, ${ }^{4}$ we have characterised $\mathbf{1 6 1}$ completely.

## Data for 161:

mp. $164-165^{\circ} \mathrm{C}$ (lit. $.^{4} \mathrm{mp} .170-173^{\circ} \mathrm{C}$ ).
IR ( $v$, nujol): $3539 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.86(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.1 \mathrm{~Hz}), 7.33(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.1 \mathrm{~Hz}), 4.81(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}$ $=4.3,10.3 \mathrm{~Hz}), 4.41(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=4.4 \mathrm{~Hz}), 3.92-4.05(\mathrm{~m}, 2 \mathrm{H}), 3.85(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=6.6,10.3$ $\mathrm{Hz}), 3.29(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.5 \mathrm{~Hz}), 2.59\left(\right.$ broad $\mathrm{s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), $2.44(\mathrm{~s}, 3 \mathrm{H}), 1.50$ $(\mathrm{s}, 3 \mathrm{H}), 1.36(\mathrm{~s}, 3 \mathrm{H}), 1.31(\mathrm{~s}, 3 \mathrm{H}), 1.22(\mathrm{~s}, 3 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 145.7,134.1,130.0,128.4,113.1,110.8,82.2,78.4,77.6,76.0$, 74.7, 74.3, 28.0, 26.8, 26.7, 25.8, 21.7.

Elemental analysis calcd for $\mathrm{C}_{19} \mathrm{H}_{26} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 55.06 ; \mathrm{H}, 6.32$. Found: C, 55.06; $\mathrm{H}, 6.26$.
Racemic 1,2:4,5-di-O-isopropylidene-3-O-(p-toluenesulfonyl)-6-O-acetyl-myoinositol (291).

The racemic tosylate $161(0.030 \mathrm{~g}, 0.07 \mathrm{mmol})$ was acetylated using acetic anhydride $(0.07 \mathrm{~mL}, 0.72 \mathrm{mmol})$ and pyridine $(0.8 \mathrm{~mL})$ at room temperature $(12 \mathrm{~h})$. The acetate $291(0.031 \mathrm{~g}, 94 \%)$ was obtained after usual work up with ethyl acetate.

Data for 291:
mp. $214-216^{\circ} \mathrm{C}$.
IR ( $v$, nujol): $1747 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.85(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.3 \mathrm{~Hz}), 7.32(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.0 \mathrm{~Hz}), 5.22(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}$ $=6.8,11.1 \mathrm{~Hz}), 4.83(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=4.2,10.4 \mathrm{~Hz}), 4.42(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=4.5 \mathrm{~Hz}), 4.11(\mathrm{dd}, 1 \mathrm{H}$, $\mathrm{J}=2.5,4.7 \mathrm{~Hz}), 4.04-4.07(\mathrm{~m}, 1 \mathrm{H}), 3.36(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=9.4,11.2 \mathrm{~Hz}), 2.44(\mathrm{~s}, 3 \mathrm{H}), 2.11$ (s, 3H), $1.55(\mathrm{~s}, 3 \mathrm{H}), 1.35(\mathrm{~s}, 3 \mathrm{H}), 1.29(\mathrm{~s}, 3 \mathrm{H}), 1.21(\mathrm{~s}, 3 \mathrm{H})$.

Racemic 1,6-O-isopropylidene-2,4,5-tri-O-benzyl-myo-inositol (293).
The racemic tribenzyl triol $292(2.00 \mathrm{~g}, 4.44 \mathrm{mmol}$ ), 2,2-dimethoxypropane ( 1.3 mL , $10.21 \mathrm{mmol})$ and CSA ( $0.516 \mathrm{~g}, 0.22 \mathrm{mmol}$ ) were refluxed in DCM ( 60 mL ) for 3 h .

The resulting solution was cooled to room temperature and neutralized with triethyl amine. Solvents were evaporated and the solid obtained was purified by column chromatography to obtain the racemic isopropylidene derivative 293 ( $1.784 \mathrm{~g}, 75 \%$ ). Data for 293:
mp. $78-80^{\circ} \mathrm{C}$
IR ( $v$, nujol): 3115-3489 $\mathrm{cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.45(\mathrm{~m}, 15 \mathrm{H}), 4.60-5.10(\mathrm{~m}, 6 \mathrm{H}), 4.15-4.30(\mathrm{~m}, 2 \mathrm{H}), 3.56-$ $3.74(\mathrm{~m}, 3 \mathrm{H}), 3.49(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2,10 \mathrm{~Hz}), 2.40-2.50\left(\right.$ broad s, $1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), 1.49 (s, 3H), 1.47 (s, 3H).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 138.6,138.5,138.0,128.4,128.3,128.2,128.0,127.8,127.6$, $127.4,111.8,84.0,80.2,77.8,76.9,75.9,74.4,74.2,73.5,72.9,27.1,26.7$.

Elemental analysis calcd for $\mathrm{C}_{30} \mathrm{H}_{34} \mathrm{O}_{6}$. $1.2 \mathrm{H}_{2} \mathrm{O}: \mathrm{C}, 70.34 ; \mathrm{H}, 7.16$. Found: C, 70.03; H, 7.33.

## Racemic 2,3,4,5-tetra-O-benzyl-myo-inositol (295).

To a solution of the racemic ketal $293(1.500 \mathrm{~g}, 3.06 \mathrm{mmol})$ in DMF $(15 \mathrm{~mL})$ sodium hydride ( $0.184 \mathrm{~g}, 4.59 \mathrm{mmol}$ ) was added and stirred for 20 min . The reaction mixture was cooled to $0^{\circ} \mathrm{C}$ and benzyl bromide ( $1.047 \mathrm{~g}, 6.12 \mathrm{mmol}$ ) was added drop-wise and the mixture stirred for 1 h . The reaction was quenched by the addition of ice and worked up as usual with ethyl acetate. The crude product obtained $(1.020 \mathrm{~g})$ was dissolved in $\mathrm{DCM}(16 \mathrm{~mL})$ and methanol $(8 \mathrm{~mL})$ and stirred with conc $\mathrm{HCl}(0.25 \mathrm{~mL})$ for 4 h . Triethylamine was added to the reaction mixture and the solvents were evaporated under reduced pressure. The solid residue was purified by column chromatography to get the racemic 1,6-diol $295(0.761 \mathrm{~g}, 74 \%)$ as a white solid.

Data for 295:
mp. $167-169^{\circ} \mathrm{C}\left(\right.$ lit. $\left.{ }^{6 \mathrm{a}} \mathrm{mp} .162-164^{\circ} \mathrm{C}\right)$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.20-7.40(\mathrm{~m}, 20 \mathrm{H}), 4.63-5.10(\mathrm{~m}, 8 \mathrm{H}), 3.96-4.10(\mathrm{~m}, 2 \mathrm{H}), 3.76-$ $3.91(\mathrm{~m}, 1 \mathrm{H}), 3.48(\mathrm{dd}, 1 \mathrm{H}, \mathrm{J}=2,10 \mathrm{~Hz}), 3.25-3.39(\mathrm{~m}, 2 \mathrm{H}), 2.47\left(\operatorname{broad} \mathrm{~s}, 1 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), 2.30 (d, 1H, J = $8 \mathrm{~Hz}, \mathrm{D}_{2} \mathrm{O}$ exchangeable).

Elemental analysis calcd for $\mathrm{C}_{34} \mathrm{H}_{36} \mathrm{O}_{6}$ : $\mathrm{C}, 75.53 ; \mathrm{H}, 6.71$. Found: $\mathrm{C}, 75.55 ; \mathrm{H}, 6.59$.
Reaction of racemic 2,3,4,5-tetra-O-benzyl-myo-inositol (295) with butyllithium / allyl bromide.

To a solution of the racemic diol $295(0.280 \mathrm{~g}, 0.52 \mathrm{mmol})$ in THF $(3 \mathrm{~mL})$ at $0{ }^{\circ} \mathrm{C}$ butyllithium ( $0.41 \mathrm{~mL}, 0.57 \mathrm{mmol}$ ) followed by a solution of allyl bromide $(0.082 \mathrm{~g}$, $0.68 \mathrm{mmol})$ in DMF ( 0.8 mL ) were added and the mixture stirred for 56 h . The reaction mixture was worked up with ethyl acetate. The mixture of monoethers 298 and $299(0.194 \mathrm{~g}, 67 \%)$ was separated by column chromatography using $10 \%$ ethyl acetate - light petroleum as eluent. This mixture $(0.194 \mathrm{~g})$ was acetylated using acetic anhydride ( 0.6 mL ) in pyridine ( 2 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 0 0}$ and $\mathbf{3 0 1}$ $(0.203 \mathrm{~g})$. Ratio of the peak integrals at 1.95 and 2.02 in the ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 0 0}$ and $\mathbf{3 0 1}$ were present in the ratio 80:20.

Reaction of racemic 2,3,4,5-tetra-O-benzyl-myo-inositol (295) with sodium hydride / allyl bromide.

A solution of the racemic diol $295(0.100 \mathrm{~g}, 0.19 \mathrm{mmol})$ in THF ( 1 mL ) was cooled to $0{ }^{\circ} \mathrm{C}$, sodium hydride $(0.008 \mathrm{~g}, 0.21 \mathrm{mmol})$ and a solution of allyl bromide $(0.030 \mathrm{~g}$, $0.25 \mathrm{mmol})$ in DMF $(0.3 \mathrm{~mL})$ were added and stirred at room temperature for 56 h . The reaction mixture was worked up with ethyl acetate and the mixture of monoethers (298 and 299, $0.083 \mathrm{~g}, 75 \%$ ) was separated by column chromatography using $10 \%$ ethyl acetate - light petroleum as eluent. The mixture of monoethers ( 0.083 g ) was acetylated using acetic anhydride $(0.2 \mathrm{~mL})$ and pyridine $(1 \mathrm{~mL})$ at room temperature
overnight. Usual work up of the reaction mixture with ethyl acetate gave the mixture of acetates ( $\mathbf{3 0 0}$ and $\mathbf{3 0 1}, 0.086 \mathrm{~g}$ ). Ratio of the peak integrals at 1.95 and 2.02 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 0 0}$ and $\mathbf{3 0 1}$ were present in the ratio 66:33.

Benzylation of racemic 3,4,5,6-tetra-O-benzyl-myo-inositol (111) with butyllithium / benzyl bromide.

Procedure A. The racemic diol $111(0.200 \mathrm{~g}, 0.37 \mathrm{mmol})$ was benzylated as in the general procedure (reaction time $26 \mathrm{~h},-78^{\circ} \mathrm{C}$ ), using butyllithium ( $0.23 \mathrm{~mL}, 0.37$ $\mathrm{mmol})$ and benzyl bromide $(0.076 \mathrm{~g}, 0.44 \mathrm{mmol})$ to obtain a mixture of pentabenzyl ethers 303 and 304 ( $0.161 \mathrm{~g}, 69 \%$ ) and the hexaether 302 ( $0.048 \mathrm{~g}, 18 \%$ ). The mixture of pentaethers $(0.040 \mathrm{~g})$ was acetylated using acetic anhydride $(0.03 \mathrm{~mL})$ and pyridine ( 1 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}(0.041 \mathrm{~g})$. Ratio of the peak integrals at 2.16 and 1.91 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ were present in the ratio 45:55.

## Data for 302:

mp. $110-112{ }^{\circ} \mathrm{C}\left(\right.$ lit. $\left.{ }^{8} \mathrm{mp} .109-110{ }^{\circ} \mathrm{C}\right)$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.12-7.39(\mathrm{~m}, 30 \mathrm{H}), 4.70-4.90(\mathrm{~m}, 8 \mathrm{H}), 4.56(\mathrm{AB} \mathrm{q}, 4 \mathrm{H}, \mathrm{J}=11.7$ Hz), 3.92-4.09 (m, 3H), 3.41 (t, 1H, J = 9.3 Hz), $3.29(\mathrm{dd}, 2 \mathrm{H}, \mathrm{J}=2.4,9.7 \mathrm{~Hz})$.

Procedure B. The racemic diol $111(0.100 \mathrm{~g}, 0.19 \mathrm{mmol})$ was benzylated as in the general procedure (reaction time $26 \mathrm{~h}, 0^{\circ} \mathrm{C}$-rt), using butyllithium ( $0.12 \mathrm{~mL}, 0.20$ $\mathrm{mmol})$ and benzyl bromide $(0.076 \mathrm{~g}, 0.44 \mathrm{mmol})$ to obtain the mixture of pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}(0.084 \mathrm{~g}, 70 \%)$ and the hexaether $\mathbf{3 0 2}(0.020 \mathrm{~g}, 15 \%)$. The mixture of pentaethers $(0.050 \mathrm{~g})$ was acetylated using acetic anhydride $(0.04 \mathrm{~mL})$ and pyridine ( 1 mL ) overnight at room temperature. Usual work up of the reaction
mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}(0.053 \mathrm{~g})$. Ratio of the peak integrals at 2.17 and 1.92 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers 307 and $\mathbf{3 0 8}$ were present in the ratio 52:48.

Procedure C. The racemic diol $111(0.200 \mathrm{~g}, 0.37 \mathrm{mmol})$ was benzylated as in the general procedure (reaction time 26 h at reflux $\sim 76^{\circ} \mathrm{C}$ ), using butyllithium ( 0.45 mL , 0.37 mmol , added at $0{ }^{\circ} \mathrm{C}$ ) and benzyl bromide ( $0.076 \mathrm{~g}, 0.44 \mathrm{mmol}$, added at reflux temperature) to obtain a mixture of pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}$ ( $0.172 \mathrm{~g}, 74 \%$ ) and the hexaether $302(0.051 \mathrm{~g}, 19 \%)$. The mixture of pentaethers $(0.100 \mathrm{~g})$ was acetylated using acetic anhydride ( 0.15 mL ) and pyridine ( 2 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}(0.103 \mathrm{~g})$. Ratio of the peak integrals at 2.16 and 1.91 in its ${ }^{1} H$ NMR spectrum showed that the isomers $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ were present in the ratio71:29.

Benzylation of racemic 3,4,5,6-tetra-O-benzyl-myo-inositol (111) with sodium hydride / benzyl bromide.

Procedure B. The racemic diol $111(0.200 \mathrm{~g}, 0.37 \mathrm{mmol})$ was benzylated as in the general procedure (reaction time $26 \mathrm{~h}, 0^{\circ} \mathrm{C}$ ), using sodium hydride ( $0.015 \mathrm{~g}, 0.37$ $\mathrm{mmol})$ and benzyl bromide $(0.076 \mathrm{~g}, 0.44 \mathrm{mmol})$ to obtain the mixture of pentabenzyl ethers $\mathbf{3 0 3}$ and $\mathbf{3 0 4}(0.177 \mathrm{~g}, 76 \%)$ and the hexaether $\mathbf{3 0 2}(0.058 \mathrm{~g}, 22 \%)$. The mixture of pentaethers $(0.100 \mathrm{~g})$ was acetylated using acetic anhydride $(0.15 \mathrm{~mL})$ and pyridine ( 2 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave the mixture of acetates $\mathbf{3 0 7}$ and $\mathbf{3 0 8}(0.105 \mathrm{~g})$. Ratio of the peak integrals at 2.16 and 1.91 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ were present in the ratio 64:36.

## Allylation of racemic 3,4,5,6-tetra-O-benzyl-myo-inositol (111) with butyllithium

 / allyl bromide.Procedure B. The racemic diol $111(0.150 \mathrm{~g}, 0.28 \mathrm{mmol})$ was allylated as in the general procedure (reaction time $20 \mathrm{~h}, 0^{\circ} \mathrm{C}$ ), using butyllithium ( $0.19 \mathrm{~mL}, 0.28 \mathrm{mmol}$ ) and allyl bromide ( $0.044 \mathrm{~g}, 0.36 \mathrm{mmol}$ ) to obtain a mixture of allyl ethers $\mathbf{3 0 5}$ and $\mathbf{3 0 6}$ $(0.107 \mathrm{~g}, 66 \%)$. The mixture of allyl ethers $(0.032 \mathrm{~g})$ was acetylated using acetic anhydride $(0.03 \mathrm{ml})$ and pyridine $(1 \mathrm{~mL})$ overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 0 9}$ and $\mathbf{3 1 0}$ $(0.034 \mathrm{~g})$. Ratio of the peak integrals at 2.15 and 2.01 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 0 9}$ and $\mathbf{3 1 0}$ were present in the ratio 67:33.

Reaction of racemic 1,2-O-isopropylidene-3,6-di-O-benzyl-myo-inositol (311) with butyllithium / allyl bromide.

The racemic diol $311(0.200 \mathrm{~g}, 0.50 \mathrm{mmol})$ was allylated as in the general procedure (reaction time $52 \mathrm{~h}, 0^{\circ} \mathrm{C}$ ), using butyllithium ( $0.67 \mathrm{~mL}, 0.55 \mathrm{mmol}$ ) and allyl bromide $(0.091 \mathrm{~g}, 0.75 \mathrm{mmol})$ to obtain a mixture of mono allyl ethers $\mathbf{3 1 2}$ and $\mathbf{3 1 3}(0.166 \mathrm{~g}$, $76 \%$ ). The mixture of monoallyl ethers ( 0.166 g ) was acetylated using acetic anhydride $(0.4 \mathrm{~mL})$ and pyridine ( 3 mL ) overnight at room temperature. Usual work up of the reaction mixture with ethyl acetate gave a mixture of acetates $\mathbf{3 1 4}$ and $\mathbf{3 1 5}$ $(0.181 \mathrm{~g})$. Ratio of the peak integrals at 2.06 and 2.04 in its ${ }^{1} \mathrm{H}$ NMR spectrum showed that the isomers $\mathbf{3 1 4}$ and $\mathbf{3 1 5}$ were present in the ratio 50:50.

## 2B.5. References

1. Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I. M.; Vacca, J. P.; deSolms, S. J.; Huff, J. R. J. Chem. Soc., Perkin Trans. 1 1989, 1423.
2. (a) Gigg, J.; Gigg, R.; Payne, S.; Conant, R. Carbohydr. Res. 1985, 142, 132; (b) Khersonsky, S. M.; Chang, Y.-T. Carbohydr. Res. 2002, 337, 75; (c) Chung, S-K.; Ryu, Y. Carbohydr. Res. 1994, 258, 145.
3. (a) Gigg, J.; Gigg, R.; Payne, S.; Conant, R. J. Chem. Soc., Perkin Trans. 1 1987, 423; (b) Chung, S-K.; Chang, Y-T.; Ryu, Y. Pure. Appl. Chem. 1996, 68, 931.
4. Verónica, S-P.; Francisco Javier, M-M.; Rosalinda, C.; Angelina, F-P. J. Carbohydr. Chem. 1997, 16, 1479.
5. Painter, G. F.; Grove, S. J. A.; Gilbert, I. H.; Holmes, A. B.; Raithby, P. R.; Hill, M. L.; Hawkins, P. T.; Stephens, L. R. J. Chem. Soc., Perkin Trans. 1 1999, 923.
6. (a) Okajima, K.; Mukae, T.; Imagawa, H.; Kawamura, Y.; Nishizawa, M.; Yamada, H. Tetrahedron 2005, 61, 3497; (b) Ozaki, S.; Kohno, M.; Nakahira, H.; Bunya, M.; Watanabe, Y. Chem. Lett. 1988, 77.
7. Baeschlin, D. K.; Chaperon, A. R.; Green, L. G.; Hahn, M. G.; Ince, S. J.; Ley, S. V. Chem. Eur. J. 2000, 6, 172.
8. Angyal, S. J.; Tate, M. E. J. Chem. Soc. 1965, 6949.
9. Fauq, A. H.; Zaidi, J. H.; Wilcox, R. A.; Varvel, G.; Nahorski, S. R.; Kozikowski, A. P.; Erneux, C. Tetrahedron Lett. 1996, 37, 1917.
10. (a) Martinelli, M. J.; Vaidyanathan, R.; Pawlak, J. M.; Nayyar, N. K.; Dhokte, U. P.; Doecke, C. W.; Zollars, L. M. H.; Moher, E. D.; Khau, V. V.; Košmrlj, B. J. Am. Chem. Soc. 2002, 124, 3578; (b) Gangadharmath, U. B.; Demchenko, A. V. Synlett 2004, 2191.
11. (a) Bernet, B.; Vasella, A. Helv. Chem. Acta. 2000, 83, 995 and references cited therein; (b) Uhlmann, P.; Vasella, A. Helv. Chim. Acta. 1992, 75, 1979; (c) Bhosekar, G.; Murali, C.; Gonnade, R. G.; Shashidhar, M. S.; Bhadbhade, M. M. Cryst. Growth. Des. 2005, 5, 1977.
12. Knapp, S.; Kukkola, P. J.; Sharma, S.; Murali Dhar, T. G.; Naughton, A. B. J. J. Org. Chem. 1990, 55, 5700 .
13. (a) Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Das, T. Chem. Rev. 2003, 103, 4477; (b) Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2000, 41, 4185. (c) Ahn, Y-H.; Chang, Y-T. Chem. Eur. J. 2004, 10, 3543.
14. Desai, T.; Fernandez-Mayoralas, A.; Gigg, J.; Gigg, R.; Payne, S. Carbohydr. Res. 1990, 205, 105.








${ }^{1}$ H NMR spectrum for the acetylated mixture of products $\mathbf{2 8 4}$ and $\mathbf{2 8 7}$ obtained on allylation of the triol 76 with sodium hydride / allyl bromide at $0^{\circ} \mathrm{C}$.










(





${ }^{1} \mathrm{H}$ NMR spectrum for the acetylated mixture of products $\mathbf{3 0 0}$ and $\mathbf{3 0 1}$ obtained on allylation of the racemic diol $\mathbf{2 9 5}$ with butyllithium / allyl bromide at $0^{\circ} \mathrm{C}$.

${ }^{1} \mathrm{H}$ NMR spectrum for the acetylated mixture of products $\mathbf{3 0 0}$ and $\mathbf{3 0 1}$ obtained on allylation of the racemic diol 295 with sodium hydride / allyl bromide at $0^{\circ} \mathrm{C}$.


Crystal Data Table for $\mathbf{3 0 4}$

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Monoclinic, P21/c |
| Unit cell dimensions | $\mathrm{a}=5.968(2) \AA$ alpha $=90^{\circ}$ <br> $\mathrm{b}=19.946(8) \AA$ beta $=92.375(10)^{\circ}$ <br> $\mathrm{c}=28.736(10) \AA$ gamma $=90^{\circ}$ |
| Volume | $3418(2) \AA^{3}$ |
| Z, Calculated density | $4,1.226 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.081 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.242 \times 0.202 \times 0.070 \mathrm{~mm}$ |
| Theta range for data collection | 1.24 to 25.00 deg. |
| Reflections collected / unique | $32577 / 6014[\mathrm{R}(\mathrm{int})=0.0808]$ |
| Completeness to theta $=25.00$ | $100.0 \%$ |
| Refinement method | Full-matrix least-squares on F ${ }^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.021 |
| Final R indices [I>2sigma(I)] | $\mathrm{R} 1=0.0625, \mathrm{wR} 2=0.1556$ |
| R indices (all data) | 0.263 and $-0.192 \mathrm{e} . \AA^{-3}$ |
| Largest diff. peak and hole | $0.1218, \mathrm{wR} 2=0.1849$ |


${ }^{1} \mathrm{H}$ NMR spectra for the acetylated mixture of products $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ obtained on benzylation of the racemic diol $\mathbf{1 1 1}$ with butyllithium / benzyl bromide.
(a) $-78{ }^{\circ} \mathrm{C}$

(b) $0^{\circ} \mathrm{C}$

(c) reflux

${ }^{1}$ H NMR spectrum for the acetylated mixture of products $\mathbf{3 0 7}$ and $\mathbf{3 0 8}$ obtained on benzylation of the racemic diol $\mathbf{1 1 1}$ with sodium hydride / benzyl bromide at $0^{\circ} \mathrm{C}$.

${ }^{1} \mathrm{H}$ NMR spectrum for the acetylated mixture of products $\mathbf{3 0 9}$ and $\mathbf{3 1 0}$ obtained on allylation of the racemic diol $\mathbf{1 1 1}$ with butyllithium / allyl bromide at $0^{\circ} \mathrm{C}$.

${ }^{1} \mathrm{H}$ NMR spectrum for the acetylated mixture of products $\mathbf{3 1 4}$ and $\mathbf{3 1 5}$ obtained on allylation of the racemic diol $\mathbf{3 1 1}$ with butyllithium / allyl bromide at $0^{\circ} \mathrm{C}$.


### 3.1. Introduction

Previous chapters described methods for the orthogonal protection of myoinositol orthoesters (2,4,6-triols) and a myo-inositol-1,3,5-triol. Another strategy that we thought of for the protection of C1-, C3- and C5-hydroxyl groups of myo-inositol was to see if the chair conformation of the myo-inositol ring as in its orthoesters could be locked by bridging the C4- and C6-hydroxyl groups and subsequently cleave the orthoester moiety to generate a 1,3,5-triaxial triol (Scheme 3.1). Two ways in which this can be achieved are, either by preparing the dimeric structure $\mathbf{3 8 0}$ or by preparing the monomeric structure 381. Availability of such triols would allow a study of the relative reactivity of the three axially disposed C1-, C3- and C5-hydroxyl groups and perhaps the preparation of orthogonally protected myo-inositol derivatives.


Apart from this synthetic interest, axially disposed 1,3,5-cyclohexane triols have other properties like metal complexing ability ${ }^{1}$ and can serve as ionophores. ${ }^{2}$ Also, natural products like muellitol (383, Chart 3.1) and other unnatural products like cis-inositol ${ }^{3}$ (43) contain triaxially disposed hydroxyl groups in a cyclohexane ring.
scyllo-Inositol orthoformate which is another molecule containing triaxial triol has been used for the preparation of enterobactin analog $16 .{ }^{4}$


16 Enterobactin analog, $\mathrm{R}^{1}=\mathrm{n}-\mathrm{C}_{10} \mathrm{H}_{21}$


383 Muellitol


43 cis -Inositol


384 Enterobactin

Chart 3.1
Angyal reported ${ }^{1}$ the synthesis of the 1,3,5-triol $\mathbf{3 8 8}$ from myo-inositol orthoformate as shown in Scheme 3.2. Angyal also found that the methylene bridged derivative (formaldehyde acetal of 385) cannot be prepared by the cyclization of the diol $\mathbf{3 8 5}$ with dihalomethanes. These results seem to suggest that the bridging of the C4- and C6-hydroxyl groups in myo-inositol orthoesters is not feasible with an acetal moiety. The carbonate moiety in the triol $\mathbf{3 8 8}$ was quite labile and underwent hydrolysis in boiling water.


Scheme 3.2. a) TBSCI, 2,6-lutidine, DMF; b) $\mathrm{CH}_{2} \mathrm{Br}_{2}$ or $\mathrm{CH}_{2} \mathrm{I}_{2}, \mathrm{NaH}, \mathrm{DMF}$;
c) 1,1-O-carbonylimidazole, THF, $70^{\circ} \mathrm{C}$; d) TFA, $\mathrm{H}_{2} \mathrm{O}$ (4:1); e) TFA, $\mathrm{H}_{2} \mathrm{O}$ (8:1).

Schlewer et al has reported ${ }^{5}$ the synthesis of 4,6-cyclic phosphate $\mathbf{3 8 9}$ (an analog of myo-inositol 1,4,5-trisphosphate) from the triol $\mathbf{6 5}$ to investigate the biological activity of C4- and C6- bridged myo-inositol phosphates (Scheme 3.3).


Cyclization of the diallyl ether $\mathbf{3 9 0}^{6}$ (Scheme 3.4) by ring closing metathesis resulted in the formation of the corresponding cyclic ether derivative in moderate to good yields. However, cleavage of the orthoformate moiety in 391 resulted in inversion of the inositol ring back to the normal form and hence the tetrol $\mathbf{3 9 2}$ could not be obtained. Similar result was obtained for the polymerization of the styrene derivative 394. From this report it is clear that larger bridges between C4- and C6oxygen atoms, perhaps more than three carbon atoms, do not induce the required rigidity to keep the myo-inositol ring in the 'axial rich' (see below) conformation.




Scheme 3.4. a) 398, DCM; b) TsOH, MeOH, THF; c) AIBN, toluene, heat, $90 \%$.
Yamada et al reported ${ }^{7}$ the ring inversion of myo-inositol derivatives from the 'equatorial rich' conformation (e.g. 399) to the 'axial rich' conformation (e.g. 400) by introducing bulky silyl protecting groups in the vicinal trans hydroxyl groups (Scheme 3.5). Protection of the C5- and C6-hydroxyl groups in myo-inositol as the corresponding silyl ethers brought about a change in the conformation of the myoinositol ring (from the 'equatorial rich' conformation to the 'axial rich' conformation) depending upon the bulkiness of the silyl protecting group. Use of TBDPS protection on C5- and C6-hydroxyl groups inverted the myo-inositol ring to the 'axial rich' conformation irrespective of the substituents present on the other hydroxyl groups. However, introduction of bulky silyl protecting group at C1- and C6-hydroxyl groups (405-410) did not bring about inversion of the myo-inositol ring. But there appears to be no way of predicting as to which groups can bring about a change in conformation
of the inositol ring. Ring inversion of carbohydrate derivatives by introducing bulky silyl protecting groups in vicinal trans diols (e.g. 413, 414) has also been reported. ${ }^{8}$


Scheme 3.5. a) $\mathrm{R}_{3}$ SiOTf, 2,6-lutidine, DMF, $100^{\circ} \mathrm{C}$; b) $\mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}, \mathrm{THF}$.
Kaup et al ${ }^{9}$ obtained the phenyl boronate $\mathbf{4 1 5}$ by the reaction of myo-inositol with phenyl boronic acid in the solid state (Scheme 3.6). Although the inositol ring in the boronate $\mathbf{4 1 5}$ is flipped, it may not be possible to obtain the 1,3,5-triaxial triol or its derivatives starting from the boronate 415. Such boronate esters are known to be quite labile to the action of water. ${ }^{10}$


Scheme 3.6. a) $\mathrm{PhB}(\mathrm{OH})_{2}, 9{ }^{\circ} \mathrm{C}, 100 \%$.
The following section describes our attempts to make rigid myo-inositol derivatives with triaxially disposed hydroxyl groups by bridging the C4- and C6hydroxyl groups through a suitable linker.

### 3.2. Results and discussion

In order to prepare a derivative of myo-inositol orthoformate, bridged between the C4- and the C6-oxygen atoms, we attempted the reaction of myo-inositol orthoformate and its 2-O-protected derivatives with the reagents shown in Chart 3.2.



416


417


418


419


420


421

Chart 3.2
Reaction of the triol 65 with tere-phthaloyl chloride (416) using sodium hydride or triethylamine as a base resulted in a mixture of products (Scheme 3.7). No attempt was made to separate and identify the products since the TLC analysis of the mixture of products obtained suggested this to be impractical. It is likely that this reaction resulted in the formation of oligomers such as $\mathbf{4 2 2}$ consisting of phthaloyl and myo-inositol orthoformate units.



Scheme 3.7. a) NaH or $\mathrm{Et}_{3} \mathrm{~N}, \mathrm{DMF}$.
Reaction of the racemic dibenzoate $\mathbf{4 2 3}$ with dibromo-p-xylene 417 (whether leq or excess) in the presence of silver(I) oxide also resulted in a mixture of products, separation of which did not appear to be practical (Scheme 3.8). Earlier work ${ }^{11}$ in our laboratory had shown that reaction of the racemic dibenzoate $\mathbf{4 2 3}$ with alkyl halides yields the corresponding 4,6-diether (e.g. 424) in good yields. It is likely that this reaction resulted in the formation of oligomers such as $\mathbf{4 2 5}$ consisting of $p$-xylene and myo-inositol orthoformate units.



Scheme 3.8. a) $\mathrm{Ag}_{2} \mathrm{O}, \mathrm{DMF}, \mathrm{BnBr}$; b) $\mathrm{Ag}_{2} \mathrm{O}, \mathrm{DMF}, 417$.

Reaction of the diol 261 with 417 in the presence of potassium carbonate or sodium hydride resulted in a mixture of products (Scheme 3.9). Again, this could be because of the formation of oligomers 425 (Scheme 3.8). Previous work ${ }^{12}$ in our laboratory had shown that the reaction of the diol $\mathbf{2 6 1}$ with benzyl bromide gave the corresponding C4-ether 263 in good yield.


Scheme 3.9. a) $\mathrm{K}_{2} \mathrm{CO}_{3}$, DMF, BnBr ; b) $\mathrm{K}_{2} \mathrm{CO}_{3}$ or NaH , DMF, 417.
In all the experiments mentioned above, all the reactants were mixed in DMF and stirred at ambient temperature. But, when the reaction of the diol $\mathbf{2 6 1}$ with the dibromide 417 was carried out by the addition of sodium hydride (1.2 equivalents) to a solution of the diol 261 and the dibromide 417 in DMF, although a mixture of products resulted, the diether 426 could be isolated in $10 \%$ yield (Scheme 3.10). Increasing the amount of sodium hydride to 3.5 equivalents improved the yield of the diether $\mathbf{4 2 6}$ to $33 \%$. Yield of the diether $\mathbf{4 2 6}$ could further be improved to $69 \%$ by the sequential addition of sodium hydride (5 equivalents) and the diol $\mathbf{2 6 1}$ to a solution of the dibromide 417 in DMF. The presence of the two bromomethyl groups in the diether $\mathbf{4 2 6}$ was confirmed by converting the dibromide $\mathbf{4 2 6}$ to the corresponding dimethyl ether $\mathbf{4 2 8}$ (Scheme 3.10). Reaction of the dibromo dixylylether $\mathbf{4 2 6}$ with the benzoate diol 261 resulted in a mixture of products (Scheme 3.10). Similar result was obtained on using the dichloride 427 as well. The dichloro derivative 427 was obtained by the reaction of 261 with dichloro-p-xylene $\mathbf{4 1 8}$ under similar conditions used for the preparation of the dibromide 426 (Scheme 3.10). The reaction conditions for the cyclization reaction shown in scheme 3.10 (condition c) were used since
earlier work ${ }^{13}$ in our laboratory had shown this condition to be good for the preparation of myo-inositol orthoformate derived crown ethers like 430.




Scheme 3.10. a) NaH (1-5 eq), 417 or 418 (6 eq), DMF; b) NaOMe , MeOH, THF (reaction with 426); c) 261, $\mathrm{NaH}, \mathrm{THF}, 75^{\circ} \mathrm{C}$.

In order to rule out the possibility of hydrolysis of the C2-benzoate in the diol 261 and in the dixylyl ether 426 and the consequential formation of oligomers 429 and a mixture of products (Scheme 3.10), we also prepared the dixylylether 432 with tosylate at the C2-position (Scheme 3.11). Previous work ${ }^{14}$ in our laboratory had shown that the tosylate at the C2-position in myo-inositol orthoformate derivatives was stable to alkylation conditions in the presence of sodium hydride. However, the attempted cyclization reaction of $\mathbf{4 3 2}$ with the diol $\mathbf{4 3 1}$ also resulted in a mixture of products.


Scheme 3.11. a) $\mathrm{NaH}(5 \mathrm{eq}), 417$ ( 6 eq ), DMF; b) 431, $\mathrm{NaH}, \mathrm{THF}, 75^{\circ} \mathrm{C}$.
Since our attempts to obtain a C4-C6 bridged cyclic ether by the reaction of myo-inositol orthoformate or its derivatives with dihalo- $p$-xylenes failed, we investigated the crystal structure of the dibenzyl ether $\mathbf{1 0 2}$ which could give us information on the relative orientation of the two axial benzyl groups. The ORTEP plot for the dibenzyl ether $\mathbf{1 0 2}$ is shown in Figure 3.1.


Figure 3.1. ORTEP of $\mathbf{1 0 2}$

The three axial positions at C1, C3 and C5 constitute the orthoformate bridge. The conformation of the molecule as observed in the crystal shows three rather weak intramolecular interactions. The equatorial hydroxyl group O2-H2A makes bifurcated $\mathrm{OH} \ldots \mathrm{O}$ contacts with the orthoformate bridge atoms O 1 and O 3 . Another bifurcated contact of the $\mathrm{CH} . . . \mathrm{O}$ type is made by the $\mathrm{C} 2-\mathrm{H} 2$ group with ether atoms O 4 and O 6 (Table 3.4, page no. 196). An almost perpendicular orientation of the phenyl rings (the dihedral angle between the rings is $83.35^{\circ}$ ) facilitates the somewhat off-centred $\mathrm{C} 21-\mathrm{H} 21 \ldots \pi$ interaction $(\mathrm{H} 21 \ldots \mathrm{C} 9=2.85 \AA$ and $\mathrm{H} 21 \ldots \mathrm{C} 14=2.83 \AA)$. The distance between the para carbons (C12 and C19) of phenyl rings was found to be $5.3 \AA$. The distance between the two axial oxygen atoms O4 and O6 in the diol 261 was $2.71 \AA .{ }^{15}$ These interatomic distances and the orientation of the two axial benzyl groups do not appear to be conducive for the cyclization reaction between diols like 261 and dibenzyl ethers such as $\mathbf{4 2 6}$ (also see discussion on crystal structures, pages 175-178).

Reaction of the diol 261 with dibromo-o-xylene 419 in the presence of potassium carbonate gave a mixture of products. The same reaction when carried out using sodium hydride as the base gave the corresponding monoether 433 in reasonably good yield (Scheme 3.12). Attempted intramolecular cyclization of this bromide $\mathbf{4 3 3}$ however resulted in a mixture of products. This could again be because of the formation of oligomers such as 434. Similar results were obtained with the $m$ xylyl ether 435.


Mixture of










Scheme 3.12. a) $\mathrm{K}_{2} \mathrm{CO}_{3}$, DMF, 419; b) DMF, NaH (1 eq), 419 (1.2 eq) in DMF; c) DMF, NaH ; d) DMF, $\mathrm{NaH}(1 \mathrm{eq}), 420$ (1.2 eq) in DMF.

Reaction of the diol 437 with thionyl chloride gave the corresponding cyclic sulfite 438 (Scheme 3.13). Treatment of the cyclic sulfite with TFA to cleave the orthoformate resulted in complete cleavage of the sulfite moiety as well. Acetylation of the product of acid hydrolysis of the sulfite 438 gave the pentaacetate 439 . Oxidation ${ }^{16,17}$ of the sulfite 438 with ruthenium chloride / sodium periodate or ruthenium chloride / $\mathrm{OXONE}^{\circledR}$ gave a mixture of products. This was perhaps because
of the concomitant oxidation of the C2-benzyl group. Reports on the oxidation of myo-inositol orthoformate derivatives containing benzyl ethers giving rise to mixture of products have appeared earlier. ${ }^{18}$


Scheme 3.13. a) $\mathrm{SOCl}_{2}$, Pyr, DCM; b) (i) TFA / $\mathrm{H}_{2} \mathrm{O}$; (ii) $\mathrm{Ac}_{2} \mathrm{O} / \mathrm{Pyr}$; c) $\mathrm{RuCl}_{3} \cdot \mathrm{H}_{2} \mathrm{O}, \mathrm{OXONE}$ or $\mathrm{NaIO}_{4}$.

The cyclic sulfite 440 of the diol 261 could also be prepared in good yield and oxidized to the corresponding cyclic sulfate 441 (Scheme 3.14). This supports our view that the formation of a mixture of products during the oxidation of the sulfite 438 was due to the presence of the benzyl ether in 438 (Scheme 3.13). But the orthoformate moiety in the sulfate 441 was resistant to hydrolysis under reaction conditions normally used to cleave the orthoformate moiety of other myo-inositol orthoester derivatives (for details see experimental section 3.4). In all these reactions the cyclic sulfate 441 was recovered quantitatively.


Scheme 3.14. a) $\mathrm{SOCl}_{2}$, Pyr, DCM; b) $\mathrm{RuCl}_{3} . \mathrm{H}_{2} \mathrm{O}, \mathrm{NaIO}_{4}, \mathrm{CH}_{3} \mathrm{CN}, \mathrm{DCM}, \mathrm{H}_{2} \mathrm{O}(10: 2: 1)$; c) DIBAL-H, DCM; d) TFA, $\mathrm{H}_{2} \mathrm{O}$; e) TFA, $\mathrm{H}_{2} \mathrm{O}$, THF; f) TsOH, MeOH.

Attempted reduction of the orthoformate moiety in 441 with DIBAL-H resulted in exclusive cleavage of the C2-benzoate (Scheme 3.14) since benzoate
carbonyl group is more electrophilic than the orthoformate carbon. An interesting observation during the determination of the melting point of the cyclic sulfate 441 was that the crystals begin to jump to heights of several centimeters when heated to $208-215{ }^{\circ} \mathrm{C}$ (well before the melting point, $230-232{ }^{\circ} \mathrm{C}$ ). Further investigations are being carried out in our laboratory to understand this phenomenon. Earlier instances of 'jumping crystals' of inositol derivatives have been reported. ${ }^{19}$

We next attempted the preparation of the cyclic sulfite from myo-inositol orthobenzoate derived diol 443 (Scheme 3.15) since the orthobenzoate moiety is amenable to cleavage under hydrogenolysis conditions. The cyclic sulfite derivative 444 of the orthobenzoate 443 was prepared easily, but its hydrogenolysis resulted in cleavage of the cyclic sulfite moiety rather than the orthobenzoate to give the C2benzoate 443 . We also oxidized the cyclic sulfite 444 to the corresponding cyclic sulfate 445 but all our attempts to cleave the orthobenzoate moiety either under hydrogenolytic conditions or under acid hydrolytic conditions failed. Reduction of the cyclic sulfate 445 with DIBAL-H resulted in cleavage of the benzoate group as earlier (Scheme 3.14).


Scheme 3.15. a) $\mathrm{SOCl}_{2}$, Pyr, DCM; b) $\mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}$, EtOAc; c) $\mathrm{RuCl}_{3} . \mathrm{H}_{2} \mathrm{O}, \mathrm{NaIO}_{4}, \mathrm{CH}_{3} \mathrm{CN}$, DCM, $\mathrm{H}_{2} \mathrm{O}(10: 2: 1)$; d) DIBAL-H, DCM; e) $\mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}, \mathrm{MeOH}$; f) $\mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}$, EtOAc, $\left.\mathrm{MeOH} ; \mathrm{g}) \mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}, \mathrm{MeOH}, \mathrm{Et}_{3} \mathrm{~N} ; \mathrm{h}\right) \mathrm{Pd}(\mathrm{OH})_{2} / \mathrm{C}, \mathrm{H}_{2}, \mathrm{MeOH}, \mathrm{CH}_{3} \mathrm{COOH} ;$ i) TFA, $\mathrm{H}_{2} \mathrm{O}$; j) TFA, MeOH.

Subsequently, we converted the benzoate 445 to the corresponding benzyl ether 447 by aminolysis with isobutylamine followed by benzylation with sodium hydride and benzyl bromide (Scheme 3.16). The orthobenzoate moiety in the benzyl ether 447 resisted reduction with DIBAL-H.


Scheme 3.16. a) $i$-Butylamine, MeOH ; b) NaH , BnBr, DMF; c) DIBAL-H, DCM.

We solved the crystal structure of cyclic sulfite and cyclic sulfate derivatives of myo-inositol orthoesters (see experimental section for details, pages 199, 203, 206, $210,213,216,220)$ and compared them with the structure of cyclic carbonate ${ }^{1}$ hoping to find a clue for understanding the experimental observations described in this chapter. The relevant ORTEP diagrams are shown in the experimental section (pages $199,203,206,210,213,216,220$ ) and the relevant bond lengths, bond angles and inter atomic distances are given in Tables 3.1-3.2.

Table 3.1. Selected bond lengths, bond angles and interatomic distance for myoinositol orthoester derivatives.

|  | Bond length ( $\AA$ ) |  |  | Bond Angle ( ${ }^{\circ}$ ) |
| :---: | :---: | :---: | :---: | :---: |
|  | O4-S | O6-S | 04-06 | O4-S-06 |
| $438 \mathrm{R}=\mathrm{H}, \mathrm{R}^{2}=\mathrm{Bn}, \mathrm{X}=\mathrm{SO}$ | 1.611 | 1.620 | 2.464 | 99.84 |
| $440 \mathrm{R}=\mathrm{H}, \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}$ | 1.620 | 1.628 | 2.476 | 99.35 |
| $444 \mathrm{R}=\mathrm{Ph}, \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}$ | 1.632 | 1.619 | 2.467 | 98.74 |
| $441 \mathrm{R}=\mathrm{H}, \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}_{2}$ | 1.561 | 1.563 | 2.445 | 103.02 |
| $445 \mathrm{R}=\mathrm{Ph}, \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}_{2}$ | 1.572 | 1.567 | 2.445 | 102.33 |
| $447 \mathrm{R}=\mathrm{Ph}, \mathrm{R}^{2}=\mathrm{Bn}, \mathrm{X}=\mathrm{SO}_{2}$ | 1.562 | 1.560 | 2.437 | 102.64 |
| $442 \mathrm{R}=\mathrm{H}, \mathrm{R}^{2}=\mathrm{H}, \mathrm{X}=\mathrm{SO}_{2}$ | 1.562 | 1.564 | 2.439 | 102.57 |
| $387 \mathrm{R}=\mathrm{H}, \mathrm{R}^{2}=\mathrm{H}, \mathrm{X}=\mathrm{CO}$ | $1.343^{\text {a }}$ | $1.332^{\text {b }}$ | 2.312 | $119.67^{\text {c }}$ |
| $\mathbf{6 5}^{20}$ | -- | -- | 2.768 | -- |

A comparison of the bond lengths and bond angles (involved in the formation of O4-O6 bridge, Table 3.1) for myo-inositol orthoester derivatives (cyclic sulfite, cyclic sulfate and cyclic carbonate) show that the bond lengths O4-S and O6-S do not vary much but the bond angle of O4-S-O6 increases by a few degrees after oxidation of the sulfites to corresponding sulfates. More significantly, the distance between O4 and O6 progressively decreases on going from myo-inositol orthoformate (65) to its cyclic sulfite (or sulfate) to its carbonate derivatives. This implies an increase in the deformation of the carbocyclic ring and strain in these molecules on forming O4-O6 bridged derivatives. Hence it is likely that the formation of O4-O6 bridge with a
tetrahedral carbon atom (as in a cyclic acetal) could result in an increase in the strain to such an extent that formation of such a bridge might not be feasible and stable.

Table 3.2. Selected bond lengths and bond angles for myo-inositol orthoester derivatives.

|  | Bond lengths ( $\AA$ ) |  |  | Bond angles ( ${ }^{\circ}$ ) |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | C-O1 | C-03 | C-05 | 01-C-O3 | O3-C-05 | O1-C-05 |
| $\begin{aligned} & \text { 438, } \mathrm{R}=\mathrm{H}, \\ & \mathrm{R}^{2}=\mathrm{Bn}, \mathrm{X}=\mathrm{SO} \\ & \hline \end{aligned}$ | 1.403 | 1.401 | 1.406 | 112.42 | 110.58 | 110.55 |
| $\begin{aligned} & \text { 440, } \mathrm{R}=\mathrm{H}, \\ & \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO} \end{aligned}$ | 1.401 | 1.405 | 1.406 | 112.34 | 110.69 | 110.96 |
| $\begin{aligned} & \text { 444, } \mathrm{R}=\mathrm{Ph}, \\ & \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO} \end{aligned}$ | 1.408 | 1.414 | 1.412 | 111.22 | 109.13 | 109.90 |
| $\begin{aligned} & \text { 441, } \mathrm{R}=\mathrm{H}, \\ & \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}_{2} \end{aligned}$ | 1.395 | 1.393 | 1.412 | 112.56 | 110.32 | 110.53 |
| $\begin{aligned} & 445, \mathrm{R}=\mathrm{Ph}, \\ & \mathrm{R}^{2}=\mathrm{Bz}, \mathrm{X}=\mathrm{SO}_{2} \\ & \hline \end{aligned}$ | 1.407 | 1.407 | 1.406 | 111.60 | 109.46 | 109.33 |
| $\begin{aligned} & 447, \mathrm{R}=\mathrm{Ph}, \\ & \mathrm{R}^{2}=\mathrm{Bn}, \mathrm{X}=\mathrm{SO}_{2} \end{aligned}$ | 1.402 | 1.401 | 1.425 | 112.19 | 109.10 | 108.83 |
| $\begin{aligned} & \text { 442, } \mathrm{R}=\mathrm{H}, \\ & \mathrm{R}^{2}=\mathrm{H}, \mathrm{X}=\mathrm{SO}_{2} \end{aligned}$ | 1.393 | 1.396 | 1.402 | 112.25 | 111.08 | 110.46 |
| $65^{20}$ | 1.401 | 1.407 | 1.404 | 111.13 | 111.21 | 111.47 |

Table 3.3. Selected bond lengths and bond angles for myo-inositol derived ketals (values taken from refs 21, 22).

|  | Bond lengths ( $\AA$ ) |  |  |  | Bond angles ( ${ }^{\circ}$ ) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | C-O1 | C-O2 | C-O4 | C-05 | O1-C-O2 | 04-C-05 |
| 56, $\mathrm{R}=\left(\mathrm{CH}_{2}\right)_{5} \mathrm{C}$ | 1.439 | 1.425 | 1.446 | 1.441 | 105.40 | 105.66 |
| $\mathbf{6 0}, \mathrm{R}=\left(\mathrm{CH}_{3}\right)_{2} \mathrm{C}$ | 1.441 | 1.425 | 1.452 | 1.442 | 105.79 | 105.96 |

Similarly a comparison of the C-O bond lengths and the corresponding bond angles for myo-inositol orthoesters and their derivatives (Tables 3.1-3.2) with the
ketal derivatives (Table 3.3) show that C-O bond lengths decrease where as bond angles increase on going from ketals to orthoesters indicating an increase in strain. Hence the orthoester moiety being very rigid may not allow introduction of rings containing a tetrahedral carbon atom on the opposite face of the carbocyclic (inositol) ring which further increases the strain in the molecule. These reasons could be responsible for the non formation of $\mathrm{O} 4-\mathrm{O} 6$ bridged myo-inositol orthoester derivatives shown in Schemes 3.7-3.12.

### 3.3. Conclusions

We attempted to prepare O4-O6 bridged myo-inositol orthoester derivatives that could be potentially useful for the preparation of cyclitol derivatives and also serve as metal complexing agents. Although we were successful in preparing myoinositol orthoester derivatives containing O4-O6 bridge (with sulfur) we were unable to cleave the orthoester moiety to release the 1,3,5-cis triaxial hydroxyl groups. A comparison of the crystal structures of various myo-inositol orthoester derivatives suggests that bridging of O4 and O6 atoms (as ethers or ketals) may not be feasible on these trioxa adamantane derivatives.

### 3.4. Experimental section

General: General experimental procedures are same as in Chapter 2, Section A except for the following: Dry DCM was used as solvent in all the experiments. Palladium hydroxide on charcoal ( $20 \mathrm{wt} . \%$ ) was used for all the hydrogenolysis reactions. myo-Inositol derivatives $\mathbf{2 6 1},{ }^{23} \mathbf{4 2 3},{ }^{24} \mathbf{4 3 1},{ }^{25}$ and $\mathbf{4 3 7}{ }^{25}$ were prepared according to literature procedures. The diol $\mathbf{4 4 3}$ was obtained as a gift from Mr. C. Murali. ${ }^{26}$ Compounds $\mathbf{4 2 6}-428,432,433,435,439,442,446$ were not characterized rigorously since they were not used further in any experiment. ${ }^{1} \mathrm{H}$ NMR spectra reported for the compounds $\mathbf{4 2 6}-\mathbf{4 2 8}, 432,433$ and $\mathbf{4 3 5}$ are for crude products.

Reaction of 2-O-benzoyl myo-inositol $1,3,5$-orthoformate (261) with $\alpha, \alpha$ '-dibromo-p-xylene (417). Method A. To a solution of the diol 261 ( $0.147 \mathrm{~g}, 0.50$ $\mathrm{mmol})$ in DMF $(4 \mathrm{~mL})$ sodium hydride $(0.012 \mathrm{~g}, 0.50 \mathrm{mmol})$ was added and stirred for 15 min . $\alpha, \alpha$ '-Dibromo- $p$-xylene $417(0.066 \mathrm{~g}, 0.25 \mathrm{mmol})$ was then added and the reaction mixture was stirred for 30 min at ambient temperature. TLC analysis of the reaction mixture showed a mixture of products. No attempt was made to isolate the products.

Method B. To a solution of the diol $261(0.147 \mathrm{~g}, 0.50 \mathrm{mmol})$ and $\alpha, \alpha$ '-dibromo-pxylene $417(0.792 \mathrm{~g}, 3 \mathrm{mmol})$ in DMF ( 8 mL ) sodium hydride ( $0.024 \mathrm{~g}, 0.60 \mathrm{mmol}$ ) was added and stirred for 20 min . The reaction mixture was worked up by adding ethyl acetate saturated with water. The organic layer was washed with water followed by brine and dried over anhydrous sodium sulfate. The products were separated by column chromatography (100-200 mesh silica gel) using $15 \%$ ethyl acetate / petroleum ether as eluent to get the diether $\mathbf{4 2 6}(0.035 \mathrm{~g}, 10 \%)$.

## Data for 426:

mp. $109-110{ }^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.15(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.3 \mathrm{~Hz}), 7.40-7.66(\mathrm{~m}, 3 \mathrm{H}), 7.19-7.35(\mathrm{~m}, 8 \mathrm{H})$, 5.55-5.63 (m, 2H), 4.66 ( $\mathrm{s}, 4 \mathrm{H}$ ), 4.39-4.60 (m, 9H).

Elemental analysis calcd for $\mathrm{C}_{30} \mathrm{H}_{28} \mathrm{O}_{7} \mathrm{Br}_{2}$ : C, 54.56 ; H, 4.27. Found: C, 54.48 ; H, 4.39.

Method C. To a solution of the diol $261(0.147 \mathrm{~g}, 0.50 \mathrm{mmol})$ and $\alpha, \alpha$ 'dibromo-pxylene $417(0.792 \mathrm{~g}, 3 \mathrm{mmol})$ in DMF ( 10 mL ), sodium hydride ( $0.070 \mathrm{~g}, 1.75 \mathrm{mmol}$ ) was added and the mixture stirred at ambient temperature for 35 min . The reaction mixture was processed as above to obtain the diether $\mathbf{4 2 6}(0.110 \mathrm{~g}, 33 \%)$.

Method D. To a solution of $\alpha$, $\alpha$ '-dibromo-p-xylene 417 ( $2.100 \mathrm{~g}, 8 \mathrm{mmol}$ ) in DMF $(16 \mathrm{~mL})$, sodium hydride $(0.200 \mathrm{~g}, 5 \mathrm{mmol})$ and the diol $261(0.294 \mathrm{~g}, 1 \mathrm{mmol})$ were added sequentially and the mixture stirred vigorously for 40 min and worked up as above to obtain the diether $\mathbf{4 2 6}$ ( $0.456 \mathrm{~g}, 69 \%$ ).

Reaction of 2-O-benzoyl-4,6-di-O-(p-bromomethylbenzyl) myo-inositol 1,3,5orthoformate (426) with sodium methoxide. To a solution of the dibromodiether $426(0.100 \mathrm{~g}, 0.15 \mathrm{mmol})$ in THF ( 2 mL ) and methanol ( 3 mL ), sodium methoxide $(0.082 \mathrm{~g}, 1.52 \mathrm{mmol})$ was added and the mixture refluxed for 1.5 h . The reaction mixture was worked up with ethyl acetate to get the dimethylether $428(0.064 \mathrm{~g}$, $75 \%) .{ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{4 2 8}$ suggested the presence of a small amount $\mathbf{4 2 6}$ in the product.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.25(\mathrm{~s}, 8 \mathrm{H}), 5.45(\mathrm{~s}, 1 \mathrm{H}), 4.15-4.70(\mathrm{~m}, 15 \mathrm{H}), 3.38(\mathrm{~s}, 6 \mathrm{H}), 2.18$ (broad s, 1H).

Reaction of 2-O-benzoyl-myo-inositol 1,3,5-orthoformate (261) with $\alpha, \alpha$ '-dichloro-p-xylene (418). To a solution of $\alpha$, $\alpha^{\prime}$-dichloro- $p$-xylene 418 ( $0.462 \mathrm{~g}, 2.64$ $\mathrm{mmol})$ in DMF $(6 \mathrm{~mL})$ sodium hydride $(0.070 \mathrm{~g}, 1.76 \mathrm{mmol})$ was added immediately followed by the diol $261(0.130 \mathrm{~g}, 0.44 \mathrm{mmol})$ and the mixture stirred vigorously for

40 min at ambient temperature. Work up and purification as above gave the dichloro diether 427 ( $0.145 \mathrm{~g}, 58 \%$ ).

## Data for 427:

mp. $115-117^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.15(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7.3 \mathrm{~Hz}), 7.40-7.67(\mathrm{~m}, 3 \mathrm{H}), 7.28(\mathrm{~s}, 8 \mathrm{H}), 5.55-$ $5.68(\mathrm{~m}, 2 \mathrm{H}), 4.35-4.80(\mathrm{~m}, 13 \mathrm{H})$.

Elemental analysis calcd for $\mathrm{C}_{30} \mathrm{H}_{28} \mathrm{O}_{7} \mathrm{Cl}_{2}$ : C, $63.10 ; \mathrm{H}, 4.94$. Found: C, $63.60 ; \mathrm{H}$, 5.40.

Reaction of 2-O-tosyl-myo-inositol 1,3,5-orthoformate (431) with $\alpha$, $\alpha$ '-dibromo-$\boldsymbol{p}$-xylene (417). To a solution of $\alpha, \alpha$ '-dibromo- $p$-xylene 417 ( $0.650 \mathrm{~g}, 2.46 \mathrm{mmol}$ ) in DMF ( 6 mL ) sodium hydride ( $0.066 \mathrm{~g}, 1.64 \mathrm{mmol}$ ) was added immediately followed by the tosylate diol $431(0.140 \mathrm{~g}, 0.41 \mathrm{mmol})$ and the mixture stirred vigorously for 40 min at ambient temperature. Usual work up with ethyl acetate and column chromatography as in the method B (preparation of 426) gave the diether 432 (0.181 $\mathrm{g}, 62 \%$ ) as a light green gummy solid.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.76(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.8 \mathrm{~Hz}), 7.05-7.45(\mathrm{~m}, 10 \mathrm{H}), 5.52(\mathrm{~s}, 1 \mathrm{H}), 4.95-$ $5.05(\mathrm{~m}, 1 \mathrm{H}), 4.20-4.62(\mathrm{~m}, 13 \mathrm{H}), 2.35(\mathrm{~s}, 3 \mathrm{H})$.

Reaction of 2-O-benzoyl-myo-inositol 1,3,5-orthoformate (261) with $\alpha, \alpha$ '-dibromo-o-xylene (419). To a solution of the diol 261 ( $0.294 \mathrm{~g}, 1 \mathrm{mmol}$ ) in DMF (4 $\mathrm{mL})$ sodium hydride ( $0.044 \mathrm{~g}, 1.10 \mathrm{mmol}$ ) was added and stirred for 15 min at ambient temperature. A solution of $\mathbf{4 1 9}(0.290 \mathrm{~g}, 1.10 \mathrm{mmol})$ in DMF ( 2 mL ) was then added in one lot and the mixture stirred for 1 h at ambient temperature. Work up of the reaction mixture as above followed by column chromatography ( $10 \%$ ethyl acetate / petroleum ether) gave the crude monoether $433(0.167 \mathrm{~g}, 35 \%)$ as a gum.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.01-8.26(\mathrm{~m}, 2 \mathrm{H}), 7.20-7.70(\mathrm{~m}, 7 \mathrm{H}), 5.43-5.65(\mathrm{~m}, 2 \mathrm{H}), 4.79-$ $5.05(\mathrm{~m}, 2 \mathrm{H}), 4.35-4.75(\mathrm{~m}, 7 \mathrm{H}), 3.71(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=9.8 \mathrm{~Hz})$.

Reaction of 2-O-benzoyl-myo-inositol $1,3,5$-orthoformate (261) with $\alpha, \alpha$ '-dibromo-m-xylene (420). To a solution of the diol $261(0.294 \mathrm{~g}, 1 \mathrm{mmol})$ in DMF (4 $\mathrm{mL})$ sodium hydride $(0.044 \mathrm{~g}, 1.10 \mathrm{mmol})$ was added and the mixture stirred for 15 min at ambient temperature. A solution of $\mathbf{4 2 0}(0.264 \mathrm{~g}, 1 \mathrm{mmol})$ in DMF $(2 \mathrm{~mL})$ was added in one lot and stirred for 1 h . Work up and purification as above gave the monoether $435(0.237 \mathrm{~g}, 50 \%)$ as a gum.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.01(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8.8 \mathrm{~Hz}), 7.17-7.60(\mathrm{~m}, 7 \mathrm{H}), 5.48(\mathrm{~s}, 1 \mathrm{H}), 5.43-$ $5.47(\mathrm{~m}, 1 \mathrm{H}), 4.68(\mathrm{AB} \mathrm{q}, 2 \mathrm{H}, \mathrm{J}=11.8 \mathrm{~Hz}), 4.47-4.55(\mathrm{~m}, 2 \mathrm{H}), 4.44(\mathrm{~s}, 2 \mathrm{H}), 4.35-$ $4.42(\mathrm{~m}, 2 \mathrm{H}), 4.26-4.34(\mathrm{~m}, 1 \mathrm{H}), 3.74(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=9.7 \mathrm{~Hz})$.

Preparation of myo-inositol derived cyclic sulfites. General procedure. The required myo-inositol orthoester derivative and dry pyridine were taken in dry DCM and cooled to $0^{\circ} \mathrm{C}$. Thionyl chloride (421) was added drop-wise using a syringe at 0 ${ }^{\circ} \mathrm{C}$ over a period of 2-3 min. The reaction mixture was stirred for 3-4 h , diluted with DCM and washed with sodium bicarbonate solution followed by water and brine. The organic layer was dried over anhydrous sodium sulfate. Evaporation of the solvent under reduced pressure followed by column chromatography (using 10-20\% ethyl acetate - light petroleum as the eluent) gave the cyclic sulfite.

2-O-benzyl-myo-inositol $\mathbf{1 , 3 , 5}$-orthoformate-4,6-cyclic sulfite (438). The cyclic sulfite $\mathbf{4 3 8}$ was prepared as above using the 2-benzyl ether $437(0.500 \mathrm{~g}, 1.80 \mathrm{mmol})$, pyridine $(2.20 \mathrm{~mL})$, $\mathrm{DCM}(6 \mathrm{~mL})$ and thionyl chloride $(0.47 \mathrm{~mL}, 5.40 \mathrm{mmol})$. The cyclic sulfite $\mathbf{4 3 8}$ was obtained as a colorless solid ( $0.380 \mathrm{~g}, 65 \%$ ).

## Data for 438:

mp. $129-131^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.30-7.45(\mathrm{~m}, 5 \mathrm{H}), 5.72-5.80(\mathrm{~m}, 1 \mathrm{H}), 5.54-5.58(\mathrm{~m}, 1 \mathrm{H}), 5.11-$ $5.20(\mathrm{~m}, 2 \mathrm{H}), 4.76(\mathrm{~s}, 2 \mathrm{H}), 4.37-4.45(\mathrm{~m}, 2 \mathrm{H}), 3.91-3.97(\mathrm{~m}, 1 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 137.2,128.6,128.2,127.9,102.4,71.9,69.0,68.7,66.8,59.7$.
Elemental analysis Calcd for $\mathrm{C}_{14} \mathrm{H}_{14} \mathrm{O}_{7} \mathrm{~S}$ : C, 51.53 ; H, 4.32. Found: C, 51.87 ; H, 4.19.

2-O-benzoyl-myo-inositol 1,3,5-orthoformate-4,6-cyclic sulfite (440). The cyclic sulfite 440 was prepared as above using the diol $261(0.600 \mathrm{~g}, 2.04 \mathrm{mmol})$, pyridine $(2.5 \mathrm{~mL}), \mathrm{DCM}(12 \mathrm{~mL})$ and thionyl chloride $(0.53 \mathrm{~mL}, 6.12 \mathrm{mmol})$. The cyclic sulfite 440 was obtained as a colorless solid ( $0.403 \mathrm{~g}, 58 \%$ ).

Data for 440:
mp. $178-180^{\circ} \mathrm{C}$.
IR (v, nujol): $1720 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.10-8.25(\mathrm{~m}, 2 \mathrm{H}), 7.40-7.70(\mathrm{~m}, 3 \mathrm{H}), 5.80-5.92(\mathrm{~m}, 1 \mathrm{H}), 5.60$ $(\mathrm{d}, 1 \mathrm{H}, \mathrm{J}=1.3 \mathrm{~Hz}), 5.45-5.55(\mathrm{~m}, 1 \mathrm{H}), 5.23(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=4 \mathrm{~Hz}), 4.55-4.65(\mathrm{~m}, 2 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 165.7,133.7,130.0,129.1,128.5,102.3,68.6,63.2,59.7$.
Elemental analysis Calcd for $\mathrm{C}_{14} \mathrm{H}_{12} \mathrm{O}_{8} \mathrm{~S}$ : C, 49 41; H, 3.55. Found: C, 49.32; H, 3.25.

2-O-benzoyl-myo-inositol $\mathbf{1 , 3 , 5 - o r t h o b e n z o a t e - 4 , 6 - c y c l i c ~ s u l f i t e ~ ( 4 4 4 ) . ~ T h e ~ c y c l i c ~}$ sulfite 444 was prepared as above using the diol $443(0.400 \mathrm{~g}, 1.08 \mathrm{mmol})$, pyridine $(1.3 \mathrm{~mL}), \mathrm{DCM}(6 \mathrm{~mL})$ and thionyl chloride $(0.28 \mathrm{~mL}, 3.24 \mathrm{mmol})$. The cyclic sulfite 444 was obtained as a colorless solid ( $0.337 \mathrm{~g}, 75 \%$ ).

## Data for 444:

mp. $162-164{ }^{\circ} \mathrm{C}$.
IR ( $v$, nujol): $1711 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.16(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=8 \mathrm{~Hz}), 7.30-7.75(\mathrm{~m}, 8 \mathrm{H}), 5.98-6.08(\mathrm{~m}, 1 \mathrm{H})$, $5.59(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=2 \mathrm{~Hz}), 5.32(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=4 \mathrm{~Hz}), 4.73-4.83(\mathrm{~m}, 2 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 165.7,135.8,133.6,130.0,129.9,129.3,128.6,128.2,125.3$, 107.3, 70.2, 68.5, 62.6, 60.9.

Elemental analysis calcd for $\mathrm{C}_{20} \mathrm{H}_{16} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 57.69 ; \mathrm{H}, 3.87$. Found: C, $57.57 ; \mathrm{H}, 3.83$.
Hydrolysis of 2-O-benzyl-myo-inositol 1,3,5-orthoformate-4,6-cyclic sulfite (438) with aqueous TFA. The cyclic sulfite $438(0.017 \mathrm{~g}, 0.05 \mathrm{mmol})$ was stirred in distilled water $(0.2 \mathrm{~mL})$ and trifluoroacetic acid $(0.8 \mathrm{~mL})$ for 12 h . The solid obtained on evaporation of the solvent under reduced pressure was acetylated with acetic anhydride $(0.5 \mathrm{~mL})$ in pyridine ( 1 mL ) overnight at room temperature, which gave the pentaacetate 439 ( $0.026 \mathrm{~g}, 100 \%$ ).

## Data for 439:

mp. 217-219 ${ }^{\circ} \mathrm{C}$.
IR ( $v$, nujol): $1744 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H} \mathbf{N M R}\left(\mathrm{CDCl}_{3}\right): \delta 7.32(\mathrm{~s}, 5 \mathrm{H}), 5.61(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=10.1 \mathrm{~Hz}), 5.12(\mathrm{t}, 1 \mathrm{H}, \mathrm{J}=9.4 \mathrm{~Hz})$, $4.95(\mathrm{dd}, 2 \mathrm{H}, \mathrm{J}=2.4,10.6 \mathrm{~Hz}), 4.67(\mathrm{~s}, 2 \mathrm{H}), 4.07-4.14(\mathrm{~m}, 1 \mathrm{H}), 1.98(\mathrm{~s}, 9 \mathrm{H}), 1.96(\mathrm{~s}$, 6 H ).

Hydrogenolysis of 2-O-benzoyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfite (444). A solution of the orthobenzoate $444(0.100 \mathrm{~g}, 0.24 \mathrm{mmol})$ in ethyl acetate ( 1.5 mL ) was hydrogenolyzed in the presence of palladium hydroxide on charcoal ( 0.100 g) at 65 psi for 7 h . The catalyst was removed by filtration through a small bed of Celite and evaporation of the filtrate gave the diol $443(0.084 \mathrm{~g}, 95 \%)$.
mp. $183-185^{\circ} \mathrm{C}\left(\right.$ Lit. $\left.{ }^{26} \mathrm{mp} .185-187^{\circ} \mathrm{C}\right)$.

Preparation of myo-inositol cyclic sulfates. General procedure. The required myoinositol orthoester cyclic sulfite was taken in a mixture of acetonitrile, dichloromethane and water ( $10: 2: 1 \mathrm{v} / \mathrm{v} 18-25 \mathrm{~mL}$ ). Sodium periodate (4 eq) and ruthenium chloride monohydrate ( $7 \mathrm{~mol} \%$ ) were added and the mixture stirred for 2-3 h at room temperature. The reaction mixture was filtered through a small bed of Celite using DCM as solvent. Evaporation of the filtrate under reduced pressure gave the cyclic sulfate

2-O-benzoyl-myo-inositol 1,3,5-orthoformate-4,6-cyclic sulfate (441). The cyclic sulfite $440(0.520 \mathrm{~g}, 1.53 \mathrm{mmol})$ was oxidized using sodium periodate $(1.300 \mathrm{~g}, 6.12$ $\mathrm{mmol})$ and ruthenium chloride monohydrate $(0.023 \mathrm{~g}, 0.11 \mathrm{mmol})$ as in the general procedure to get cyclic sulfate 441 as a colorless solid ( $0.524 \mathrm{~g}, 96 \%$ ).

## Data for 441:

mp. $230-232{ }^{\circ} \mathrm{C}$.
IR ( $v$, nujol): $1726 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.12-8.20(\mathrm{~m}, 2 \mathrm{H}), 7.58-7.69(\mathrm{~m}, 1 \mathrm{H}), 7.44-7.55(\mathrm{~m}, 2 \mathrm{H}), 5.55-$ $5.70(\mathrm{~m}, 3 \mathrm{H}), 5.30-5.45(\mathrm{~m}, 2 \mathrm{H}), 4.70-4.85(\mathrm{~m}, 2 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR (125.8 MHz, $\mathrm{CDCl}_{3}$ ): $\delta 165.4,133.8,130.0,128.9,128.6,101.8,73.8$, 68.6, 62.4, 59.1.

Elemental analysis Calcd for $\mathrm{C}_{14} \mathrm{H}_{12} \mathrm{O}_{9} \mathrm{~S}: \mathrm{C}, 47.19$; H, 3.40. Found: C, 47.06; H, 3.72.

2-O-benzoyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfate (445). The cyclic sulfite $444(0.850 \mathrm{~g}, 2.04 \mathrm{mmol})$ was oxidized using sodium periodate $(1.750 \mathrm{~g}, 8.16$ $\mathrm{mmol})$ and ruthenium chloride monohydrate $(0.030 \mathrm{~g}, 0.14 \mathrm{mmol})$ as in the general procedure to get the cyclic sulfate $445(0.865 \mathrm{~g}, 98 \%)$.

## Data for 445:

mp. $80-82^{\circ} \mathrm{C}$.
IR ( $\mathrm{v}, \mathrm{CHCl}_{3}$ ): $1728 \mathrm{~cm}^{-1}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 8.16(\mathrm{~d}, 2 \mathrm{H}, \mathrm{J}=7 \mathrm{~Hz}), 7.57-7.71(\mathrm{~m}, 3 \mathrm{H}), 7.34-7.55(\mathrm{~m}, 5 \mathrm{H})$, 5.70-5.82 (m, 2H), 5.43-5.54 (m, 2H), 4.92-5.02 (m, 2H).
${ }^{13} \mathbf{C}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 165.4,134.7,133.8,130.4,130.0,128.9,128.6,128.3,125.3$, 107.0, 73.8, 70.0, 61.8, 60.3.

Elemental analysis calcd for $\mathrm{C}_{20} \mathrm{H}_{16} \mathrm{O}_{9} \mathrm{~S}: \mathrm{C}, 55.55 ; \mathrm{H}, 3.73$. Found: C, 55.85; H, 3.66. Attempted hydrolysis of 2-O-benzoyl-myo-inositol 1,3,5-orthoformate-4,6-cyclic sulfate (441). The orthoformate 441 was stirred with reagents shown below for 20-24 h. There was no reaction and the starting material was recovered after evaporation of the solvent.
a) Cyclic sulfate $441(0.200 \mathrm{~g}, 0.56 \mathrm{mmol}$ or $0.100 \mathrm{~g}, 0.28 \mathrm{mmol})$, distilled water $(0.8 \mathrm{~mL}$ or 0.2 mL$)$ and trifluoroacetic acid ( 1.2 mL or 0.8 mL ) at room temperature.
b) Cyclic sulfate $441(0.100 \mathrm{~g}, 0.28 \mathrm{mmol})$, THF $(0.4 \mathrm{~mL})$, distilled water ( 0.2 $\mathrm{mL})$ and trifluoroacetic acid $(0.4 \mathrm{~mL})$ at room temperature.
c) as in (b) at $60^{\circ} \mathrm{C}$.
d) Cyclic sulfate $441(0.100 \mathrm{~g}, 0.28 \mathrm{mmol})$ and $\mathrm{TsOH}(0.004 \mathrm{~g}, 0.02 \mathrm{mmol})$ in methanol $(1 \mathrm{~mL})$ at room temperature.

Reduction of 2-O-benzoyl-myo-inositol 1,3,5-orthoformate-4,6-cyclic sulfate (441) with DIBAL-H. A solution of the cyclic sulfate $441(0.200 \mathrm{~g}, 0.56 \mathrm{mmol})$ in DCM ( 5 mL ) was cooled to $0{ }^{\circ} \mathrm{C}$ and a 1 M solution of DIBAL-H in toluene ( $5.62 \mathrm{~mL}, 5.62$ mmol ) was added and stirred for 12 h at room temperature. The reaction mixture was worked up ${ }^{27}$ by stirring with a solution of sodium potassium tartrate and saturated
ammonium chloride solution and extracted with DCM. The organic layer was washed with water, dried over anhydrous sodium sulfate and the solvent evaporated under reduced pressure to get the product $442(0.098 \mathrm{~g}, 70 \%)$.

## Data for 442:

mp. $179-181^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 5.57(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=1.2 \mathrm{~Hz}), 5.50-5.55(\mathrm{~m}, 1 \mathrm{H}), 5.29-5.40(\mathrm{~m}, 2 \mathrm{H})$, 4.46-4.55 (m, 2H), 4.32-4.45 (m, 1H), $3.22(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=12 \mathrm{~Hz})$.

Attempted hydrogenolysis of 2-O-benzoyl-myo-inositol 1,3,5-orthobenzoate-4,6cyclic sulfate (445). A solution of the orthobenzoate 445 in methanol or ethyl acetate was hydrogenolyzed (various conditions as below) in the presence of palladium hydroxide on charcoal at 60 psi for 30 h . There was no reaction and the starting material was recovered by filtering through a small bed of Celite and evaporation of the solvent from the filtrate.
a) Cyclic sulfate $445(0.250 \mathrm{~g}, 0.578 \mathrm{mmol})$ and palladium hydroxide on charcoal ( 0.250 g ) in methanol ( 2 mL ).
b) Cyclic sulfate $445(0.200 \mathrm{~g}, 0.46 \mathrm{mmol})$ and palladium hydroxide on charcoal $(0.200 \mathrm{~g})$ in ethyl acetate $(2 \mathrm{~mL})$.
c) Cyclic sulfate $445(0.250 \mathrm{~g}, 0.58 \mathrm{mmol})$ and palladium hydroxide on charcoal $(0.250 \mathrm{~g})$ in ethyl acetate ( 1 mL ), methanol ( 1 mL ).
d) Cyclic sulfate $445(0.200 \mathrm{~g}, 0.46 \mathrm{mmol})$, triethylamine ( 0.04 mL ) and palladium hydroxide on charcoal $(0.200 \mathrm{~g})$ in methanol $(2 \mathrm{~mL})$.
e) Cyclic sulfate $445(0.200 \mathrm{~g}, 0.46 \mathrm{mmol})$, acetic acid $(0.04 \mathrm{~mL})$ and palladium hydroxide on charcoal $(0.200 \mathrm{~g})$ in methanol ( 2 mL ).

Attempted solvolysis of 2-O-benzoyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfate (445) with aqueous TFA. The orthobenzoate $445(0.100 \mathrm{~g}, 0.23 \mathrm{mmol})$ was stirred in distilled water $(0.2 \mathrm{~mL})$ and trifluoroacetic acid $(0.8 \mathrm{~mL})$ for 20 h . There was no reaction; the starting material was recovered after evaporation of the solvent. Similar result was obtained on using methanol ( 0.2 mL or 0.8 mL ) and trifluoroacetic acid $(0.8 \mathrm{~mL}$ or 0.2 mL$)$.

Reduction of 2-O-benzoyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfate (445) with DIBAL-H. To a solution of the orthobenzoate $445(0.250 \mathrm{~g}, 0.58 \mathrm{mmol})$ in DCM ( 6 mL ) cooled to $0{ }^{\circ} \mathrm{C}$, a solution of DIBAL-H in toluene $(5.8 \mathrm{~mL}, 5.78$ mmol ) was added and stirred for 12 h at room temperature. The reaction mixture was worked up ${ }^{27}$ by stirring with a solution of sodium potassium tartrate and saturated ammonium chloride solution and extracted with DCM. The organic layer was washed with water, dried over anhydrous sodium sulfate and the solvent evaporated under reduced pressure to get the product 446 ( $0.173 \mathrm{~g}, 91 \%$ ).
${ }^{1} \mathbf{H}$ NMR $\left(\mathrm{CDCl}_{3}\right): \delta 7.55-7.67(\mathrm{~m}, 2 \mathrm{H}), 7.34-7.51(\mathrm{~m}, 3 \mathrm{H}), 5.59-5.70(\mathrm{~m}, 1 \mathrm{H}), 5.44$ $(\mathrm{t}, 2 \mathrm{H}, \mathrm{J}=4.5 \mathrm{~Hz}), 4.62-4.74(\mathrm{~m}, 2 \mathrm{H}), 4.35-4.51(\mathrm{~m}, 1 \mathrm{H}), 3.30(\mathrm{~d}, 1 \mathrm{H}, \mathrm{J}=11.4 \mathrm{~Hz}$, $\mathrm{D}_{2} \mathrm{O}$ exchangeable).

## Preparation of 2-O-benzyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfate

 (447). A solution of the benzoate $445(0.600 \mathrm{~g}, 1.40 \mathrm{mmol})$ and iso-butylamine (1.4 $\mathrm{mL})$ in methanol ( 5.6 mL ) was refluxed for 6 h . The solvents were evaporated and the residue was chromatographed to obtain the sulfate $446(0.446 \mathrm{~g})$. A portion of the crude sulfate $446(0.400 \mathrm{~g}, 1.22 \mathrm{mmol})$ and benzyl bromide $(0.626 \mathrm{~g}, 0.44 \mathrm{~mL}, 3.66$ mmol ) were dissolved in DMF ( 5 mL ), sodium hydride ( $0.976 \mathrm{~g}, 2.44 \mathrm{mmol}$ ) was added and the mixture stirred for 1 h at room temperature. The reaction mixture was worked up as usual with ethyl acetate and the product 447 ( $0.490 \mathrm{~g}, 96 \%$ ) wasseparated by column chromatography using $10 \%$ ethyl acetate - petroleum ether as eluent.

## Data for 447:

mp. $151-153{ }^{\circ} \mathrm{C}$.
${ }^{1} \mathbf{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.57-7.69(\mathrm{~m}, 2 \mathrm{H}), 7.31-7.48(\mathrm{~m}, 8 \mathrm{H}), 5.59-5.68(\mathrm{~m}$, $1 \mathrm{H}), 5.34-5.42(\mathrm{~m}, 2 \mathrm{H}), 4.80(\mathrm{~s}, 2 \mathrm{H}), 4.68-4.76(\mathrm{~m}, 2 \mathrm{H}), 4.18-4.23(\mathrm{~m}, 1 \mathrm{H})$.
${ }^{13} \mathbf{C}$ NMR ( $100 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 137.0,134.9,130.1,128.6,128.2,128.1,127.9$, $125.4,106.9,74.2,72.0,70.1,65.4,60.3$.

Elemental analysis calcd for $\mathrm{C}_{20} \mathrm{H}_{18} \mathrm{O}_{8} \mathrm{~S}: \mathrm{C}, 57.41 ; \mathrm{H}, 4.34$. Found: C, 57.23; H, 4.28.
Reduction of 2-O-benzyl-myo-inositol 1,3,5-orthobenzoate-4,6-cyclic sulfate (447) with DIBAL-H. A solution of the orthobenzoate $447(0.200 \mathrm{~g}, 0.48 \mathrm{mmol})$ in DCM $(6 \mathrm{~mL})$ was cooled to $0{ }^{\circ} \mathrm{C}$ and a solution of DIBAL-H $(4.8 \mathrm{~mL}, 4.80 \mathrm{mmol})$ in toluene was added and the mixture was stirred for 12 h at room temperature. The reaction mixture was then worked $u^{27}$ by stirring with a solution of sodium potassium tartrate and saturated ammonium chloride solution and extracted with DCM. The organic layer was washed with water, dried over anhydrous sodium sulfate and the solvent evaporated under reduced pressure to get the starting material ( $0.187 \mathrm{~g}, 94 \%$ ).

### 3.5. References

1. Angyal, S. J. Carbohydr. Res. 2000, 325, 313.
2. Paquette, L. A.; Tae, J.; Gallucci, J. C. Org. Lett. 2000, 2, 143.
3. Angyal, S. J.; Odier, L.; Tate, M. E. Carbohydr. Res. 1995, 266, 143.
4. Tse, B.; Kishi, Y. J. Am. Chem. Soc. 1993, 115, 7892.
5. Ballereau, S.; Poirier, S. N.; Guillemette, G.; Spiess, B.; Schlewer, G. J. Chem. Soc., Perkin Trans. 1 1998, 1859.
6. Kim, T-H.; Giles, M.; Holmes, A. B. Chem. Commun. 2000, 2421.
7. (a) Yamada, H.; Okajima, K.; Imagawa, H.; Mukae, T.; Kawamura, Y.; Nishizawa, M. Tetrahedron Lett. 2004, 45, 3157; (b) Yamada, H.; Okajima, K.; Imagawa, H.; Nagata, Y.; Nishizawa, M. Tetrahedron Lett. 2004, 45, 4349; (c) Okajima, K.; Mukae, T.; Imagawa, H.; Kawamura, Y.; Nishizawa, M.; Yamada, H. Tetrahedron 2005, 61, 3497.
8. (a) Hosoya, T.; Ohashi, Y.; Matsumoto, T.; Suzuki, K. Tetrahedron Lett. 1996, 37, 663; (b) Yamada, H.; Tanigakiuchi, K.; Nagao, K.; Okajima, K.; Mukae, T. Tetrahedron Lett. 2004, 45, 5615.
9. Kaupp, G.; Naimi-Jamal, M. R.; Stepanenko, V. Chem. Eur. J. 2003, 9, 4156.
10. Frechet, J. M. J.; Nuyens, L. J.; Seymour, E. J. Am. Chem. Soc. 1979, 101, 432.
11. Das, T.; Shashidhar, M. S. Carbohydr. Res. 1997, 297, 243.
12. Das, T.; Praveen, T.; Shashidhar, M. S. Carbohydr. Res. 1998, 313, 55.
13. Sureshan, K. M.; Shashidhar, M. S.; Varma, A. J. J. Org. Chem. 2002, 67, 6884.
14. Sarmah, M. P.; Shashidhar, M. S.; Sureshan, K. M.; Gonnade, R. G.; Bhadbhade, M. M. Tetrahedron 2005, 61, 4437.
15. Samanta, U.; Puranik, V. G.; Chakrabarti, P.; Thoniyot, P.; Shashidhar, M. S. Acta. Cryst. Sec C. 1998, 54, 1289.
16. Gao, Y.; Sharpless, K. B. J. Am. Chem. Soc. 1988, 110, 7538.
17. (a) Schröder, M.; Griffith, W. P. J. Chem. Soc., Chem. Commun. 1979, 58; (b) Ko, H.; Kim, E.; Park, J. E.; Kim, D.; Kim, S. J. Org. Chem. 2004, 69, 112.
18. (a) Lee, H. W.; Kishi, Y. J. Org. Chem. 1985, 50, 4402; (b) Sarmah, M. P.; Shashidhar, M. S. Carbohydr. Res. 2003, 338, 999.
19. Steiner, T.; Hinrichs, W.; Saenger, W.; Gigg, R. Acta. Cryst. Sec B. 1993, 49, 708.
20. Uhlmann, P.; Vasella, A. Helv. Chim. Acta. 1992, 75, 1979.
21. Spiers, I. D.; Schwalbe, C. H.; Blake, A. J.; Solomons, K. R. H.; Freeman, S. Carbohydr. Res. 1997, 302, 43.
22. Chung, S-K .; Ryu, Y.; Chang, Y-T.; Whang, D.; Kim, K. Carbohydr. Res. 1994, 253, 13.
23. Ozaki, S.; Koga, Y.; Ling, L.; Watanabe, Y.; Kimura, Y.; Hirata, M. Bull. Chem. Soc. Jpn. 1994, 67, 1058.
24. Banerjee, T.; Shashidhar M. S. Tetrahedron Lett. 1994, 35, 8053.
25. Sureshan, K. M.; Shashidhar, M. S. Tetrahedron Lett. 2001, 42, 3037.
26. Murali, C.; Shashidhar, M. S. unpublished results.
27. Painter, G. F.; Grove, S. J. A.; Gilbert, I. H.; Holmes, A. B.; Raithby, P. R.; Hill, M. L.; Hawkins, P. T.; Stephens, L. R. J. Chem. Soc., Perkin Trans. 1 1999, 923.






Crystal Data Table for $\mathbf{1 0 2}$

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Triclinic, P-1 |
| Unit cell dimensions | $\mathrm{a}=9.0511(11) \AA$ alpha $=68.849(2)^{\circ}$ <br> $\mathrm{b}=10.0953(12) \AA \quad \mathrm{beta}=81.453(2)^{\circ}$ <br> $\mathrm{c}=10.6586(13) \AA$ gamma $=87.433(2)^{\circ}$ |
| Volume | $898.18(19) \AA^{3}$ |
| Z, Calculated density | $2,1.370 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.100 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.29 \times 0.22 \times 0.13 \mathrm{~mm}$ |
| Theta range for data collection | 2.07 to 25.50 deg. |
| Reflections collected / unique | $13398 / 3328[\mathrm{R}($ int $)=0.0252]$ |
| Completeness to theta $=25.00$ | $99.6 \%$ |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.086 |
| Final R indices [I>2sigma(I)] | $\mathrm{R} 1=0.0448, \mathrm{wR} 2=0.1081$ |
| R indices (all data) | $\mathrm{R} 1=0.0518, \mathrm{wR} 2=0.1126$ |
| Largest diff. peak and hole | 0.185 and $-0.184 \mathrm{e} . \AA^{-3}$ |

Table 3.4. Cg 1 is the centroid of the $\mathrm{C} 16-\mathrm{C} 21$ ring

| $D-\mathrm{H} \cdots A$ | $D-\mathrm{H}$ | $\mathrm{H} \cdots \mathrm{A}$ | $D \cdots \mathrm{~A}$ | $D-\mathrm{H} \cdots \mathrm{A}$ |
| :---: | :---: | :---: | :---: | :---: |
| $\mathrm{C} 2-\mathrm{H} 2 \cdots \mathrm{O} 6$ | 0.98 | 2.47 | $2.8550(19)$ | 103 |
| $\mathrm{C} 2-\mathrm{H} 2 \cdots \mathrm{O} 4$ | 0.98 | 2.69 | $3.0463(18)$ | 102 |
| $\mathrm{O} 2-\mathrm{H} 2 A \cdots \mathrm{O} 3$ | 0.82 | 2.63 | $2.9385(18)$ | 104 |
| $\mathrm{O} 2-\mathrm{H} 2 A \cdots \mathrm{O} 1$ | 0.82 | 2.66 | $2.9408(18)$ | 102 |
| $\mathrm{C} 10-\mathrm{H} 10 \cdots \mathrm{O}^{\mathrm{iii}}$ | 0.93 | 2.67 | $3.517(2)$ | 152 |
| $\mathrm{C} 15-$ <br> $\mathrm{H} 15 B \cdots \mathrm{O}^{\mathrm{ii}}$ | 0.97 | 2.64 | $3.514(2)$ | 150 |
| $\mathrm{C} 4-\mathrm{H} 4 \cdots \mathrm{O}^{\mathrm{ii}}$ | 0.98 | 2.62 | $3.466(2)$ | 145 |
| ${\mathrm{O} 2-\mathrm{H} 2 A \cdots \mathrm{O}^{\mathrm{i}}}^{\mathrm{i}}$ | 0.82 | 2.15 | $2.8558(16)$ | 144 |
| $\mathrm{C} 15-$ <br> $\mathrm{H} 15 A \cdots \mathrm{Cg}^{\text {iv }}$ | 0.97 | 2.69 | 3.602 | 158 |

Symmetry codes: (i) $-\mathrm{x},-\mathrm{y}+2,-\mathrm{z}+1$; (ii) $-\mathrm{x}+1,-\mathrm{y}+2,-\mathrm{z}+1$; (iii) $-\mathrm{x}+1,-\mathrm{y}+1,-\mathrm{z}+2$; (iv) $-\mathrm{x}+1,-\mathrm{y},-\mathrm{z}$.





ORTEP of $\mathbf{4 3 8}$
Crystal Data Table for $\mathbf{4 3 8}$

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Monoclinic, $\mathrm{P} 2{ }_{1} / \mathrm{n}$ |
| Unit cell dimensions | $\mathrm{a}=8.549(2) \AA \quad$ alpha $=90^{\circ}$. <br> $\mathrm{b}=11.112(3) \AA \quad$ beta $=99.109(7)^{\circ}$. <br> $\mathrm{c}=14.775(4) \AA \quad$ gamma $=90^{\circ}$ |
| Volume | $1386.0(7) \AA^{3}$ |
| Z, Calculated density | $4,1.564 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.268 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.28 \times 0.26 \times 0.21 \mathrm{~mm}$ |
| Theta range for data collection | 2.30 to 25.00 deg. |
| Reflections collected $/$ unique | $6625 / 2432[\mathrm{R}($ int $)=0.0254]$ |
| Completeness to theta $=25.00$ | $99.8 \%$ |
| Refinement method | $\mathrm{Full-matrix} \mathrm{least-squares} \mathrm{on} \mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.044 |
| Final R indices [I $>2$ sigma(I) $]$ | $\mathrm{R} 1=0.0342, \mathrm{wR} 2=0.0888$ |
| R indices (all data) | $\mathrm{R} 1=0.0404, \mathrm{wR2}=0.0933$ |
| Largest diff. peak and hole | 0.244 and $-0.279 \mathrm{e} . \AA^{-3}$ |









ORTEP of 440
Crystal Data Table for 440

| Wavelength | 0.71073 Å |
| :---: | :---: |
| Crystal system, space group | Triclinic, P-1 |
| Unit cell dimensions | $\begin{aligned} & \mathrm{a}=6.225(2) \AA \quad \text { alpha }=93.169(5)^{\circ} . \\ & \mathrm{b}=7.268(3) \AA \quad \text { beta }=97.104(5)^{\circ} . \\ & \mathrm{c}=16.151(6) \AA \quad \text { gamma }=109.801(5)^{\circ} . \end{aligned}$ |
| Volume | 678.5(4) $\AA^{3}$ |
| Z, Calculated density | $2,1.666 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.283 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.64 \times 0.55 \times 0.11 \mathrm{~mm}$ |
| Theta range for data collection | 2.56 to 25.00 deg . |
| Reflections collected / unique | $6506 / 2392[\mathrm{R}(\mathrm{int})=0.0236]$ |
| Completeness to theta $=25.00$ | 99.6 \% |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.067 |
| Final R indices [ $1>2$ sigma(I)] | $\mathrm{R} 1=0.0344, \mathrm{wR} 2=0.0960$ |
| R indices (all data) | $\mathrm{R} 1=0.0356, \mathrm{wR} 2=0.0970$ |
| Largest diff. peak and hole | 0.263 and -0.332 e. $\AA^{-3}$ |






ORTEP of 441
Crystal Data Table for 441

| Wavelength | $0.71073 \AA$ |
| :--- | :---: |
| Crystal system, space group | Triclinic, P-1 |
| Unit cell dimensions | $\mathrm{a}=5.9115(8) \AA$ alpha $=101.408(2)^{\circ}$. <br> $\mathrm{b}=10.0414(14) \AA$ beta $=92.553(2)^{\circ}$. <br> $\mathrm{c}=12.5917(17) \AA$ gamma $=103.111(2)^{\circ}$ |
| Volume | $710.44(17) \AA^{3}$ |
| Z, Calculated density | $2,1.666 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.280 \mathrm{~mm} \mathrm{~m}^{-1}$ |
| Crystal size | $1.06 \times 0.28 \times 0.17 \mathrm{~mm}$ |
| Theta range for data collection | 2.13 to 25.00 deg. |
| Reflections collected / unique | $6824 / 2465[\mathrm{R}(\mathrm{int})=0.0209]$ |
| Completeness to theta $=25.00$ | $99.4 \%$ |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.069 |
| Final R indices [I>2sigma(I)] | $\mathrm{R} 1=0.0322, \mathrm{wR2}=0.0867$ |
| R indices (all data) | $\mathrm{R} 1=0.0345, \mathrm{wR} 2=0.0886$ |
| Largest diff. peak and hole | $0.205 \mathrm{and}-0.349 \mathrm{e} . \AA^{-3}$ |

DSC of $\mathbf{4 4 1}$


|  | $\begin{aligned} & \text { Maximum } \\ & { }^{\circ} \mathrm{C} \\ & 234.31 \end{aligned}$ | $\begin{gathered} \text { Stop } \\ { }^{\circ} \mathrm{C} \\ 242.11 \end{gathered}$ | Area J/g 93.86 |
| :---: | :---: | :---: | :---: |
| Glass Transition  <br> Onset Midpoint (I) <br> ${ }^{\circ} \mathrm{C}$ ${ }^{\circ} \mathrm{C}$ <br> 204.51 204.54 | $\begin{gathered} \text { End } \\ { }^{\circ} \mathrm{C} \\ 204.54 \end{gathered}$ | $\begin{aligned} & \text { Height } \\ & \text { mW } \\ & 0.03447 \end{aligned}$ | $\begin{gathered} \text { Delta Cp } \\ \mathrm{J} /\left(\mathrm{g} \cdot{ }^{\circ} \mathrm{C}\right) \\ 0.1880 \end{gathered}$ |

Enlarged region $\left(170-230^{\circ} \mathrm{C}\right)$ in DSC of 441


| Peak Integration |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Start | Onset | Maximum | Stop | Area |
| 225.56 | 232.42 | 234.31 | 242.11 | 93.86 |
| Glass Transition |  |  |  |  |
| Onset | Midpoint (I) | End | Height | Delta Cp |
| ${ }^{\circ} \mathrm{C}$ | ${ }^{\circ} \mathrm{C}$ | ${ }^{\circ} \mathrm{C}$ | mW | $\mathrm{J} /\left(\mathrm{g} \cdot{ }^{\circ} \mathrm{C}\right)$ |
| 194.96 | 202.25 | 201.50 | 0.04154 | 0.2266 |




ORTEP of 442
Crystal Data Table for 442

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Orthorhombic, Pna $2_{1}$ |
| Unit cell dimensions | $\mathrm{a}=11.999(2) \AA$ alpha $=90^{\circ}$. |
|  | $\mathrm{b}=8.3099(15) \AA \quad$ beta $=90^{\circ}$. <br> $\mathrm{c}=8.8995(16) \AA$ gamma $=90^{\circ}$. |
| Volume | $887.4(3) \AA^{3}$ |
| Z, Calculated density | $4,1.888 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.396 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.17 \times 0.15 \times 0.09 \mathrm{~mm}$ |
| Theta range for data collection | 2.98 to 25.00 deg. |
| Reflections collected / unique | $5988 / 1566[\mathrm{R}($ int $)=0.0242]$ |
| Completeness to theta $=25.00$ | $100.0 \%$ |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.131 |
| Final R indices [I>2sigma(I)] | $\mathrm{R} 1=0.0310, \mathrm{wR} 2=0.0727$ |
| R indices (all data) | $\mathrm{R} 1=0.0331, \mathrm{wR} 2=0.0737$ |
| Largest diff. peak and hole | 0.150 and $-0.265 \mathrm{e} . \AA^{-3}$ |
|  |  |







ORTEP of 444
Crystal Data Table for $\mathbf{4 4 4}$

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Triclinic, P-1 |
| Unit cell dimensions | $\mathrm{a}=6.150(3) \AA$ alpha $=97.946(11)^{\circ}$. |
|  | $\mathrm{b}=12.327(6) \AA \quad$ beta $=100.537(9)^{\circ}$. |
| $\mathrm{c}=12.441(6) \AA$ gamma $=96.257(9)^{\circ}$. |  |
| Volume | $909.6(8) \AA^{3}$ |
| Z, Calculated density | $2,1.520 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.227 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.45 \times 0.33 \times 0.31 \mathrm{~mm}$ |
| Theta range for data collection | 2.18 to 25.00 deg. |
| Reflections collected / unique | $4626 / 3166[\mathrm{R}($ int $)=0.0265]$ |
| Completeness to theta $=25.00$ | $98.6 \%$ |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.318 |
| Final R indices $[\mathrm{I} 2$ sigma(I) $]$ | $\mathrm{R} 1=0.0741, \mathrm{wR} 2=0.1756$ |
| R indices (all data) | $\mathrm{R} 1=0.1123, \mathrm{wR} 2=0.1875$ |
| Largest diff. peak and hole | 0.182 and $-0.216 \mathrm{e} . \AA^{-3}$ |






ORTEP of 445
Crystal Data Table for $\mathbf{4 4 5}$

| Wavelength | $0.71073 \AA$ |
| :--- | :--- |
| Crystal system, space group | Monoclinic, P2, $\backslash \mathrm{N}$ |
| Unit cell dimensions | $\mathrm{a}=8.549(2) \AA$ alpha $=90^{\circ}$. <br> $\mathrm{b}=11.112(3) \AA$ beta $=99.109(7)^{\circ}$. <br> $\mathrm{c}=14.775(4) \AA \quad$ gamma $=90^{\circ}$. |
| Volume | $1386.0(7) \AA^{3}$ |
| Z, Calculated density | $8,1.529 \mathrm{Mg}^{-1} \mathrm{~m}^{3}$ |
| Absorption coefficient | $0.227 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.28 \times 0.26 \times 0.21 \mathrm{~mm}$ |
| Theta range for data collection | 2.30 to 25.00 deg. |
| Reflections collected / unique | $6625 / 2432[\mathrm{R}($ int $)=0.0254]$ |
| Completeness to theta $=25.00$ | $99.8 \%$ |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.044 |
| Final R indices [I>2sigma(I) $]$ | $\mathrm{R} 1=0.0342, \mathrm{wR} 2=0.0888$ |
| R indices (all data) | $\mathrm{R} 1=0.0404, \mathrm{wR} 2=0.0933$ |
| Largest diff. peak and hole | 0.244 and $-0.279 \mathrm{e} . \AA^{-3}$ |









ORTEP of 447

Crystal Data Table for 447

| Wavelength | 0.71073 A |
| :---: | :---: |
| Crystal system, space group | Monoclinic, C2/c |
| Unit cell dimensions | $\begin{aligned} & \mathrm{a}=22.887(12) \AA \quad \text { alpha }=90^{\circ} . \\ & \mathrm{b}=6.109(3) \AA \quad \text { beta }=99.784(8)^{\circ} . \\ & \mathrm{c}=26.752(14) \AA \quad \text { gamma }=90^{\circ} . \end{aligned}$ |
| Volume | 3686(3) $\AA^{3}$ |
| Z, Calculated density | $8,1.508 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.224 \mathrm{~mm}^{-1}$ |
| Crystal size | $0.99 \times 0.105 \times 0.061 \mathrm{~mm}$ |
| Theta range for data collection | 2.17 to 25.00 deg. |
| Reflections collected / unique | 8574 / 3235 [R(int) $=0.0456]$ |
| Completeness to theta $=25.00$ | 99.7 \% |
| Refinement method | Full-matrix least-squares on $\mathrm{F}^{2}$ |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.045 |
| Final R indices [I>2sigma(I)] | $\mathrm{R} 1=0.0646, \mathrm{wR} 2=0.1629$ |
| R indices (all data) | $\mathrm{R} 1=0.0917, \mathrm{wR} 2=0.1774$ |
| Largest diff. peak and hole | 0.653 and -0.449 e. $\AA^{-3}$ |

## References:

1 (a) Thomas, J. R.; Dwek, R. A.; Rademacher, T. W. Biochemistry, 1990, 29, 5413. (b) Potter, B. V. L.; Lampe, D. Angew. Chem. Int. Ed. Engl. 1995, 34, 1933. (c) Hinchliffe, K.; Irvine, R. Nature, 1997, 390, 123.

2 (a) The Inositol Phosphates. Chemical Synthesis and Biological Significance, Billington, D. C. VCH, New York, N.Y. 1993. (b) Phosphoinositides: Chemistry, Biochemistry and Biomedical Applications, Bruzik, K. S. Ed.; ACS Symposium Series 718. American Chemical Society, Washington D. C. USA, 1999.

3 Suzuki, T.; Suzuki, S. T.; Yamada, I.; Koashi, Y.; Yamada, K. and Chida, N. J. Org. Chem., 2002, 67, 2874. (b) Chida, N.; Ogawa, S. Chem. Commun., 1997, 807.

4 Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Das, T. Chem. Rev. 2003, 103, 4477.

5 Angyal, S. J. Carbohydr. Res., 2000, 325, 313 and the references cited therein.

## List of publications:

1. Chelation controlled regiospecific O-substitution of myo-inositol orthoesters: Convenient access to orthogonally protected myo-inositol derivatives.
S. Devaraj, M. S. Shashidhar and S. S. Dixit, Tetrahedron 2005, 61, 529.
2. $\mathrm{OH} . . . \mathrm{O}$ bridge dimmers link via $\mathrm{CH} . . . \mathrm{O}$ and $\mathrm{CH} \ldots . \pi$ interaction in the crystal structure of 4,6-di-O-benzyl myo-inositol 1,3,5-orthoformate.
K. Manoj, S. Devaraj, Rajesh Gonnade, M. M. Bhadbhade, M. S. Shashidhar, Acta Crystallographica C. 2005, 61, 628.
3. Cyclitol based metal complexing agents. Preference for the extraction of lithium by myo-inositol based crown-4-ethers depends on the relative orientation of crown ether oxygen atoms.
S. S. Dixit, M. S. Shashidhar and S. Devaraj, Tetrahedron 2006, 62, 4360.
4. Chelation controlled regioselective O-substitution in cyclohexanepolyols.
S. Devaraj and M. S. Shashidhar (Manuscript under preparation).
5. Intramolecular acyl migration in trioxaadamantane derivatives (myo-inositol 1,3,5-orthoesters): transesterification reaction without the involvement of the tetrahedral intermediate.
K. M. Sureshan, S. Devaraj and M. S. Shashidhar (Manuscript under preparation).

## Posters presented in conferences:

1. Effect of metal ion on regioselective alkylation and acylation of myo-inositol orthoesters.
S. Devaraj and M. S. Shashidhar.

Poster presented at the Fourth National Symposium in Chemistry, Feb. 1-3, 2002
(Organized by Chemical Research Society of India); National Chemical
Laboratory; Pune-411 008; Maharashtra, India.

## Oral presentations in conferences:

1. Effect of metal ion on the regioselective nucleophilic $O$-substitution in myoinositol orthoesters.
S. Devaraj and M. S. Shashidhar.

Presented at the first RSC-West India section students symposium, 2003, National Chemical Laboratory, Pune, India.

